# Effect of *In ovo*Injection of Various Nutrients on the Hatchability, Mortality Ratio and Weight of the Broiler Chickens

Najih Jabir Al-Shamery<sup>1</sup> and Mohammed Baqur S. Al-Shuhaib<sup>2</sup> Department of Animal Resources/College of Agriculture/Al Qasim Green University

**Abstract:** The main goal of this paper is to study the effect of injectingthe eggs embryos with certain levels of nutrient solutions (vitamin E 1.5 mg / egg, the amino acid lysine 1.5 mg / egg, the amino acid methionine 1.5 mg egg and a mixture of vitamin E and lysine and methionine 1.5 mg / egg) in the proportion of mortality of embryonic and hatchability and initial weight 1 g / day. Results wereindicated a significant decrease (P < 0.05) in the percentage of embryonic mortality in the treatmentinjection with methionine and the treatment of the mixture of vitamin E, lysine and methionine. Though arithmetic improvements in the percentage of hatching and in the initial weight of the hatched chicks (1g/day) were achieved in case of vitamin E injection treatment, these improvements weren't getinto the significance (P < 0.05) in the percentage of hatching ratio, and in the initial weight of hatched chicks. Moreover, arithmetic improvements weren't get the significance (P < 0.05) in case of vitamin E, lysine, and methionine mixture injection treatment. We conclude from this study, the in ovo possibility of injecting eggs embryoswith 1.5 mg / egg of vitamin E solution to improve hatchability and initial weight, methionine solution and lysine - methionine – vitamin E solution to reduce the proportion of embryonic mortality.

Keywords: Nutrients, In-ovo injection, Mortality, Weight, Broiler Chickens

### I. Introduction

Avian embryogenesis is an external process (Foye, 2005). Chicken embryos are dependent on the nutrients within the egg, which provide the energy and building blocks required for the metabolic needs of the growingembryo during the normal 21-day incubation (Foyeet al., 2006). One unique way of introducing nutrients to the incubating embryo is through in ovoinjection (Macalintal, 2012). Studies have shown that in ovoinjection of nutrients, such as carbohydrate, proteins, vitamins and amino acids as well as vaccines, can enhance growth and development of the embryo, improve energy status, promote early gut development and improve immune status, as well as alleviate the stress of hatching (Takoet al, 2005; Gore and Qureshi, 1987; Ohtaet al., 2001; Johnston et al., 1997; Gaafaret al.;., 2013). Thus, any embryonic growth and development inside the fertilized egg is taken place under the effect of the concentration and the type of nutrients that involved in the constitution of the fertilized egg (Al-Murraniet al., 1982). However, the injection of fertilized eggs with nutrients plays an important role in replacing any deficiency in the synthesis of food materials involved in the formation of eggs that is taken place as a result for the possible maternal malnutrition (Selimet al., 2012). Moreover, it was mentioned that in ovo injection with nutrients might be useful and much more efficient than the normal nutrition (Bhanjaet al., 2007). Add to that, in ovoinjection with nutrients reduces posthatching mortalities and diseases as long as enhances the immune responses (Gore and Qureshi, 1987). Also, early development by enhancing intestinal function and development is enhanced too (Takoet al., 2004).

The in ovo nutrient administration could be considered as an alternative method to improve hatchability and duckling weights since either vitamin E or ascorbic acid resulted in significantly (P < 0.05) higher body weights at hatch, final body weights, and feed intakes than the un-injected control group (Selimet al., 2012). Vitamin E is regarded as the major lipid soluble antioxidant vitamin, which plays in turn on preventing the oxidation of phospholipids that involved in the construction of brain cells (Suraiet al.,1999). The use of antioxidants, especially vitamin E, has been proven to reduce harmful peroxidation of lipids and cholesterols in animal models (Singh, et al., 2005). Therefore, developed and improved nutritional status afforded by in ovo feeding subsequently improved hatchability percentage, hatching weight, growth performance and immune responses (Al-Zuhairyet al., 2013; Bakyarajet al., 2012; Selimet al., 2012).Moreover, the injection of hatching eggs with vitamin E leads to an enrichment of eggs contents of this vitamin, thus, it's possible to be exploited for chicks' development and growth (Cherianet al., 1997;Suraiet al., 1999). The in ovo injected vitamin E plays a significant role in the increasing of fatty acids oxidation since the day 14 of hatching (Cherianet al., 1997).

