

## Profile of Staphylococcus Aureus Associated With HIV Patients in University Of Maiduguri Teaching Hospital (UMTH), Maiduguri, Borno State

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### Abstract:

**Aims:** This study was aimed at determining the characteristic of nasal carriage status of HIV patients. *Staphylococcus aureus* has emerged as a significant opportunistic pathogen among HIV and AIDS patients in both nosocomial and community settings.

**Methodology and results:** A total of 200 nasal swabs were collected and analysed bacteriologically. Profile of 42 (21.0%) were characterized as *Staphylococcus aureus*. Gender distribution of HIV positive patients with *Staphylococcus aureus* infections were 16 (38.0%) males and 26 (61.9%) females respectively. The age group with high positive *Staphylococcus aureus* (38.09%) was seen among 31- 40years while the least was among 0 – 20years (2.3 %). *Staphylococcus aureus* exhibited alpha 7(16.7%), beta 22(52.4%) and gamma 22(52.4%) haemolysis on blood agar.

**Conclusion, significance and impact study:** High prevalence of nasal carriage among these patients (21.0%) has been documented. These results provide strategies to prevent systemic infections by eliminating nasal carriage of *Staphylococcus aureus*. Antibiotic susceptibility pattern of the isolates was high to quinolone drugs. Therefore the use of nasal topical antibiotics as prophylaxis is highly recommended.

**Key words:** *Staphylococcus aureus*, HIV profile, Maiduguri, Nigeria.

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### I. Introduction

*Staphylococcus aureus* has emerged as a significant opportunistic pathogen among HIV and AIDS patients in both nosocomial and community settings, and recent studies have shown greater frequency and morbidity of this organism among HIV positive individuals (Chacko et al., 2009., Hidron et al., 2010). Clinical manifestations ranged from superficial to systemic disease conditions, responsible for high morbidity and mortality rate (Lowry et al., 1998). *Staphylococcus aureus* normally localize in the skin and mucous membranes in the nose of healthy humans and about 30% of the normal healthy population are transiently colonised by the organism (Liu., 2009). There are more than 30 species of coagulase negative *Staphylococci* (CNS). *S. epidermidis* and *S. saprophyticus* are the species most often associated with infection but *Staphylococcus capitis*, *Staphylococcus cohnii*, *Staphylococcus haemolyticus*, *Staphylococcus hominis*, *Staphylococcus lugdenensis*, *S. schleiferi* subspecies *schleiferi*, *Staphylococcus simulans* and *Staphylococcus warneri* have also been implicated (Chacko et al., 2009). The unique characteristic of *S. aureus* isolates is the diverse mechanism of antibiotic resistance and variety of virulence factors responsible for establishment of staphylococcal infections (Lowry et al., 1998). Of the *Staphylococcus aureus* strain that had attracted public health concern globally is the methicillin-resistant *Staphylococcus aureus* (MRSA), because of its multi-resistant pattern to all classes of antimicrobial agents and rapid dissemination within hospital environment (Grundmann et al., 2006).

*Staphylococcus aureus* is an important pathogen in patients with HIV infection. Despite multiple reports on the severity and recurrent nature of *S. aureus* infection, the factors predisposing patients infected with HIV to *S. aureus* infection have not been well studied (Bowersox et al., 1999).

Apart from being primary causative agent of nosocomial infections, MRSA isolates are being detected in the community setting in both pediatrics and adult population, termed as Community-associated *S. aureus* (CAMRSA) (Groom et al., 2001). Due to rapid dissemination of MRSA isolates, it has been reported worldwide. However, the prevalence of MRSA isolates varies with type of health institution, studied population, geographical location and subclinical condition of the patients. In Asia, the prevalence level is approximately 80%, in Europe, 20-35% in US, 20-40% and Latin America, it is up to 70% (Diekema et al., 2001). In sub-

Saharan African, the level is assumed to be high in countries like South Africa, Zimbabwe, Egypt and Algeria, while low in countries like Sudan and Somali (Adesida et al., 2006).

A number of recent investigations have indicated that *Staphylococcus aureus* is the main aetiological agent of many infections in Nigeria (Anah et al., 2008., Adeleke et al., 2009., Bekibele et al., 2009).

This work was aimed at determining the phenotypic variability of nasal carriage *Staphylococcus aureus* among HIV patients in UMTH.

