

## Antibacterial Activity of n-Hexane Extract of *Ocimum gratissimum* leaves

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**Abstract:** The antibacterial activity of n-hexane extract of *Ocimum gratissimum* leaves was investigated against gram negative bacteria (*Salmonella typhi*) and gram positive bacteria (*Staphylococcus aureus*). Agar well diffusion and broth dilution methods were employed. The results revealed that both first and second n-hexane extracts (4days and 8 days n-hexane extract respectively) inhibited the microorganisms with the second being more active as it inhibited at 30mg/ml, 40mg/ml, 50mg/ml and 100mg/ml with zone of inhibition ranging from 8.5mm to 15.5mm in diameter. The first extract showed no inhibition at 30mg/ml and 40mg/ml respectively but showed inhibition at 50mg/ml and 100mg/ml with zone of inhibition ranging from 9mm to 12mm in diameter. The Minimum Inhibitory Concentration (MIC) of the second n-hexane extract ranged from 25mg/ml to 100mg/ml whereas, that for the first was at 100mg/ml. The result was compared to conventional drug (Gentamicin 20ug/ml) with zone of inhibition at 20mm in diameter. The Gas Chromatography Mass Spectrometry (GC/MS) identified antibacterial compounds which include Hexadecanoic acid, Phytol, Ricinoleic acid, Dehydrodihydrodiisoeugenol and 9-Octadecenamide. Based on the results it can be concluded that *Ocimum gratissimum* leaves extract possess antibacterial activity and can be applied in development of potent antibacterial drugs.

**Keywords:** *Ocimum gratissimum*, n-hexane extract, antibacterial activity, Gas Chromatography Mass Spectrometry (GC/MS).

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### I. Introduction

Diseases are growing more resistant to synthetic drugs leading to researchers finding better alternatives from plant resulting to the comeback of 'green medicine'. *Salmonella typhi* a gram negative, rod shaped motile and anaerobic bacteria from genus *Salmonella* and family Enterobacteriaceae [1], the causal organism of typhoid fever. There has been resistance of this disease to ampicillin, chloramphenicol, trimethoprim-sulfamethoxazole and streptomycin [2]. *Staphylococcus aureus* gram positive cocci and anaerobic bacteria, causal organism of skin infections, pneumonia and food poisoning.

*Ocimum gratissimum* from the family Lamiaceae and genus *Ocimum* is a perennial herb found in tropical and temperate region. It is the species commonly found in Nigeria. It is well known for its nutritional and medical properties. It has long been used in folkloric medicine to treat fever, skin infections and respiratory tract infections [3]. It is antifungal [4], [5], antibacterial [6], [7], antiviral [8], antileishmanial [9], antitrypanosomal [10], antidiabetic [11], anticonvulsant [12], antioxidant [13]. Phytochemical analysis [14] confirmed the presence of alkaloids, tannins, phenolics, saponins, glycoside, and resins. Steroids and terpenoids were also confirmed to be present in the plant [15]. GC/MS of its essential oil (ocimum oil) [16] showed eugenol to be major component, thymol, germacrene, bisabolene and other volatiles as minor component. This present study is aimed at determining the antibacterial activity of *Ocimum gratissimum* leaves.

### II. Materials And Method

#### 2.1 COLLECTION AND EXTRACTION OF PLANT

*Ocimum gratissimum* leaves were collected from a vegetable garden along Zaria road in Jos North area of Plateau State in April 2013. It was identified by Mr Agyeno of Botany Department University of Jos, Plateau state Nigeria. The leaves were rinsed with water and shade dried at room temperature then pulverized. 50g of the powder was extracted for 4 days with 250ml research grade n-hexane with occasional stirring (twice a day). The marc was separated from the solvent by decantation and labeled first n-hexane extract of *Ocimum gratissimum* leaves (OGH<sub>1</sub>E). Another 250ml of n-hexane was added to the marc and left for another 4 days with occasional stirring then decanted and labeled second n-hexane extract of *Ocimum gratissimum* leaves (OGH<sub>2</sub>E). Both extracts (first and second n-hexane) were evaporated to constant weight using a steam bath.