It was referred that methionine has an active role in enhancing body weight and growth and a fundamental role in the formation of choline that considered as one of B complex compounds (Coskunet al., 2014). Add to that, the embryonic development might be enhanced after the injection of some amino acids in ovo (Al-Murraniet al., 1982).

It was suggested that DL-methionine may use as an in ovo feed additive for obtain heavier chick (Coskunet al., 2014). Moreover, the in ovo injection of amino acids reduces the danger of some diseases through blocking the deficiency in the concentration of these amino acids in hatching eggs in such away it leads into acceleration of immune response (Khalid et al., 2013). Therefore, this study was designed to investigate the effects of administration of a mixture of amino acids and vitamins ((vitamin E); (Lys); (Met); (vitamin E + Lys + Met)) at a late stage of incubation on hatchability traits and performance of hatching chicks.

## **II.** Materials And Methods

**2.1 Eggs pre-injection incubation:** Standard size fertile broilers eggs (n= 200) were collected from a commercial breeders chicks from a maternal flock of only one strain (Ross) that fed on adequate nutritional diet. Eggs were distributed into five groups of 40 eggs each. The eggs were incubated under optimal conditions (at 37 to 37.5°C and 70% relative humidity) at the Department of Animal Resources at Al-Qasim Green University.

**2.2 Eggs injection:** The groups of the chosen fertile eggs were distributed into five groups. Each treatment that contained 40 eggs was randomly distributed on two repeats. Each egg was injected with 0.1ml (1.5%) nutrient saline as the following;  $T_1$  control treatment (without injection),  $T_2$  injection treatment with vitamin E (1.5 mg/egg),  $T_3$  injection treatment with lysine (1.5 mg/egg),  $T_4$  injection treatment with methionine (1.5 mg/egg), and  $T_5$  injection treatment with the mixture of vitamin E, lysine, and methionine (1.5 mg/egg). The right eggs for hatching were chosen with average weight 62 g/bird (± 1g). Next Day, the package was introduced into the hatcher. The candling was performed at 7 days after hatching depending on each individual treatment. Before injecting the desired solution, the surface of eggs was disenfected by 70% ethanol. In their broad end, the eggs were pierced by a fine needle (gauge 23) of the commercially available 1ml insulin syringe.

**2.3 Eggs post-injection incubation:** After performing injection, the egg hole was sealed with a sterile paraffin wax immediately after injection. The injected eggs were returned into the incubator. After hatching, the candling was repeated again after 24 and 18 days to identify the mortalities ratio in these ages. After the last test, the eggs were transported into the hatcher to complete hatching. After 21 days after hatching, the hatched and non-hatched eggs ratios were calculated. The initial body weight of the hatched chicks of one day age was calculated as well.

**2.4 Statistical Analysis**: The obtained data were represented by the completely randomized design (CRD) to analyze the effect of different treatments on the studied characters. Analysis of variance (ANOVA) was used to test the significance of the difference between different treatments and statistical differences were established using a Duncan's Multiple Range Test (Duncan, 1955) at the level of P < 0.05. Also, the significant differences were compared among the averages were represented by using SAS program (2001) in the statistical analysis according into the following equation;

 $Y_{ijk} = M + T_i + e_{ijk}$ 

### **III. Results And Discussion**

The Different nutrients supplements that in ovo administrated in the present study had shown an increased rate of hatchability and initial body weight. It was noticed from table (1) the presence of a significant reduction of the embryonic mortality at 14 days after hatching at significant level (P < 0.05) for the mixture injection treatment with the nutrient solutions (T<sub>5</sub>) that notified (0.0%). On the other hand, an important reduction for methionine injection (T<sub>4</sub>) treatment was occurred but without reaching the level of significance(P < 0.05). In T4 treatment the ratio of embryonic mortalities was estimated to be 3.33% compared with T1 control treatment and T<sub>2</sub> and T<sub>3</sub> injection treatments, in which any significant differences among them weren't observed. However, the differences among T<sub>1</sub>, T<sub>2</sub>, and T<sub>3</sub> treatments were only 6.66%. Add to that, any significant difference between T<sub>5</sub> and T<sub>4</sub> injection treatment wasn't observed as well.