## **II. Materials And Methods**

### **Study Area**

The study was carried out in the University of Maiduguri Teaching Hospital, located at Maiduguri, the capital of Borno State, bordered by 3 countries, Republic of Cameroon to the East, Chad to the Northeast, Niger to the North. Maiduguri is a cosmopolitan town which is inhabited by various ethnic groups.

### **Study Population**

A total of 200 HIV-positive patients attending PEPFAR accredited clinic at the University of Maiduguri Teaching Hospital were recruited into the research study. Consent forms were administered for this study. Study questionnaire were administered on identified HIV patients that signed the consent forms. Demographic information on the questionnaire included, age, sex, duration of diagnosis, type of drug and other associated clinical details.

### **Sample Collection**

The nasal swab was taken by inserting a sterile swab into the patients' anterior nares and gently rotated. The nasal swab were properly labeled and taken to the laboratory for analysis.

### **Microbiological Analysis and Identification**

All samples (nasal swabs) were analysed bacteriologically as described by Esan et al., (2009).

### **Characterization of *Staphylococcus aureus*:**

#### **DNase Test**

DNase test was performed according to the method previously described by Kateete et al., (2010).

#### **Haemolytic activity**

The haemolytic activity testing of *Staphylococcus aureus* isolates was performed according to the method previously described by Jimenez et al., (2008).

#### **□-Lactamase test**

The β-lactamase test was performed using the tube based iodometric method as previously described (Oncel et al., 2004).

#### **Detection of methicillin-resistant *Staphylococcus aureus***

Mannitol salt agar was prepared and performed according to previously described method by Arora et al., (2010).

#### **Antibiotic susceptibility profiles**

The in-vitro susceptibility of *Staphylococcus aureus* isolates to various routine antimicrobial drugs was tested by the standard disc diffusion technique using guidelines established by NCCLS., (2002).

#### **API Staph-Ident System**

Identification of *Staphylococcus aureus* was based on conventional criteria (including the coagulase tube test and the API Staph system [ATB32 Staph, BioMérieux, Marcy-l'Etoile, France]). API staph – Identification test was performed as documented by Christof et al., (2001).

#### **Statistical analysis**

All data was entered into a Microsoft excel sheet. The analysis was conducted using the statistical package for social sciences (SPSS) program, version 17.1. Chi-square test was used for comparison of the different variables and the correlation between all the tests performed. A p value of <0.05 was considered to be statistically significant.

### Ethical clearance

Ethical clearance was obtained from UMTH research committee and Principal investigator of PEPFAR before the commencement of the study.

## III. Results

### Demographic Information

Of 200 nasal swab samples analysed, 92 (46.0%) were from males and 108 (54.0%) females. Forty-two (21.0%) of 200 nasal swabs were positive for *S. aureus* by both tube coagulase and DNase test respectively. Of the 42(21.0%) *S. aureus* positive nasal swabs, 16 (38.09%) were males and 26 (61.9%) were females, and duration of HIV diagnosis were within 2weeks and 10years. The age grouping of the patients with nasal carriage for *S. aureus* as presented in Table 2, showed high frequency within 31 – 40 years (38.09), followed by 21 – 30 years (35.7%) and least percentage among 0 – 20 years (2.3%). Table 3: showed the phenotypic characteristic of *S. aureus* isolated.13(30.9%) exhibited  $\alpha$ -haemolysis, 7 (16.7%)  $\beta$ -haemolysis and 22 (52.4%)  $\gamma$ -haemolysis.

The antibiotic susceptibility pattern of the *S. aureus*, showed that 78.6% isolates were sensitive to Ciprofloxacin (CPX).

**Table 1. Sex distribution of patients with *Staphylococcus aureus* infection**

SEX	TEST SAMPLES(%)	NO. POSITIVE(%)
MALE	100 (50.0)	16 ( 16 )
FEMALE	100 (50.0)	26 ( 26 )
TOTAL	200 (100)	42 ( 42 )

**Table 2. Age group distribution of patients with *Staphylococcus aureus* infection**

Age group (years)	Frequency (%)
0 – 20	1 (2.30)
21 – 30	15 (35.7)
31 – 40	16 (38.09)
41 – 50	8 (19.04)
≥ 51	2 (4.76)
Total	42 (100)