#### 2.2 ANTIBACTERIAL ASSAY

The microorganisms which were *Salmonella typhi* and *Staphylococcus aureus* were sourced from the Department of Pharmaceutical Science and Technology University of Jos. They were sub cultured in 5ml sterile nutrient broth then incubated at 37<sup>o</sup>C for 24hrs. 0.1ml of the 24hr broth culture was aseptically introduced into a

sterile nutrient agar, 20ml each was then transferred aseptically into petri dishes .Using a 3mm cork borer, holes were bored in each agar plate .The wells were filled with 30mg/ml, 40mg/ml, 50mg/ml and 100mg/ml of the extract dissolved in n-hexane. The plates were then incubated at 37<sup>0</sup>C for 24 hrs. The zone of inhibition of each well was obtained in millimeters. Gentamicin served as the positive control while n-hexane was used as the negative control.

**2.3 DETERMINATION OF MINIMUM INHIBITORY CONCENTRATION (MIC)**

This was done by the broth dilution method and turbidity of each tube was observed. The controls contained no microorganism only the extract .The lowest concentration showing no turbidity was the MIC.

**2.4 DETERMINATION OF MINIMUM BACTERICIDAL CONCENTRATION (MBC)**

The MBC was further carried out using the tubes that showed no turbidity and then smearing a loopful on a sterile nutrient agar plate, then incubating at 37<sup>0</sup>C for 24hrs and growth of microorganisms were observed.

**2.5 CHEMICAL ANALYSIS OF EXTRACT**

The n-hexane extract was chemically analyzed using a QP2010 PLUS SHIMADZU, JAPAN Gas Chromatograph interfaced to a Mass Spectrometer (GC/MS) instrument under the following conditions : AOC-20i auto injection , fused silica capillary column (RTX-5MS : 30m x 0.25mm I.D . d .f = 0.25µm)which was programmed as follows : flow rate of carrier gas(He) was 1ml/min , injection port temperature 200<sup>0</sup>C, initial temperature of column being 50<sup>0</sup>C increased and kept constant at 300<sup>0</sup>C for 9mins with heating rate of 8<sup>0</sup>C/min.The volume injected was 1µl with split mode. Identification of the extract constituents was made based on retention times and by comparison of mass spectra with the computer search using NIST05 library of mass spectral data.

**III. Result**

The percentage yields of first and second n-hexane extract were 3% and 2% respectively. Antibacterial activity showed that n-hexane extract had zones of inhibition between 9 – 12 mm for the first extract (Table 1) and 8.5 – 15.5mm for the second extract (Table 4). Minimum Inhibitory Concentration (MIC) and Minimum Bactericidal Concentration (MBC) for the first and second extracts are as shown in Tables 2, 3, 5 and 6. The chromatograms for OGH<sub>1</sub>E and OGH<sub>2</sub>E are shown in figures 1 and 2a respectively. The mass spectra of compounds responsible for antibacterial activity are presented in figures 2a<sub>1</sub> to 2a<sub>5</sub>.

**Table 1: Antibacterial activity of first n-hexane extract of *Ocimum gratissimum* leaves (OGH<sub>1</sub>E).**

Microorganisms	Zones of inhibition of OGH <sub>1</sub> E (mm) / Concentration (mg/ml)				Gentamicin (20ug/ml)	n-Hexane
	30	40	50	100		
<i>Salmonella typhi</i>	-	-	11	12	20	-
<i>Staphylococcus aureus</i>	-	-	9	12	20	-

- = No inhibition

**Table 2: Minimum Inhibitory Concentration (MIC) of the first n-hexane extract of *Ocimum gratissimum* leaves.**

Microorganisms	MIC / Concentration (mg/ml)				
	6.125	12.5	25	50	100
<i>Salmonella typhi</i>	+	+	+	+	-
<i>Staphylococcus aureus</i>	+	+	+	+	+

+ = Turbidity  
- = No turbidity

**Table 3: Minimum Bactericidal Concentration (MBC) of the first n-hexane extract of *Ocimum gratissimum* leaves.**

Microorganisms	MBC / Concentration (mg/ml)				
	6.125	12.5	25	50	100
<i>Salmonella typhi</i>	N.T	N.T	N.T	N.T	+
<i>Staphylococcus aureus</i>	N.T	+	+	+	+