Concerning the effect of the nutrient solutions in the embryonic mortalities ratio after 18 days of hatching, it was noticed that there is a significant reduction (P < 0.05) in T5 injection treatment since it reached 0.0%. Though it was an important reduction, it wasn't reached the level of significance concerning  $T_2$  injection treatment, which reached 7.17% compared with  $T_1$  control treatment and other injection treatments ( $T_3$  and  $T_4$ ), in which any significant differences weren't observed. These results were come in accordance with what it was come out some papers (Selimet al., 2012). Since vitamin E participated in turn in the preventing of the oxidation phospholipids that involved in the growth and development of brain cells and other variable embryonic tissues properly (Dror and Bartov, 1982; Bhanjaet al., 2007). Moreover, this vitamin acts as a reducer for free radicals that cause deleterious damages for unsaturated fatty acids that existing in the embryonic cellular membranes (Cherian and Sim, 1992 and 1997).

The effect of injection of nutrient solutions into the eggs didn't hatched was clarified by the reduction of the injection treatment with methionine ( $T_4$ ) compared with the control treatment ( $T_1$ ) and other treatments ( $T_2$ ,  $T_3$ , and  $T_4$ ). But these reductions were only mathematical and weren't getting the level of significance (P < 0.05). These results, however, were come in accordance several observations that reported a significant reduction in the embryonic mortalities in injection treatments with lysine and methionine, in which the reason behind the reduction of mortal embryos might be attributed into the important roles played by these two amino acids in the reduction of glycolysis (Uni and Ferkit, 2003, Peebles et al., 2006). Consequently, the body energy resources will be conserved inside the body and the energy level will be elevated through the formation glucose from non-carbohydrate sources, such as amino acids (Sunny et al., 2007).

Table (1): the effect of injecting eggs with numeric solutions on the ratio of mortainty									
Treatment	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	$T_4$	T <sub>5</sub>				
Feature									
Embryonic mortalities ratio (14 days after	6.66 a	6.66 a	6.66 a	3.33 ab	0.0 b				
hatching)	±2.67	$\pm 2.67$	$\pm 2.67$	$\pm 1.69$	$\pm 0.70$				
Embryonic mortalities ratio (18 days after	10.71 a	7.17 a	10.71 a	14.28 a	0.0 b				
hatching)	$\pm 3.30$	$\pm 2.76$	$\pm 3.30$	$\pm 3.72$	$\pm 0.70$				
Eggs didn't hatched ratio	8.01 a	3.84 b	8.33 a	11.68 a	9.99 a				
	±2.91	$\pm 1.78$	$\pm 2.42$	$\pm 3.47$	± 3.19				

**Table (1):** the effect of injecting eggs with nutrient solutions on the ratio of mortality

• Different letter within the same column indicate to the presence of significant differences among treatments within the probability levels (P < 0.05)

It was noticed from table (2) the presence of an important superiority in the hatching ratio for the injection treatment with vitamin E ( $T_2$ ), and the injection treatment with methionine ( $T_5$ ) compared with the control treatment ( $T_1$ ) and the remaining injection treatments ( $T_3$  and  $T_4$ ). But, this superiority was only mathematical, and wasn't getting the level of significance (P < 0.05). These results were come in accordance with the results obtained by Selim and his colleagues (2012). Vitamin E plays on the increasing of fatty acids oxidation that begins at the day 14 after hatching (Chereian and Sim, 1992 and 1997). The situation was the same for the initial weight at one day age, in which it was noticed that superiority of the injection treatments ( $T_3$ ,  $T_4$ , and  $T_5$ ). But this superiority was mathematical and wasn't getting the level of significance (P < 0.05). The in ovo injection of vitamin E plays on the increasing of the vitamin E egg's content, and consequently it will be exploited in the development and growth in addition to the ability of chick to utilize of this vitamin after hatching (Cherianet al., 1997; Suraiet al., 1999).