**Table 3. Profile of *Staphylococcus aureus* isolates in the HIV positive patients**

Biochemical parameter	Catalase	Coagulase	Haemolysis			DNase
			$\alpha$	$\beta$	$\gamma$	
No. tested	42	42	42	42	42	42
No. positive	42	42	13	7	22	42

## IV. Discussion

*Staphylococcus aureus* are both commensal organism and versatile pathogen capable of causing a wide range of human diseases resulting in high morbidity and mortality in tropical Africa. Over the past two decades, there has been an increase in the rate of infection and diseases caused by *S. aureus* particularly MRSA throughout the world (Sadaka et al., 2009). The situation is even more alarming among patients with reduced immunity such as those undergoing chemotherapy or surgery, children, elders and patients with HIV and AIDS.

The prevalence of nasal carriage of *Staphylococcus aureus* documented in this finding (21.0%) is similar to previous studies but however considered low when compared to other findings in North Central (Abuja, 78%) and SouthEastern (Aba, 80%) Nigeria as documented by Onunuga et al., 2005. The anterior nares are the main ecological niche for *Staphylococcus aureus* and the natural history of nasal carriage of *Staphylococcus aureus* in HIV-infected patients has not been well delineated.

The differences in the prevalence level as observed in our study might be related to the number and clinical condition of the patients, and the duration of study. This is further emphasized by the gender distribution of *Staphylococcus aureus* among HIV patients with high positive nasal carriage of 42% (males 16% and females 26%). The distribution of nasal carriage of *S. aureus* has been documented (Hidron et al., 2010) by the behavioral influence, social, environmental, biologic, HIV host-specific risk factors, and probably, a combination of all these play a significant role in explaining the increased prevalence and incidence identified with female patients investigated. High prevalence of *Staphylococcus aureus* was observed among the age group 21- 40 years and this may be due to activities like nose picking. This further buttresses the findings of Christof et al., (2001) who reported persistent nasal carriage of *Staphylococcus aureus* in children. Although nasal carriage of *S. aureus* has been suggested as the source of subsequent infections, previous studies were limited to single hospitals (Mest et al., 1994., Weinke et al., 1992., Ena et al., 1994) or to defined patient groups such as patients infected with the human immunodeficiency virus or receiving hemodialysis (Chow and Yu, 1989., Holton et al., 1991).

Intake of the ARD has little or no effect on the nasal carriage of *Staphylococcus aureus* as the highest number of isolates was recorded among those on ARD intake for over five years. The least number of isolates came from those on ARD for less than a year.

The reason for the higher colonization rates observed are unclear, but could include factors such as frequent contact with both health care and community settings and frequent exposure to antibiotics, leading to a greater likelihood of becoming colonized with resistant strains. Some authors argued that this increased susceptibility to colonization with *S. aureus* could be HIV-specific because of depleted immunity (Sharpiro et al., 2000).

*S. aureus* is the most frequent cause of both community and hospital-acquired bacteremia in HIV-positive patients (Pedro-Botet et al., 2002., Stroud et al., 1997) and MRSA can explain 32%–67% of cases of *S. aureus* bacteremia among HIV patient population as previously reported (Tumbarello et al., 2002., Senthilkumar et al., 2001., Uche and Forrest, 2006).

Phenotypic tests are costly in resource limited settings but the mainstay in the diagnosis of staphylococcal infections, in which coagulase tests are usually confirmatory (Christof et al, 2001) as reported in our study. Phenotypic characteristic investigated included production of hemolytic activities among other tests as in the assessment of the virulence factors documented in a similar study by Samie and Shivambu (2011) and antibiotic susceptibility (Esan et al, 2009).

There is no single phenotypic test (including the tube coagulase test) that can guarantee reliable results in the identification of *Staphylococcus aureus*.

Therefore, the ideal identification of *Staphylococcus aureus* clinical isolates requires a battery of tests such as API Staph. Modern methods of molecular typing, which are highly discriminatory are also recommended. These data will improve on the identification of *Staphylococcus aureus* in clinical specimens.

## V. Conclusions

In the light of the forgoing, therefore, the finding of this study has opened up the epidemiological pattern of *Staphylococcus aureus* in HIV AIDS patients in the study area. Similarly, it is believed that information generated from the study could serve as a guide in clinical management of HIV/AIDS patients with *S. aureus* infection and baseline for further epidemiological studies. Some of the risk factors for colonization among HIV-infected patients suggest immunologic and virologic control, as well as the use of prophylaxis as control or protective measures.