+ = Growth  
N.T = Not tested

**Table 4: Antibacterial activity of second n-hexane extract of *Ocimum gratissimum* leaves (OGH<sub>2</sub>E).**

Microorganisms	Zones of inhibition of OGH <sub>2</sub> E (mm) / Concentration (mg/ml)				Gentamicin (20ug/ml)	n-Hexane
	30	40	50	100		
<i>Salmonella typhi</i>	9	10.5	12	15.5	20	-
<i>Staphylococcus aureus</i>	8.5	10	11	12	20	-

- = No inhibition

**Table 5: Minimum Inhibitory Concentration (MIC) of second n-hexane extract of *Ocimum gratissimum* leaves.**

Microorganisms	MIC / Concentration (mg/ml)				
	6.125	12.5	25	50	100
<i>Salmonella typhi</i>	+	+	+	-	-
<i>Staphylococcus aureus</i>	+	+	-	-	-

+ = Turbidity

- = No turbidity

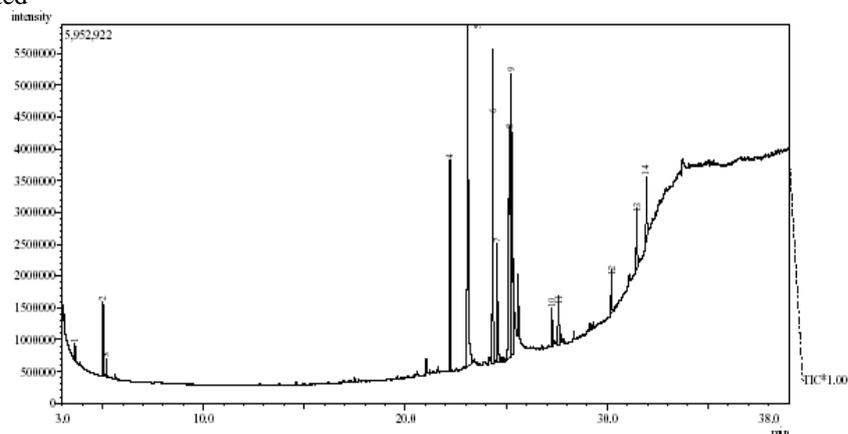
**Table 6: Minimum Bactericidal Concentration (MBC) of second n-hexane extract of *Ocimum gratissimum* leaves.**

Microorganisms	MBC / Concentration (mg/ml)				
	6.125	12.5	25	50	100
<i>Salmonella typhi</i>	N.T	N.T	N.T	-	-
<i>Staphylococcus aureus</i>	N.T	N.T	+	-	-

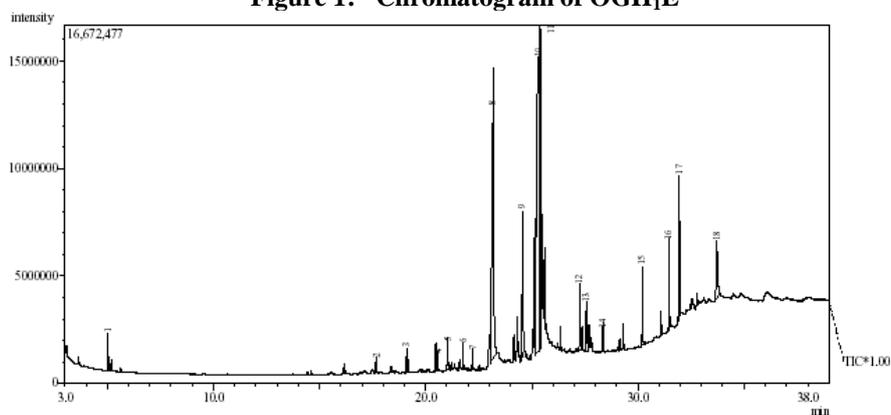
- = No growth

+ = Growth

N.T = Not tested

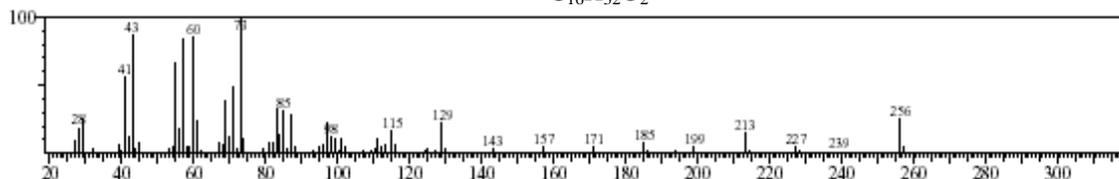
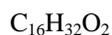


