Chemical Composition, In Vitro Digestibility And Gas Production Characteristics Of Diets Containing Sweet Orange (*Citrus Sinensis*) Peel Meal

¹J. Oloche., I.O.A. Oluremi and Ayoade, J.A

¹Department of Animal production, University of Agriculture, Makurdi *Department of Animal Nutrition, University of Agriculture, Makurdi

Abstract: The nutritive value of four diets containing different levels of sweet orange peel meal (SOPM), $D_1(0\%SOPM)$, $D_2(12.5\% SOPM)$, $D_3(25\% SOPM)$ and $D_4(50\% SOPM)$ were evaluated in terms of their chemical composition, in vitro gas production (IVGP), metabolizable energy (ME), organic matter digestibility (OMD), short chain fatty acids (SCFA) and in vitro gas production characteristics. Results showed that the in vitro gas production characteristics did not show any significant difference (p>0.05) among treatments. The values for potential gas production 'b' and the potentially degradable fractions 'a+b' for all the dietary treatments were similar (p>0.05), but absolute values of $D_4(50\% SOPM)$ for 'b' and 'a+b' (30.88 and 38.59) respectively, were higher than those of D_1 , D_2 and D_3 . The rate of fermention 'c' was also similar across the treatments. In vitro gas production volumes were significantly affected (p<0.05) among the treatments. Metaboliazable energy for $D_4(6.56MJ/kgDM)$ was significantly higher than $D_1(5.30 MJ/kgDM)$, $D_2(5.32 MJ/kgDM)$. The OMD and SCFA followed the same trend.

Key words: Sweet orange peel meal, in vitro fermentation, in vitro gas production parameters, in vitro gas production characteristics,

I. Introduction

Increasing demand and consequently high cost of conventional animal feedstuffs in the tropics has created the need for sustained alternatives, particularly natural feed resources indigenous to the region. In Nigeria, ruminants slowly gain weight in the rainy season and rapidly loose it in the dry season, yet in the traditional animal husbandary, ruminants in the tropics especially in Nigeria, are mainly fed with grasses, so that improved livestock production is not likely attainable and sustainable by grasses forage alone (Babayemi and Bamikole, 2006). Babayemi et al., (2003) had earlier reported that the forages are unimproved and low in nutritive values during the wet season, while during the dry season proper, they are fibrous, lignified with low protein values and even in short supply. Recently, Lamidi et al., (2010) agreed to this by reporting that, the available forages for most part of the year are low in protein content which leads to marked decrease in voluntary intake and digestibility, and subsequently leads to substantial weight loss of the animals during this period. The success of the livestock industry anywhere in the world depends greatly on feed quantity and quality. Ruminants do better when energy and protein rich diets are strategically combined for feeding (Adegbola and Asaola, 1986). However, the expensive nature of conventional feed as a result of competition between man and livestock (Ogunbosove and Babayemi, 2010), makes this combination difficult. Thus, there is the need to source for locally available feedstuffs which are less expensive and viable. In Nigeria, sweet orange (*Citrus sinensis*) peels is one such agricultural by-product. In vitro gas production technique is a quick, cheap and accurate method of assessing nutritive values of feedstuff (Menke et al., 1979). It involves volumetric measurement of gas production in phosphate and bicarbonate buffered in vitro incubation in syringes. Although, gas produced during rumen fermentation are waste products and of no nutritive value to the ruminants, gas production tests are used routinely in feed research as gas volumes are related to both the extent and rate of substrate degradation (Blummel et al., 1997). This study was therefore designed to assess the chemical composition, in vitro gas production parameters and in vitro gas production characteristics of diets containing varying levels of sweet orange (Citrus sinensis) peel meal

II. Material and Method

Sweet orange peels were collected from sweet orange retailers who peel and sell oranges for direct consumption. They were sun-dried on concrete floor for 48 hrs and when dried, it was packed and crushed using a mortar and pestle and packed into synthetic bags. Four diets were compounded to contain 0%, 12.5%, 25% and 50% SOPM designated as treatment 1, 2, 3 and 4 respectively. Sample from each treatment was ground in a hammer mill to pass through a 1mm sieve. Each sample was analysed for crude protein, crude fibre, ether extract and ash according to the method of AOAC (1995). The *in vitro* gas production volume was determined