**Conflict of interest:** The authors declared no conflict of interest.

## References

- [1]. Adeleke SI, Asani MO (2009). Urinary tract infection in children with nephritic syndrome in Kano, Nigeria. *Ann. Afr. Med.* 8: 38-41.
- [2]. Adesida S, Boelens H, Babajide B, Kehinde A, Snijders S, Van Leeuwen W, Coker A, Verbrugh H, Van Belkum A (2005). Major epidemic clones of *Staphylococcus aureus* in Nigeria. *Microb. Drug Resist.* 11: 115-121.
- [3]. Anah MU, Udo JJ, Ochigbo SO, Abia-Basse LN (2008). Neonatal septicaemia in Calabar, Nigeria. *Trop. Doct.* 38: 126-128.
- [4]. Arora S, Devi P, Arora U, and Devi B (2010). Prevalence of Methicillin-resistant *Staphylococcus aureus* (MRSA) in a Tertiary Care Hospital in Northern India. *J. Lab. Phys.*, 2(2): 78–81.
- [5]. Bowersox, J., (1999) Experimental StaphVaccine Broadly Protective in Animal Studies". <http://web.archive.org/web/20070505050641/http://www3.niaid.nih.gov/news/newsrelases/1999/staph.htm>.
- [6]. Bekibebe CO, Kehinde AO, Ajayi BG (2009). Upper lid skin bacterial count of surgical eye patients in Ibadan, Nigeria. *Afr. J. Med. Med. Sci.* 37: 273-277.
- [7]. Chacko J, Kuruvila M, Bhat AK (2009). Factors affecting the nasal carriage of MRSA in human immunodeficiency Virus- infected patients. *Indian J. Med. Microbiol.*, 25: 146-148.
- [8]. Christof V. E., Karsten B., Konstanze M., Holger S., and Georg P(2001) Nasal carriage as a source of *Staphylococcus aureus* bacteremia. *Engl J. Med.*, Vol. 344, No. 1
- [9]. Chow J.W, Yu VL. (1989) *Staphylococcus aureus* nasal carriage in hemodialysis patients: Its role in infection and approaches to prophylaxis. *Arch Intern Med* 149:1258-62.
- [10]. Diekema, D.J., Pfaller, M.A., Schmitz, F.J., Smayevsky, J., Bell, J., Jones, R.N., and Beach, M.; SENTRY Participants Group: (2001). Survey of infections due to *Staphylococcus* species: Frequency of occurrence and antimicrobial susceptibility of isolates collected in the United States, Canada, Latin America, Europe, and the Western Pacific region for the SENTRY Antimicrobial Surveillance Program, 1997- 1999. *Clin. Infect. Dis.* 32 (Suppl. 2), S114-S132
- [11]. Esan, Clement Olawale, Oladiran Famurewa, Johnson Lin and Adebayo Osagie Shittu, (2009) Characterization of *Staphylococcus aureus* isolates obtained from health care institutions in Ekiti and Ondo States, South-Western Nigeria. *African Journal of Microbiology Research* Vol. 3(12) pp. 962-968
- [12]. Ena J, Boelaert J.R, Boyken L.D, Van Landuyt H.W, Godard C.A, Herwaldt L.A. (1994) Epidemiology of *Staphylococcus aureus* infections in patients on hemodialysis. *Infect Control Hosp Epidemiol*;15:78-81.
- [13]. Grundman H Sousa, MA, Boyce J Tickersima (2006). Emergency and resurgence of methicillin-resistant *Staphylococcus aureus* as a public health threat, 68854-3.
- [14]. Groom, AV, Wolsey DH, Naimi TS, Smith K et al (2001). community-acquired methicillin *Staphylococcus aureus* in a rural American Indian community. *J. AM Med association*:1201-1205.
- [15]. Hidron A, Kempker R, Moanna A, Rimland D (2010). Methicillin-resistant *Staphylococcus aureus* in HIV-infected patients. *Infect. Drug. Res.*, 3: 73–86

