Confirmation and Quantification of Genotoxic Impurity 2-Dimethylaminoethyl chloride hydrochloride (DMC HCl) by GCMS in Chlorpheniramine/Chlorphenamine Maleate

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Abstract: The purpose of this research work is to develop a suitable GCMS method for the quantitative determination of Genotoxic impurity 2-Dimethylaminoethyl chloride hydrochloride (DMC HCl). The compound is inorganic and it is not easy to quantify at ppm level present in Chlorpheniramine/ Chlorphenamine Maleate Active Pharmaceutical Ingredients (APIs) with any other methods. Hence the GCMS method was developed on Thermo Scientific, WAXMS, 30 m x 0.32 mm x 0.25µmcolumn using Thermo Scientific Trace 1310 and TSQ at a flow rate of 1.0 mL/min. Under these conditions impurity was quantified by selecting mass range 40 – 700 amu. The limit of detection and the limit of quantitation for the impurity were established. Validation of the developed GCMS method was carried out as per ICH requirements and the data shows that, the proposed method is specific, linear, accurate, precise and robust. This method has been tested in a number of Chlorpheniramine/ Chlorpheniramine/ Chlorpheniramine dust successfully for quantification of the impurity at ppm level. The developed GCMS method was found to be suitable to quantify the Genotoxic impurity 2-Dimethylaminoethyl chloride hydrochloride (DMC HCl) at ppm level present Chlorpheniramine/ Chlorphenamine Maleate. **Keywords:** (GCMS), Genotoxic impurity; 2-Dimethylaminoethyl chloride hydrochloride (DMC HCl); Thermo

Scientific, Chlorpheniramine/ Chlorphenamine Maleate; Threshold of Toxicological Concern (TTC).

I. Introduction

2-Dimethylaminoethyl chloride hydrochloride (DMC HCl)[1]is Process Impurity of Chlorpheniramine/ Chlorphenamine Maleate (Fig.1). 2-Dimethylaminoethyl chloride hydrochloride (DMC HCl) shows presence of structural alert for genotoxic mutagenicity and carcinogenicity. The QSTR models predict the compound positive for genotoxicity, mutagenicity and carcinogenicity the compound is shown the positive for mutagenicity in training set used for Ames mutagenicity model [2-5].In genetics, Genotoxicity describes as property of chemical compound which may damage the genetic information within a cell leading mutations, which can lead to different types of Cancers in Human body in any forms.

While genotoxicity is often confused with mutagenicity, all mutagens are genotoxic; however, not all genotoxic substances are mutagenic. The alteration can have direct or indirect effects on the DNA: the induction of mutations, mistimed event activation and direct DNA damage leading to mutations. The permanent, heritable changes can affect either somatic cells of the organism or germ cells to be passed on to coming/future generations. Cells prevent expression of the genotoxic mutation by either DNA repair or apoptosis; however, the damage may not always be fixed leading to mutagenesis[4,5] Specifically, there is evidence that genotoxic substances may bind directly to DNA and may also act indirectly by affecting enzymes involved in DNA replication. There are three primary effects that Genotoxins can have on organisms by affecting their genetic information. Genotoxins can be carcinogens, or cancer-causing agents, mutagens, or mutation-causing agents, or teratogens, birth defect-causing agents [6]. The toxicological assessment of these genotoxic impurities and the determination of acceptable limits for such impurities in active substances is a difficult issue and not addressed in sufficient detail in the existing International Conference on Harmonization (ICH) Q3X guidelines [7]. The presence of trace level of the Genotoxic Impurity in drug substance or drug product is of genotoxicity concern and has been closely monitored by regulatory agencies and pharmaceutical industries [8]. The 'threshold of toxicological concern' (TTC) of 1.5 µg/person/day (exposure of genotoxic impurity in drugs that will be tested or dosed for longer than 12 months) has been suggested by the European Medicines Agency's (EMEA) "Guideline on the limits of genotoxic impurities" [9] and the Pharmaceutical Research and Manufacturers of America's (PhRMA) white paper [13]. Based on the TTC, the concentration limits of genotoxic impurity in drug substances or drug products can then be derived based on the maximum daily dose: concentration limit (ppm) =

