The Effects Of Casuarina Bark On Lipid Profile And Random Blood Sugar Level In Albino Rats

Aloh Godwin Sunday¹, Obeagu Emmanuel Ifeanyi², Umechukwu Chioma Eucharia³

1. Lecturer, Department of Biochemistry, Michael Okpara University of Agriculture, Umudike, Abia State, Nigeria.
2. Diagnostic Laboratory Unit, University Health Services Department, Michael Okpara University of Agriculture, Umudike, Abia State, Nigeria.
3. Department of Biochemistry, Ebonyi State University, Abakaliki, Nigeria.

Abstract: The effect of Casuarina equisetifolia bark incorporated into rat feed at 10-40% on the lipid profiles and blood sugar of albino rats was investigated. The rats were fed ad lib for 21 days. The parameters studied were triacylglycerol (TGL), total cholesterol (TC), total lipid (TL), phospholipids (PHOS), high-density lipoprotein (HDL) and random blood sugar (RBS). Their means were subjected to a t-Test at P<0.05. There was no significant change (P>0.05) in the TGL levels of all the rats, including the control, as they all range between 0.18-0.22(mg/dl). The effects on TC and TL were irregular as they did not display any dose dependence. The mean plasma PHOS levels did not change significantly (P>0.05) between the control and the rats fed on 10% feed (0.19±0.00 vs 0.18±0.00 mg/dl), was significantly lowered (P<0.05) at 20-40% feed content but did not change significantly (P>0.05) in the rats maintained on the feeds with the 20-40% bark contents. The mean HDL level rose, although insignificantly (P>0.05) with the percentage contents of the feeds increased, indication that it could have anti-diabetic properties.

Keywords: Casuarina equisetifolia bark, Triacylglycerol, Total cholesterol, Total lipid, phospholipid, high density lipoprotein, random blood sugar.

I. Introduction

Casuarina equisetifolia L. (Casuarinaceae) is an evergreen tree to 25m high with drooping branches and needle-like branchlets. The Bark is light greyish-brown, smooth on young trunks, rough thick, furrowed and flaking into oblong pieces on older trees; inner bark reddish or deep dirty brown, astringent (Cambine and Ash, 1994). Common along the beaches, rocky coasts, limestone outcroppings, dry hillside and open forests in both wet and dry zones from sea-level to mid montane. Native to South-East Asia, Australia and Polynesia. It is also cultivated as an ornamental, for wind-breaks, or as a medical plant in some tropical countries in the South Pacific.

The chief chemical constituents include Caffeic acid; chlorogenic acid; d-galactocatechin; ellagic acid; prodelphinidin; propelargonidin; quercetin; rutin. It is also accounted to contain 98% of asparagine and glutamine of the total amino acid in the nodules. The bark contains 10% catchol tannin, the root 15%.

Phytosterol from the leaves of the plant shows antibacterial activity, hypoglycaemic, antifungal, molluscidal, cytotoxic. In Tahiti, the plants are used to treat nervous disorders, diarrhoea and gonorrhoea (Whistler, 1992). Tongans use it to treat cough, ulcers, stomach aches and constipation. Dysuria and menorrhagia are treated with an infusion of the bark. An infusion of the bark, in Tonga is used as an emetic to treat throat infections.

Cardiovascular system refers to the passage through which blood flows - the heart, arteries, veins and capillaries. It is the portion of the circulatory system that includes the heart and blood vessels. It moves the blood between the body cells and organs of the integumentary, digestive, respiratory and urinary systems that communicate with the external environment.

Cardiovascular disease is a class of disease that involves the heart or blood vessels (arteries, capillaries and veins). Cardiovascular disease refers to any disease that affects the cardiovascular system, principally cardiac disease like coronary heart disease. The causes of cardiovascular disease are diverse but atherosclerosis and hypertension are the most common. (Burtis et al., 2008).

Lipid profile is used to diagnose, detect, evaluate and distinguish cardiovascular disease or injuries. Lipid profile is a collective term for tests carried out for total cholesterol, high-density lipoproteins, low-density lipoproteins and triglycerides. An extended lipid profile may include very low-density lipoproteins. This is used to identify hyperlipidemia (various disturbances of cholesterol and triglyceride levels), many forms of which are recognized risk factors for cardiovascular disease and sometimes pancreatitis (Sagara et al., 2008).
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Aims and objectives
The sole aim of this project is to determine the effects of bark of *Casuarina equisetifolia* plant on cardiovascular by monitoring lipid profile and blood glucose level of albino rats treated with it.

II. Materials And Method

**Biological Materials**
Albino rats
*Casuarina equisetifolia* plant (Bark)

**METHODS**

**Collection of samples**

**Collection of plants**
The *Casuarina equisetifolia* plant (Bark) was collected from the people’s club garden Abakalili, Ebonyi State.

**Collection of Albino rats**
The albino rats were purchased from the zoology department of University of Nigeria (UNN). They were fifteen (15) in number, which was later kept in biochemistry department animal house.