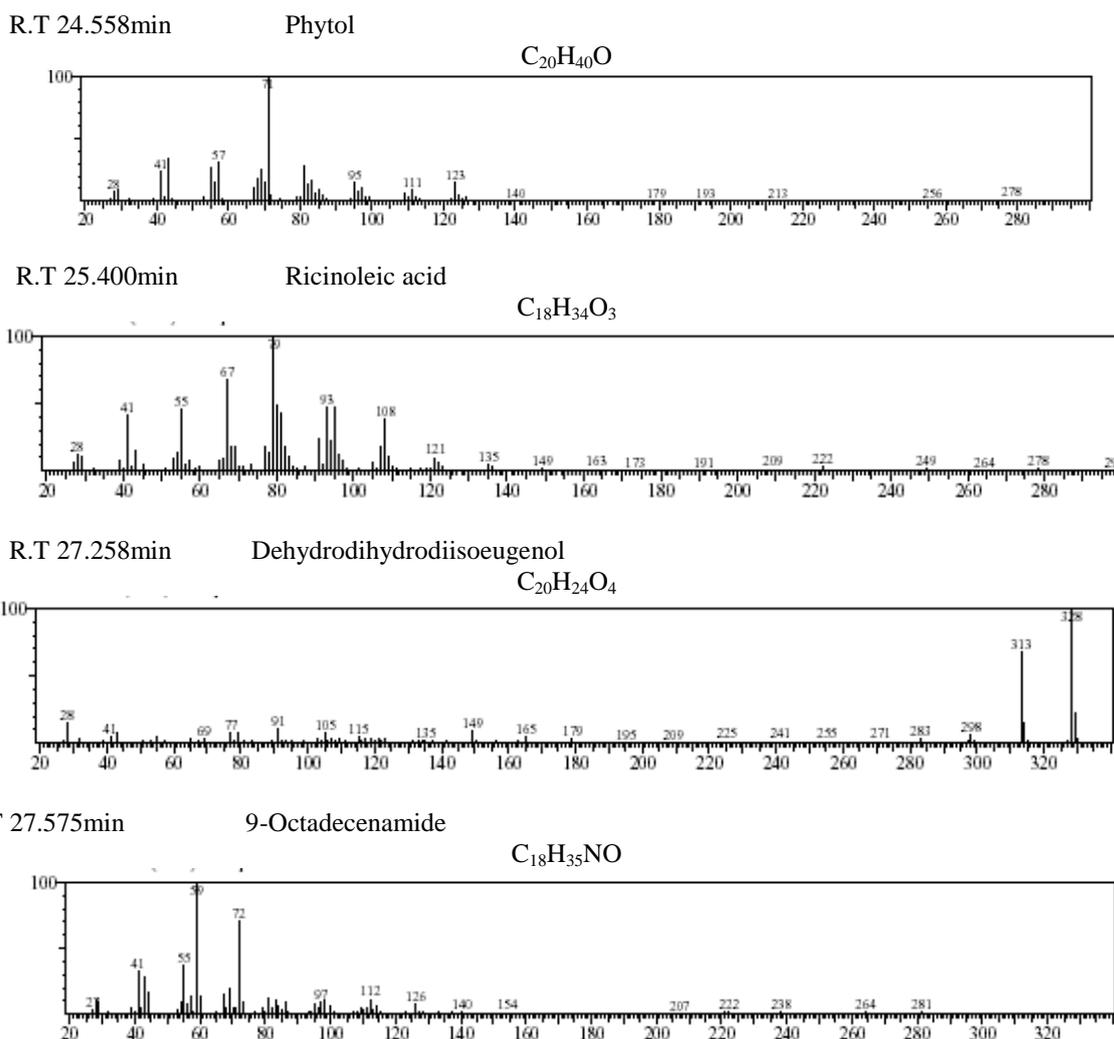
**Figure 1: Chromatogram of OGH<sub>1</sub>E**



**Figure 2a: Chromatogram of OGH<sub>2</sub>E**

R.T 23.108min Hexadecanoic acid





Figures 2a<sub>1</sub> to 2a<sub>5</sub>: Mass spectra of compounds responsible for antibacterial activity according to their retention times (R.T).