according to Menke and Steingass (1988). Rumen liquor was obtained from four male West African dwarf goats in the morning before feeding. 200mg of each sample was placed in a 120ml calibrated syringe in triplicates and 30ml innoculum containing cheese strained rumen liquor and buffer solution in a ratio of 1:4 was added to the 200mg of feed in the plastic syringes. Metabolizable energy (ME/MJ/kgDM) and organic matter digestibility (OMD%) were estimated as established by Menke and Steingass (1988), while short chain fatty acids (SCFA) were calculated as described by Getachew *et al.* (1998). Gas production characteristics were estimated using the equation Y=a+b (1-e^{-ct}) (Orskov and MacDonald, 1979), where Y = volume of gas produced in time t, a = gas produced from the soluble fraction, b = gas produced from the insoluble but fermentable fraction, c = gas production rate constant for the insoluble but fermentable fraction 'b' while t = incubation time. Metabolizable energy (ME, MJ/kgDM) and organic matter digestibility (OMD, %) were estimated as established by Menke and Steingass, 1988) and short chain fatty acids (SCFA) were computed as reported by Getachew *et al.* (1998). ME = 2.20 + 0.136GV + 0.057CP + 0.002CF, OMD = 14.89 + 0.889GV + 0.45CP + 0.651XA

SCFA = 0.0239GV - 0.0601, Where GV, CP, CF and XA = gas production volume (ml/200mgDM), crude protein (%), crude fibre (%) and ash (%) of the incubated sample.

Data collected were subjected to analysis of variance (ANOVA) using the Minitab (1991) software in completely randomized design. Significant differences were separated using the least square difference (LSD) as outlined by Akindele (1996).

III. Results and discussion

Table 1 presents the gross and chemical composition of the four diets. Crude protein values were between 14.72-16.41%, $D_1(16.41\%)$, $D_2(16.00\%)$ and $D_3(16.23\%)$ had similar values while $D_4(14.72\%)$ was lower. This was probably so because of the higher percentage of SOPM in D_4 and also that SOPM is lower in CP than maize offal. Observed values were comparable to 16.97 -17.48% reported by Arigbede et al. (2010) for diets containing graded levels of cocoa yam. Crude fibre values ranged between 11.05-14.22%, while the NFE was 40.61- 45.84%. The NFE values indicated that there was sufficient fermentable carbohydrates in the diets for energy production. Nitrogen free extract values were highest in $D_4(54.93)$ this may be as a result of the higher level of SOPM in D₄. Observed NFE values were comparable with 42.87-43.77% reported by Ukanwoko et al. (2009) for WAD goats fed cassava peel-cassava leaf meal, but it was higher than 28.04-32.97% reported by Maigandi and Abubakar (2004) for diets containing Faidherbia albida pods. Ash values which were between 9.01-9.82% indicated that, there was appreciable amount of minerals in the diets. Table 2 presents the *in vitro* gas production volumes, metabolisable energy, organic matter digestibility and short chain fatty acid of the diets containing varying levels of sweet orange peel meal. Values of IVGP for D₄(24.67ml/200mgDM) was significantly higher (p < 0.05) than those of D₁(15.33ml/200mgDM) D₂(17.33ml/200mgDM) and $D_3(20.33 \text{m}/200 \text{mgDM})$. This means that more carbohydrate fermentation took place in $D_4(50\% \text{SOPM})$ and also that D_4 had higher nutritive potentials than the other diets. The ME values were 5.30-6.50 MJ/kgDM, OMD was 40.57-49.18% and SCFA was 0.30-0.53 mmol/200mgDM. Diet 4 was significantly higher (p<0.05) in ME, OMD and SCFA, implying that D₄ had higher energy potentials than the other diets because high ME and SCFA implies energy availability. This agrees with the report of Sallam et al. (2007) that, high SCFA is an indicator of energy availability. Observed ME, OMD and SCFA values were comparable with the report of Abegunde et al. (2010) who reported 4.45-5.99 MJ/kgDM, 38.32-50.33% and 0.25-0.51 mmol/200mgDm for ME, OMD and SCFA respectively for *Ficus exasperata* diets. Table 3 shows the *in vitro* gas production characteristics of the experimental diets. The gas production characteristics of the diets did not show any significant difference (p>0.05) among the treatments. The soluble fraction 'a' (6.08-8.25ml/200mgDM) and the potential gas production 'b' (16.23-30.88ml/200mg) were all similar. This indicates that there was no difference in nutritive value among dietary treatments in the soluble carbohydrates and the insoluble but fermentable carbohydrates. The 'b' values were comparable with 19.00-26.00ml/200mgDM reported by Binuomote et al. (2010) for ensiled cassava tops and guinea grass mixture. The rate constant of fermentation 'c' for D_1 - D_4 which ranged between 0.03-0.07ml/hr was comparable with 0.04-0.06ml/hr reported by Binuomote et al. (2010) for ensiled cassava tops and guinea grass mixture.