**Preparation of plant sample**
The casuarinas bark which has been kept under room temperature for about 2 weeks grinded with a blender into a powder form.

**Animal handling and treatment**
The animals were kept in five cages. Three animals per cage and were labelled properly, control, Group 1 – Group 4. For control animals poultry food and water was given to them, 1, 2, 3 and 4 were given 10%, 20%, 30% and 40% of the compounded feed. The feed was compounded as follows:

- 10% of feed = 900g of poultry food + 100g of powdered casuarina bark.
- 20% of feed = 800g of poultry food + 200g of powdered casuarina bark.
- 30% of feed = 700g of poultry food + 300g of powdered casuarina bark.
- 40% of feed = 600g of poultry food + 400g of powdered casuarina bark.

**Collection of blood samples from the animals**
The blood sample was collected from the heart after being anaesthetized. The blood was dispensed into a sterile specimen bottle and were appropriately labelled and allowed to clot before centrifuged. The supernatant (serum) was collected using micropipette.

**Determination of lipid profile**

**Estimation of total serum cholesterol** (Ochei and Kalhatkar, 2007).

**Principle:** in the presence of excess acid such as phosphoric acid and ferric (Fe$^{3+}$) ions, cholesterol is oxidized to disulphuric acid which is reddish purple in colour (Salkoroske reaction). It is read spectrophotometrically at 560nm (green yellow filter).

**Specimen:** Serum after 12 hours fast.

**Procedure:** Using ferric reagent, cholesterol standard (250mg/dl).

- The serum is diluted with water in the ration of 1: 20 (0.1ml of serum + 1.9ml distilled water)
- Cholesterol standard is diluted with glacial acetic acid in the ratio of 1: 20.
- The three test tubes were shook for 10 seconds to mix the contents properly
- The test tubes were placed immediately in a boiling water baht for exactly 90 seconds.
- They were cooled in a running tap water for 5 minutes.
- Absorbances were read for all the test tubes (mixtures) using spectrophometer at 560nm

Where 250 = concentration of standard

The values were in mg/dl but can be converted to mmol/L by multiplying by 0.0259.

**ESTIMATION OF SERUM HDL CHOLESTEROL** (Ochei and Kalhatkar, 2007).

**Principle:** Chylomicrons, VLDL and LDL are precipitated by phosphotungstic acid in the presence of magnesium ions, leaving HDL in solution, which is read using spectrophometer.

**Procedure:** Using phosphotungstic acid reagent, magnesium chloride solution colour reagent concentrated sulphuric acid, cholesterol standard.
Two test tubes were set up and labelled test and standard.
- 1ml of serum was added to the test using micropipette.
- 0.1ml of phosphotungstic acid reagent was added to both test tubes and mixed well
- 0.05ml magnesium chloride solution was also added immediately and stirred.
- After stirring, the mixture was centrifuged using a centrifuge at 2500rpm for 30 minutes.
- The supernatants were transferred to another test tube using Pasteur pipette, then absorbance was read using spectrophotometer at 560nm.

The values were in mg/dl but can be converted to mmol/L by multiplying by 0.0259.

**Estimation of serum Triacylglycerol (OCHI AND KALHATKAR, 2007)**

**Procedure:** Using colour reagent (mixture of solution A and solution B) and triglyceride standard.

- Three tubes were set up and properly labelled test, standard and blank.
- 3.0ml of colour reagent was added to the three test tubes.
- 0.03ml of serum was added to the test using micropipette while 0.03ml of triglyceride and distilled water was added to the standard and blank respectively.
- The three test tubes were shaken well and the mixture mixed well then allowed/ incubated at 37°C for 15 minutes in a water bath.
- Absorbance was obtained using spectrophotometer at 42nm using blank to zero.

The values were in mg/dl but can be converted to mmol/L by multiplying by 0.0113.

**Estimation of total lipid (Burtis et al., 2008)**

**Procedure:** Using trichloroacetic acid, 0.6 in aqueous solution of ammonium molybdate reagent concentrated perchloric acid, reducing agent.

- 0.2ml of blood serum and 2.8ml of distilled water was added to a sample test tube, 3ml of the distilled water was added to a control test tube.
- 3ml of trichloroacetic acid was added to each test tube and the contents were mixed by shaking.
- The sample solution was allowed centrifuge for 15 minutes at 3000 rpm.
- The supernatant liquid was decanted and placed in the test tube upside down on a piece of filter paper to let the residual liquid flow out.
- 1ml of concentrated perchloric acid was poured into both test tubes and two glass beads placed in each and the contents mixed.
- The test tubes were placed in the sand bath at 180°C for 20-30 minutes to hydrolyze the contents (until the solution decolorize).
- The test tubes are allowed to cool under room temperature and then 3ml of distilled water. 1ml of ammonium molybdate reagent and 2ml of freshly prepared reducing agent was added.
- The contents were mixed thoroughly and allowed to stand for 10 minutes at room temperature.
- The absorbance of the sample solution was measured against control solution on the photocalorimeter at 630nm.
- The total phospholipids concentration x (g/litre) in blood serum is calculated by making use of the formular

\[
\text{Where } m \text{ is the mass of inorganic phosphate in the sample solution mg; } 5000; \text{ is the sealing factor for conversion to per litre blood serum; } 10 \text{ is the sealing factor for milligram to gram conversion. } 25; \text{ is the sealing factor (Lipid phosphorus accounts for 4% of relative molecular mass of phospholipids) the values were in (g/litre) but can be converted to mmol/litre by multiplying by 0.323.}
\]