#### IV. Discussion

The present study showed that the n-hexane extract of *Ocimum gratissimum* leaves inhibited the growth of both microorganisms. It can also be seen that the extract was both bacterostatic and bactericidal. From chromatogram of OGH<sub>1</sub>E, Peak 5 which is the highest is hexadecanoic acid with R.T 23.108min. Peak 7 is phytol with R.T 24.550min. Dehydrodihydrodiisoeugenol is peak 10 with R.T 27.267min. 9-Octadecenamide is peak 11 with R.T 27.600min. From chromatogram of OGH<sub>2</sub>E, peak 8 is hexadecanoic acid also in high proportion with R.T 23.175min. Peak 9 is phytol higher than that in OGH<sub>1</sub>E with R.T 24.558min. Peak 11 being the highest peak is ricinoleic acid with R.T 25.400min. Peak 12 is dehydrodihydrodiisoeugenol higher than that in OGH<sub>1</sub>E with R.T 27.258. 9-octadecenamide is peak 13 with R.T 27.575min. From the chromatograms it can be inferred that the retention times of the compounds are similar and increase in dehydrodihydrodiisoeugenol, phytol and especially the presence of ricinoleic acid in OGH<sub>2</sub>E lead to increase in antibacterial activity and inhibition of bacterial strains at low concentrations. Hexadecanoic acid has been shown to be a potent antibacterial compound [17], [18]. Phytol has been shown to possess activity against *Salmonella typhi* and resistant gonorrhoea [19]. Eugenol and isoeugenol has also been observed to possess antibacterial activity [20], so dehydrodihydrodiisoeugenol being a phenolic compound could exhibit certain antibacterial activity. Ricinoleic acid being the major component of castor oil also possesses antibacterial and antifungal properties [21].

## V. Conclusion

The data analysis of this work has shown *Ocimum gratissimum* to be effective against gram positive and gram negative strains of bacteria making it a broad spectrum antibacterial drug and provides the possibility of treating various bacterial infections by applying it in production of pharmaceuticals.