IV. Conclusion

In vitro gas production parameters improved with increasing levels of SOPM in the diets, while there was no treatment effect on the gas production characteristics. The study showed that treatment with the highest percentage of SOPM had the highest energy potential, and increasing the levels of the test ingredient in the diets did not exert harmful effect on the activities of the rumen micro-organisms so as to cause retardation.

	(DM basis)						
Feed Ingredients		Experimental Diets					
	D_1	D_2	D_3	D_4			
Rice offal	20.00	20.00	20.00	20.00			
Maize offal	48.81	42.71	36.61	24.41			
Sweet orange peel meal	0	6.10	12.20	24.40			
Full fat Soyabean meal	28.19	28.19	28.19	28.19			
Bone ash	2.00	2.00	2.00	2.00			
Common salt	1.00	1.00	1.00	1.00			
Total	100.00	100.00	100.00	100.00			
Determined							
Crude prorein	16.41	16.00	16.23	14.72			
Crude fibre	14.22	13.16	11.05	12.27			
Ether extract	9.55	7.58	8.67	6.02			
Nitrogen free extract	50.81	53.84	54.93	57.93			
Ash	9.01	9.42	9.12	9.82			

Table 1:	Gross and chemical	composition	of experin	mental die	ets fed to	West African	Dwarf goats
			(D)	(f 1 : .)			

DM= Dry matter, T1 = 0%SOPM, T2 = 12.5%SOPM

T3 = 25%SOPM. T4 = 50%SOPM

Table 2: In vitro gas production	, metabolisable energy,	, organic matter dig	estibility and short
chain fatty acid	of diets containing gra	ded levels of sweet	orange peel meal

		In vitro parameters		
Diets	IVGP	ME (MJ/kgDM)	OMD (%)	SCFA (µmol/200mgDM)
D ₁ (0% SOPM)	15.33 ^b	5.30 ^c	40.57 ^c	0.30 ^d
D ₂ (12.5% SOPM)	17.33 ^b	5.52 ^{bc}	42.42 ^{bc}	0.35 ^b
D ₃ (25% SOPM)	20.33 ^b	5.39 ^b	41.41 ^b	0.32 ^c
D ₄ (50% SOPM)	24.67 ^a	6.56 ^a	49.18 ^a	0.53 ^a
SEM	2.08*	0.08*	0.51*	0.01*

a,b,c = Means on the same column with different superscripts are significantly different (p < 0.05) * = significant (p<0.05), SEM = Standard error of the mean, IVGP = In vitro gas production ME = Metabolisable energy, OMD = Organic matter digestibility, SCFA = Short chain fatty acid, and SOPM = Sweet orange peel meal.

I wore 5, in the cas production ended of the experimental dress (in 200 mgb it)	Table 3: In vitro gas	production	characteristics	of the ex	perimental	diets	(ml/200mgDN	1)
---	-----------------------	------------	-----------------	-----------	------------	-------	-------------	----

Fermentation	Treatment diets					
Characteristics	D_1	D_2	D_3	D_4	SEM	
а	7.81	8.25	6.08	7.70	1.37 ^{ns}	
a + b	24.04	27.49	23.79	38.59	10.17 ^{ns}	
В	16.23	19.20	17.70	30.88	9.15 ^{ns}	
С	0.05	0.03	0.07	0.05	0.01 ^{ns}	
Lag	8.58	8.30	5.30	5.40	2.02 ^{ns}	