- [16]. Holton DL, Nicolle LE, Diley D, Bernstein K (1991) Efficacy of mupirocin nasal ointment in eradicating *Staphylococcus aureus* nasal carriage in chronic haemodialysis patients. *J. Hosp Infect*;17:133-7.
- [17]. Jimenez E, Delgado S, Fernandez L, Garcia N, Albuja M, Gomez A, Rodriguez JM (2008). Assessment of the bacterial diversity of human colostrums and screening of *Staphylococcal* and *Enterococcal* population for potential virulence factors. *Res. Microbiol.*, 159: 595- 601.
- [18]. Liu GY (2009). Molecular Pathogenesis of *Staphylococcus aureus* Infection. *Int. Ped. Res. Found.*, 65: 71-77.
- [19]. Kateete et al. (2010) *Annals of Clinical Microbiology and Antimicrobials*, 9:23 <http://www.ann-clinmicrob.com/content/9/1/23>
- [20]. Lowry, F.D., (1998). *Staphylococcus aureus* infection. antibiotics, such as penicillin G and ampicillin. *N. England J. Med.*, 339: 520-532.
- [21]. Mest DR, Wong DH, Shimoda KJ, Mulligan ME, Wilson SE (1994) Nasal colonization with methicillin-resistant *Staphylococcus aureus* on admission to the surgical intensive care unit increases the risk of infection. *Anesth Analg*;78:644-50.
- [22]. National Committee for Clinical Laboratory Standards (2002). Performance standards for antimicrobial disc and dilution susceptibility tests for bacteria isolated from animals. Approved Standard 5th ed. M31-A.
- [23]. Oncel,T., Iça, T., Akan, M (2004) Beta lactamase production rate and antimicrobial susceptibility of *Staphylococcus aureus* isolated from clinical and subclinical mastitis cases in Turkey. *Revue Med Vet* 155, 7, 385-388
- [24]. Onanuga A, Oyi AR, Onaolapo JA (2005). Prevalence and susceptibility pattern of methicillin resistant *Staphylococcus aureus* isolates among healthy women in Zaria, Nigeria. *Afr. J. Biotechnol.*, 4: 1321-1324.
- [25]. Pedro-Botet ML, Modol JM, Valles X, et al. (2002) Changes in bloodstream infections in HIV-positive patients in a university hospital in Spain (1995–1997). *Int J Infect Dis*.6(1):17–22.
- [26]. Sadaka SM, El-Ghazzawy EF, Harfoush RA, Meheissen MA (2009).Evaluation of different methods for the rapid diagnosis of methicillinresistance in *Staphylococcus aureus*. *Afr. J. Microbiol.*, 3:049-055.
- [27]. Shapiro M, Smith KJ, James WD, et al. (2000) Cutaneous microenvironment of human immunodeficiency virus (HIV)-seropositive and HIV-seronegative individuals, with special reference to *Staphylococcus aureus* colonization. *J. Clin Microbiol*: 38(9):3174–3178.
- [28]. Stroud L, Srivastava P, Culver D et al. (1997) Nosocomial infections in HIV-infected patients: preliminary results from a multicenter surveillance system (1989–1995). *Infect Control Hosp Epidemiol*.18(7):479–485.
- [29]. Senthilkumar A, Kumar S, Sheagren JN (2001). Increased incidence of *Staphylococcus aureus* bacteremia in hospitalized patients with acquired immunodeficiency syndrome. *Clin Infect Dis*.;33(8):1412–1416.
- [30]. Samie, A., Shivambu, N. (2011) Biofilm production and antibiotic susceptibility profiles of *Staphylococcus aureus* isolated from HIV and AIDS patients in the Limpopo Province, South Africa. *African Journal of Biotechnology* Vol. 10(65), pp. 14625-14636.
- [31]. Tumbarello M, de Gaetano Donati K, Tacconelli E, et al. (2002). Risk factors and predictors of mortality of methicillin-resistant *Staphylococcus aureus* (MRSA) bacteraemia in HIV-infected patients. *J Antimicrob Chemother*.50(3):375–382.
- [32]. Uche A, Forrest G. (2006).*Staphylococcus aureus* bacteremia in HIV-infected patients in the HAART era. Presented at 44th Annual Meeting of the Infectious Diseases Society of America (IDSA). Boston, MA, October 12–15
- [33]. Weinke T, Schiller R, Fehrenbach FJ, Pohle HD (1992)Association between *Staphylococcus aureus* nasopharyngeal colonization and septicemia in patients infected with the human immunodeficiency virus. *Eur J Clin Microbiol Infect Dis*;11:985-9.