[1.5 µg /day] / [dose (g/day)][2]. For a drug dosed at 1g per day, for example, 1.5 ppm would be the limit of a specific genotoxic impurity which would also be the 'target analyte level' (TAL) from an analytical perspective [7-9]. Given such a low ppm concentration limit, besides the control challenges in process chemistry, developing sensitive and robust methodology for their detection poses a tremendous analytical challenge for the pharmaceutical industry[10-15]. Therefore, potential genotoxins must be minimized during the synthesis the compounds and where there is difficulty achieving this, the method of manufactureshould preferably be changed 'As 2-Dimethylaminoethyl chloride hydrochloride (DMC HCl) is a genotoxic compound, the regulators may require the toxin levels to be controlled to 37.5 ppm in the drug substance on the basis of Maximum Daily Dose of drug substance. The Quantification at such low level can be possible only by using GCMS or LCMS/MS and also there is no method for the quantification of this impurity hence a high sensitive GCMS method developed for the quantification of this genotoxic impurity 2-Dimethylaminoethyl chloride hydrochloride (DMC HCl). The present trend in pharmaceutical analysis shows tremendous advancement in physio-chemical method of analysis for drugs which are very precise and accurate, the modern methods of choice involves sophisticated instruments like High performance liquid chromatography(HPLC),gas chromatography(GC), Mass spectrometry, Nuclear magnetic resonance spectroscopy(NMR)[16-17].

II. Experimental

Chemicals and reagents: The samples of Chlorpheniramine/Chlorphenamine maleate and 2-Dimethylaminoethyl chloride hydrochloride (DMC HCl) (Fig. 2) were collected from Supriya Lifescience Ltd., Mumbai, Maharashtra, India. GCMS grade Dimethyl Sulfoxide was purchased from Advent, Mumbai, India.

Equipment: The GCMS method development and validation were done using Thermo Scientific Trace 1310 and TSQ 8000 Connected with Thermo Scientific mass detector GCMS system. The data were collected usingChromeleonTM6.8 Chromatography Data System (CDS) Software.

GCMS chromatographic conditions: The GC chromatographic separations were achieved on Thermo Scientific TRACETM 1310 Gas Chromatograph with TSQTM 8000 Evo Triple Quadrupole GC-MS/MS. The GC was equipped with a capillary column (Thermo Scientific, TG-WAXMS, 30 m x 0.32 mm x 0.25µm) and run in full-scan mode (scan range 40-700 m/z with Detector voltage 2160V). Helium was employed as the carrier gas. The injector temperature was 250 °C and the initial oven temperature was 50 °C, which was held for 5 minutes. The oven was ramped at 10 °C/min to 230°C. The final temperature was held for 7 minutes for a total run time of 30.00 minutes. Three mass spectral libraries were used: an in-house library created using neat reference samples materials (OCME), the 2008 Scientific Working Group for the Analysis of Seized Drugs (SWGDRUG) library, and a combined Wiley and National Institute of Standards and Technology (NIST) library. ChromeleonTM6.8 Chromatography Data System (CDS) Software was used to quantitatively analyse the samples and qualifier ions were used to determine the presence and concentration of the analytes of interest.

Preparation of genotoxic impurity standard and test sample Solution: The stock solution of impurity standard prepared at approximately0.0375 mg/ml (37.5 ppm) in dimethyl Sulfoxide. For linearity, the stock solution impurity was diluted using diluents to give standards at 0.23, 0.47, 0.94, 1.87, 3.75 ppm with respect to test concentration. The testing API samples were typically prepared at approximately 100 mg/mL in dimethyl Sulfoxide.

III. Result And Discussion

Linearity: The linearity of 2-Dimethylaminoethyl chloride hydrochloride (DMC HCl) was satisfactorily done. A series of solutions were prepared using 2-Dimethylaminoethyl chloride hydrochloride (DMC HCl) at concentration levels from around quantification level to 150% and the concentration levels are 18.75, 30.00, 37.50, 45.00, 56.25 ppm respectively. The peak area versus concentration data was done by linearity plot slope, intercept, and residual sum of squares analysis. The calibration curve was given based on response over the concentration range for 2-Dimethylaminoethyl chloride hydrochloride (DMC HCl). The correlation coefficient2-Dimethylaminoethyl chloride hydrochloride (DMC HCl) was 0.996 and the Linearity results are tabulated in table 1 and Fig.3.