^{ns} = Not significant, SEM = Standard error of the mean

a = Soluble fraction

a + b = Potentially degradable fraction

b = Potential gas production

c = Rate of fermentation constant

References

- Abegunde, T.O., Mako, A.A., Adene, I.C., Akinsovinu, A.O and Babavemi, O.J. (2010).
- [1]. [2]. Chemical compostion and in vitro gas production parameters of Ficus exasperate diets in the dry and wet season of Nigeria. Proc. 35^{th} Conf., Nig. Soc. for Anim. Prod. held on the 14^{th} - 17^{th} March, 2010, Univ. of Ibadan. Nig. Pp. 580 – 583.
- [3]. Adegbola, A. A. and Asaola, V. O. (1986). Preparation of cassava peels used in small ruminant production in Western Nigeria. Proc. of workshop held at the University of Alexandria. Egypt. Oct. 1985. ILCA. Addis Ababa. Ethiopia. Pp. 109-115.
- Aganga, A. A., Umunna, N. N., Okoh, P. N. and Oyedipe, E. O. (1986). Water metabolism of ruminants. J.Anim. Prod. Res. 6 (2): [4]. 171 - 187.
- Akindele, S. O. (1996). Basic Experimental Designs in Agricultural Research. Montem paperbacks "akure Nigeria. Pp. 25 34. [5].
- AOAC (1995). Official method of Analysis (16th edition) Association of official Analytical Chemists, Washington DC. [6].
- [7]. Babayemi, O.J., Bamikole, M.A and Odunguwa B.O. (2003). Haematological and biochemical components of West African dwarf goats fed Tephrosia bracteolata - based forage. Trop. Anim. Prod. Invest. 6:31-38.
- Babayemi, O. J. and Bamikole, M. A. (2006). Nutritive value of Tephosia candida seed in West African Dwarf goats. J. Cent. Eur. [8]. Agric. 7:4, 731 - 738.

Binuomote, R. T. and Babayemi, O. J. (2010). Nutritive evaluation of ensile cassava (Manihot esculentus, crants) tops and guinea [9]. grass mixtures. Proc. 35th Conf., Nig. Soc. for Anim. Prod. 14th - 17th March, 2011, Univ. of Ibadan, Nigeria. Pp. 601 - 603

Blummel, M., Makkar, H.P.S., Chisanga, G., Mtimuni and Becker, K. (1997). The prediction of dry matter intake and in vitro [10]. digestibility of African roughages in relation to ruminant live weight gain. Anim. Feed Sci. and Tech. 69:131-141.

- [11]. Getachew, G., Blummel, M., Makkar, H. P and Becker, K. (1998). *In vitro* gas measuring techniques for assessment of nutritional quality of feeds. A review. *Anim. feed. Sci. Technolo.* 72: 261 281.
- [12]. Lamidi, A.A., Aina, A.B.J and Sowande, S.O. (2010). Nutrient digestibility and nitrogen balance in West African dwarf goats fed blended diets for dry season. Proc. 35th Conf. Nig. Soc. Anim. Prod. held 14th – 17th March, 2010 at the Univ. of Ibadan, Nigeria. Pp. 499 – 501.
- [13]. Maigandi, S.A and Abubakar, S. (2004). Nutrient intake and digestibility by red Sokoto goats fed varying levels of *Faigherbial albida* pods. Proc. 29th Ann. Conf., Nig Soc. Anim. Prod. Vol. 29: 325 328.
- [14]. Menke, K. H and Steinguss, H. (1988). Estimation of the energetic feed value from chemical analysis and *in vitro* gas production using rumen fluid. *Anim. Res. Develop.* 28: 7 55
- [15]. Menke, K. H and Steinguss, H. (1988). Estimation of the energetic feed value from chemical analysis and *in vitro* gas production using rumen fluid. *Anim. Res. Develop.* 28: 7 55
- [16]. MINITAB (1991). Statistical Software. Vol.10. 2 MINITAB Inc. P.A., USA.
- [17]. Ogunbosoye, D. O. and Babayemi, O. J. (2010). Voluntary intake of Non-legume fodders offered simultaneously to West African Dwarf goats for a period of six hours. Proc. 35th Conf. Nig. Soc. for Anim. Prod. 14 – 17th March, 2010, Univ. of Ibadan, Nigeria. Pp. 518 – 520.
- [18]. Sallam, S.M.A., Nasser, M.E.A., El-Waziri, A.M., Bueno, I.C.S and Abdalla, A.L. (2007). Use of *in vitro* rumen gas production technique to evaluate feedstuffs. *J. Appl. Sci. and Res.* 3 (1): 34 41.
- [19]. Ukanwoko, A. I., Ibeawuchi, J. A. and Ukachukwu, N. N. (2009). Growth Performance and carcass characteristics of West African dwarf goats fed cassava peel-cassava leaf meal based diet. Proc. 34th Ann. Conf., Nig. Soc. for Anim. Prod. 15-18th March, 2009. Univ. of Uyo, Uyo. Pp. 476 – 479.
- [20]. Menke, K.H., Ranb, L., Salewski, A., Steingass., Fritz, D. and Schneider, W. (1979). The estimation of the digestibility and Metabolizable energy content of ruminant feedstuffs from gas Production when they are incubated with human liquors nitrogen. J. Agric. Sc. 93: 217 – 222.