for natoriou enters (5, 1 day).									
Feature	$T_1$	T <sub>2</sub>	T <sub>3</sub>	$T_4$	T <sub>5</sub>				
Hatching ratio	83.33 a ±9.15	89.99 a ±9.53	83.33 a ± 9.13	76.66 a ± 8.78	89.99 a ± 9.51				
Initial body weight 1g/day	42.50 a ±4.25	44.0 a ± 4.40	42.50 a ± 4.23	$42.50 \text{ a} \pm 4.28$	43.50 a ± 4.35				

**Table (2):** the effect of injecting eggs with nutrient solutions on the hatching ratio and initial weight for hatched chicks (g/1 day).

• Different letter within the same column indicate to the presence of significant differences among treatments within the probability levels (**P** < **0.05**)

### **IV.** Conclusion

It may be concluded that early supplementation of nutrients through inovoinjection such as amino acids and vitamin mixture can beregarded as a possible method to improve hatchability, body weight at hatch, marketing weights of broilers. Further investigations are needed to highlight the effect of in-ovo injection of amino acids mixture on the humoral, cell mediated immunity, and development of digestive organs in broilers.

#### References

- [1]. Al-Murrani,W.K.1978.Maternal effect on embryonic and post embryonic growth inPoultry.Br.poult. Sci. 19:277-281.
- [2]. Al-Murrani,W.K.1982.Effect of injecting amino acids into the egg on Embryonic and Subsequent growth in the domestic fowl.Br.Poultry.Sci.23:171-174.
- [3]. Al-Zuhairy M., Alasadi Y. Effect of in ovo injection with Newcastle disease vaccine, multivitamins AD3E, and Omega-3 on performance and immune response of broiler. I.J.A.B.R, 3(2) 2013: 208-211.
- [4]. Bakyaraj, S., S. K. Bhanja, S. Majumdar and B. Dash. 2012. Modulation of post-hatch growth and immunity through in ovo supplemented nutrients in broiler chickens. J. Sci. Food Agric. 92:313-320.
- [5]. Bhanja, S.K., Mandal A.B., AgarwalS.K., MajumdarS. and Bhattacharyya A.2007.Effectof in ovo injection of vitamins on the chick weight and post-hatch Growth performancein broiler chickens .World poultry Science Association, Proceedings of the 16<sup>th</sup>Eurpean Symposium on poultry Nutrition, India.