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## References

- [1]. OcheiKolhatkar, Medical laboratory science theory and practice, six edition 2007.
- [2]. Typhoid fever-Wikipedia, the free encyclopedia. <http://en.m.wikipedia.org/wiki/Typhoid-fever>.
- [3]. Adebolu T.T, Oladimeji S.A. Antimicrobial Activity of Leaf Extract of *Ocimum gratissimum* on Selected Diarrhea causing Bacteria in South West Nigeria. African Journal of Biotechnology, 4(7), 2005, 682- 684.
- [4]. Dubey N.K, Tiwari T.N, Mandin D, Andriamboavonjy H, Chaumont J.P. Antifungal Properties of *Ocimum gratissimum* Essential oil (ethyl cinnamatechemotype). Fitoterapia, 71, 2000, 567-569.
- [5]. Terezinha de Jesus Faria, Rafael Sottero Ferreira, LidianeYassumoto, Jose' Robert, Pinto de Souza, Noemia Kazue Ishikawa and Aneli de Melo Barbosa. Antifungal Activity of Essential oil Isolated from *Ocimum gratissimum* L. (eugenolchemotype) against Phytopathogenic fungi. Brazilian Archives of Biology and Technology, 49(6), 2006, 867-871.
- [6]. Junaid S.A, Olabode A.O, Onwuliri F.C, Okwori A.E and Agina S.E. The Antimicrobial Properties of *Ocimum gratissimum* Extract on some Selected Bacterial Gastrointestinal Isolates. African Journal of Biotechnology, 5(22), 2006, 2315-2321.
- [7]. Matasyoh L.G, J.C Matasyoh, F.N Wachira, M.G Kingua, A.W ThairuMuigai, T.K Mukiama. Antimicrobial Activity of Essential oils of *Ocimum gratissimum* L. from Different Populations of Kenya. Afr.J.Trad.CAM, 5(2), 2008, 187-193.
- [8]. Ayissi N.K and Nyadedzor C. Comparative in-vitro effects of AZT and Extracts of *Ocimumgratissimum*, *Ficuspolita*, *Clausernaanisata*, *Alchorneacordifolia*, and *Elaeophorbiadrupifera* against HIV-1 and HIV-2 infections. Antiviral Res, 58, 2003, 25-33.
- [9]. Ueda-Nakamura T, R.R Mendonca- Filho, J.A Moryo do-diaz, P.K Maza and B.P.D Fiho. Antileishmanial Activity of eugenol-rich Essential oil from *Ocimum gratissimum*. Parasitol.Int, 55, 2006, 99-105.
- [10]. Adamu M, Nwonsu C.O, Agbede R.I.S. Antitrypanosomal Effects of Aqueous Extract of *Ocimumgratissimum* (Lamiaceae) Leaf in rats infected with Trypanosome bruceibrucei. Afr.J.Trad CAM, 6(3), 2009, 262-267.
- [11]. Aguiyi J.C, E.L Obi, S.S Gand and A.C Igweh. Hpglycemic Activity of *Ocimum gratissimum* in rats. Fitoterapia, 71, 2000, 444-446.
- [12]. Freire C.M.M, Marques M.O.M and Costa M. Effects of Seasonal Variation on the Central Nervous System Activity of *Ocimum gratissimum* L. Essential oil. J.Ethnopharmacol, 105, 2006, 161-166.
- [13]. Aprioku J.S, Obianime A.W. Antioxidant Activity of Aqueous Crude Extract of *Ocimumgratissimum*Linn.Leaf on Basal and Cadmium-induced Serum Levels of Phosphatases in Male guinea-pigs. J.Appl.Sci.Environ.Manage, 12(4), 2008, 33-39.
- [14]. Koche D.K, Kokate P.S, Suradkar S.S and Bhadange D.G. Preliminary Phytochemical and Antibacterial Activity of Ethanolic Extract of *Ocimum gratissimum* L. Bioscience Discovery , 3(1), 2012, 20-24.
- [15]. Ladipo M.K, Doherty V.F and Kanife U.C. Phytochemical screening and Antibacterial Investigation of the Extract of *Ocimum gratissimum* (scent leaf) on Selected Enterobacteriaceae. PAT, 6(2), 2010, 75-84.
- [16]. Janine de Aquino Lemos, XistoSenaPassos, Orionaida de Fatima LisboaFernande, Jose'Realino de Paula, Pedro Henrique Ferri, Lucia KiokoHasimoto de Souza, Aline de Aquino Lemos, Maria de Rosario, Rodriques Silva. Antifungal Activity from *Ocimum gratissimum* L. toward *Cryptococcus neoformans*. Mem. Inst. Oswaldo Cruz, 100(1), 2005, 55-58.
- [17]. M.C Graw, L.J Jager, A.K Vanstaden J. Isolation of Antibacterial fatty acids from *Schotiabrachypetala*. Fitoterapia, 73, 2002, 431-433.
- [18]. Seidel V, Taylor P.W. In-vitro Activity of Extract and Constituents of *Pelegonium* against rapidly growing Mycobacteria. Int.J.Antimicrob.Agent, 23, 2004, 613-619.
- [19]. Akpuaka A, Ekwenchi M.M, Dashak D.A, Dildar A. Biological Activities of Characterized Isolates of n-Hexane Extract of *Azadirachtaindica*A.Juss (Neem) Leaves. Nature and Science, 11(5), 2013, 141-147.
- [20]. Matasyoh L.G, Josphat C.M, Francis N.W, Miriam G.K, Anne W.T.M, Titus K.M. Chemical Composition and Antimicrobial Activity of Essential oil of *Ocimum gratissimum* L. growing in Eastern Kenya. Afr.J.Biotech, 6, 2007, 760-5.
- [21]. Medical uses of castor oil. <http://sobelarry.hubpages.com/hub/Medical> uses of castor-oil.