Limit of Detection (LOD) and Limit of Quantitation (LOQ):The LOD and LOQ values of 2-Dimethylaminoethyl chloride hydrochloride (DMC HCl) were predicted from the predication linearity data. Each predicted concentration was verified for precision by preparing the solutions at about predicted concentration and injecting each solution six times for GCMS/MS study and the predicted concentration for LOQ was 3.7 ppm and LOD was 0.94ppm (Fig.4 ABC & D) and the results are tabulated in table 2.

Precision: The precision of the developed method was checked by preparing solutions by spiking the impurity at 100% level with the drug substance for six times and injected each once also injected100% spiked solution for 6 times to show the system precision. The %relative standard deviation (RSD) of the areas at each level 5.3% and 13.21 confirming developed that method is précised.

Accuracy: The accuracy of the method was evaluated in sample solutions were prepared in triplicate by spiking 2-Dimethylaminoethyl chloride hydrochloride (DMC HCl) at LOQ level, 50%, 100% and 150% with Chlorpheniramine/Chlorphenamine Maleate and injected each solution in to GCMS as per methodology. The percentage of recovery for the impurity was calculated and the values are 92.9%, 93.1%, 98.8% and 101.4%. At such low levels, these recoveries and % relative standard deviation (RSD) were satisfactory and the results are tabulated in table 3.



IV. Tables And Figures

Fig. 1: Reaction Scheme of Chlorpheniramine Maleate.



Fig. 2: Structures of Chlorpheniramine Maleate and DMC HCl (its genotoxic impurity).

Level	Conc. (ppm)	Mean Area		
Level				
1	3.713	936638		
2	18.565	4365170		
3	29.704	8567094		
4	37.13	10829855		
5	44.556	13351036		
6	55.695	15695812		
	Slope	297028		
	Correlation	0.996		
	Intercept	-416765.5		
	Intercept (%)	-3.64		
	Residual sum of squares	0.992572		

Table 1: The regression analysis data for DMC HCl





Table 2: Table for LOQ Precision					
Injection	jection Area of DMC HCL (3.7 ppm)				
1	728611				
2	731306 771110 970365 953198				
3					
4					
5					
6	886350				
Mean	840156.66				
SD	110385.33				
%RSD	13.1				







Table 3: % recoveries found for spiked DMC HCl in Chlorpheniramine Maleate

Level	Qty. Added (ppm)	Mean Qty. Added (ppm)	Area	Qty. Recovered (ppm)	Mean Qty. Recovered (ppm)	% Recovery	Mean %Recovery
LOQ-1	3.74		1129101	3.45		92.26	
LOQ-2	3.74	3.74	1136523	3.51	3.52	93.85	94.03
LOQ-3	3.74		1228675	3.59		95.99	
50%-1	18.68		6426461	17.12		91.65	
50%-2	18.68	18.68	7067019	18.83	17.39	100.80	94.01
50%-3	18.68		6292017	16.92		90.58	
100%-1	37.35		14044889	37.42		100.15	
100%-2	37.35	37.35	13709442	36.52	36.91	97.78	98.80
100%-3	37.35		13807542	36.78		98.47	
150%-1	56.03		20201040	53.82		96.06	
150%-2	56.03	56.03	20435043	54.44	54.80	97.16	97.81
150%-3	56.03		21327091	56.15		100.21	
						Mean	96.16
						SD	2.51
						% RSD	2.61

SD= Standard Deviation, RSD= Relative Standard Deviation

V. Conclusion

On the basis of above study, reported method is sensitive specific, accurate, validated and well-defined GCMS/MS method for the Quantification of genotoxic impurity 2-Dimethylaminoethyl chloride hydrochloride (DMC HCl) at ppm level in Chlorpheniramine/Chlorphenamine maleate. The detection limit and quantification limit found to be 0.94 ppm and 3.75 ppm respectively. The described method is highly reliable technique for the quantification of the genotoxic impurity present in the Chlorpheniramine/Chlorphenamine maleateduring quality control testing.

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