- [6]. Cherian ,G. and J. Sim. 1992.Preferential accumulation of n-3 fatty acids in the brain of chicks from eggs enriched with n-3 fatty acids. Poult.Sci.71:1658-1668.
- [7]. Cherian ,G. and J. Sim. 1997. Egg yolk polyunsaturated fatty acids and vitamin EContent alters the tocopherol status of hatched chicks.Poult.Sci.76:1753-1759.
- [8]. Coşkun I., Erener G., Şahin A., Karadavut U., Altop A., Okur A. (2014). Impacts of In Ovo Feeding of DL-Methionine on Hatchability and Chick Weight. Turkish Journal of Agriculture - Food Science and Technology, 2(1): 47-50.
- [9]. Dror, Y. and Bartov, i. 1982.Dietary factors affecting experiment models of nutritionalEncephalomalacia.poult.Sci.61:84-93.Duncan, B.D, 1955. Multiple range and multiple test. Biometrics, 11:1-42.
- [10]. Foye OT. The biochemical and molecular effects of amnionic nutrient administration, "in ovo feeding" on intestinal development and function and carbohydrate metabolism in turkey embryos and poults. A Dissertation submitted to the Graduate Faculty of North Carolina State University in partial fulfillment of the requirements for the Degree of Doctorate of Philosophy in Nutrition Department of Poultry Science/Nutrition Raleigh, NC, 2005.
- [11]. Foye OT, Uni Z, Ferket PR. 2006. Effect of in ovo feeding egg white protein, β-hydroxy-β-methylbutyrate, and carbohydrates on glycogen status and neonatal growth of turkeys, Poult Sci., 85: 1185-1192.
- [12]. Gore, A. B. and M. A. Qureshi.1987. Enhancement of humoral and cellularImmunitybyvitamin Eafter embryonic exposure. Poultry.Sci.76:984-991.
- [13]. Gaafar K.M., Selim S.A., El-ballal S.S. 2013.Effect of in-ovo administration with two levels of amino acids mixture on the performance of Muscovy ducks.Emir. J. Food Agric. 25 (1): 58-65.
- [14]. Johnston P. A., H. Liu, T. O'Connell, P. Phelps, M. Bland, J. Tyczkowski, A. Kemper, T. Harding, A. Avakian, E. Haddad, C. Whitfill, R. Gildersleeve, and C. A. Ricks. 1997. Applications in ovo technology. PoultSci 76:165-178.
- [15]. Khalid, M. Gaafar, ShaimaaA. Selimand Salah .S. El-ballal.2013. Effect of in ovoadministration with two levels of amino acids mixture on the performance of Muscovy ducks .Emir .J . Food Agric. 25(1):58-65.
- [16]. Macalintal L. M. 2012. In ovo selenium (SE) injection of incubating chicken eggs: effects on embryo viability, tissue SE concentration, lipid peroxidation, immune response and post hatch development. Theses and Dissertations--Animal and Food Sciences, University of Kentucky UKnowledge.
- [17]. Ohta Y, Kidd MT. 2001. Optimum site for in ovo amino acid injection in broiler breeder eggs. Poult. Sci., 80: 1425-1429.
- [18]. SAS, 2001. SAS User's Guide: Statistics (version 6.0) SAS Inst. Inc. Cary, NC, USA.
- [19]. Peebles, E.D., W.D. Berry, R.W. Keris, L.W. Bennett, and P.D.Gerard. 2006.Effect of injected gluconeogenic supplementation on the performance of broiler from young breeders.Poult.Sci.8:371-376.
- [20]. Selim ,Sh.A., K.M. Gaafar and S.S. El-ballal.2012.Ifluence of in ovo administration with vitamin E and ascorbic acid on the performance of Muscovy ducks. Emir. J. Food. Agric. 24(3):264-271.
- [21]. Singh, U., Devaraj, S. and Jialal, I. (2005) Vitamin E, oxidative stress, and inflammation. Annu. Rev. Nutr. 2005. 25:151–174.
- [22]. Surai, P.F., R. C. Noble, B. K.Speake.1999.Relationship between vitamin EContent and susceptibility to lipid peroxidation in tissues of the newly Hatched chick. Br. Poult. Sci.40:406-410.
- [23]. Sunny, E., Admany J., Owens S.L. ,Bequette B.J. 2007.Gluconeogenesis and Carbon utilization in day 20 chicken embryo supplemented in ovo with glucose and amino acid. Poult. Sci. 86(suppl 1):214.(Abstract).
- [24]. Tako,E., P.R. Ferket and Z. Uni.2004.Effects of in ovo feeding of carbohydrates and beta-hydroxy-beta-methyl butyrate on the development of chicken intestine.Poult.Sci.83:2023-2028.
- [25]. Tako E, Ferket PR, Uni Z. 2005. Changes in chicken intestinal zinc exporter mRNA expression and small intestine functionality following intra-amniotic zinc-methionine administration.JNutr.Biochem., 15: 339-346.
- [26]. Uni,Z., and P.R.Ferket. 2003. Enhancement of development of oviparous species by in ovo feeding.Patent 6,529,878 .North Carolina state university, Raleigh, NC.