**Estimation of blood sugar**

**Copper-Reaction Method (Renschler et al., 2005)**

**Procedure:** Using sodium tungstate (10%), 2/3N sulphuric acid, Alkaline copper tartrate reagent, phosphomolybdic acid reagent, stock glucose standard solution (1g/dl), working glucose standard solution (10mg/dl).

- O.1ml of serum and 7.0ml of distilled water are mixed then 1.0ml of 2/3N H₂SO₄ was added and allowed to stand for 5 minutes at room temperature.
- It was filtered using whatman No. 1 filter paper.
- Three test tubes were set up and properly labelled test, blank and standard.
- 1.0ml of the distilled water was added to blank.
- 1.0ml of working standard was added to standard.
- 1.0ml of supernatant was added to the test tube.
- 1.0ml of alkaline copper tartrate was added to the blank, standard and test.
- The test tubes were placed in boiling water bath for 10 minutes.
- 1.0ml of phosphomolybdic acid reagent was added to each test tube and cooled.
- The absorbance was read at 680nm.

**Statistical Analysis**

The results obtained were subjected to statistical analysis using ANOVA

### III. Results

Table 4.1: The results shows the mean lipid and sugar levels of rats after 21 days of feeding with Casuarina compounded feeds.

<table>
<thead>
<tr>
<th>GROUP</th>
<th>TGL (mg/dl)</th>
<th>TC (mg/dl)</th>
<th>TL (mg/dl)</th>
<th>PHOS (mg/dl)</th>
<th>HDL (mg/dl)</th>
<th>RBS (mg/dl)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1. Control</td>
<td>0.218±0.022</td>
<td>0.065±0.015</td>
<td>0.037±0.020</td>
<td>0.192±0.003</td>
<td>0.002±0.002</td>
<td>0.406±0.046</td>
</tr>
<tr>
<td>2. 10% feed</td>
<td>0.215±0.004</td>
<td>0.085±0.017</td>
<td>0.029±0.001</td>
<td>0.180±0.004</td>
<td>0.003±0.001</td>
<td>0.154±0.000</td>
</tr>
<tr>
<td>3. 20% feed</td>
<td>0.219±0.001</td>
<td>0.076±0.019</td>
<td>0.170±0.001</td>
<td>0.010±0.001</td>
<td>0.125±0.001</td>
<td>0.194±0.000</td>
</tr>
<tr>
<td>4. 30% feed</td>
<td>0.183±0.060</td>
<td>0.074±0.003</td>
<td>0.020±0.010</td>
<td>0.169±0.001</td>
<td>0.003±0.001</td>
<td>0.119±0.000</td>
</tr>
<tr>
<td>5. 40% feed</td>
<td>0.222±0.016</td>
<td>0.087±0.009</td>
<td>0.031±0.015</td>
<td>0.170±0.001</td>
<td>0.002±0.000</td>
<td>0.199±0.001</td>
</tr>
</tbody>
</table>

**NOTE:** Means with the same letter are not significantly different

**KEY**

TGI: Triacylglycerol
TC: Total choleaterol
PHOS: Phospholipids
HDL: High Density Lipoproteins
RBS: Random Blood sugar

### IV. Discussion

Cholesterol is an essential substance involved in many cellular functions, including the maintenance of membrane fluidity, production of vitamin D on the surface of the skin, production of hormones and possibly helping cell connections in the brain (Daniels *et al.*, 2009). It is of vital necessity that the body cells should have adequate supply of cholesterol. However, when cholesterol levels rise in the blood, they can deleterious consequences; in particular, cholesterol has generated considerable notoriety for its causative role in atherosclerosis, the leading cause of death in developed countries around the world (Stamler *et al.*, 2000). Great efforts have been put into reducing the risk of cardiovascular diseases through the regulation of cholesterol, traditional plant remedies have been used in the treatment of illness (Akhtar and Ali, 2004) but only few have been evaluated scientifically. Therefore, the effects of the diet preparations of bark of *Casuarina equisetifolia* on the lipid profile of rats were studied and compared.

The analysis of lipid profile reviewed significantly high levels (P<0.005) of total cholesterol, no significant difference in triacylglycerol and total lipid (P>0.005). The biochemical mechanism responsible for this is an interesting subject for further research, since studies made on the ethanolic extract of the plant shows it has anti-hyperlipidaemic effect (Bucholtz *et al.*, 1997)

Phospholipids are majorly components of all cell membranes which are usually found with cholesterol molecules (Choi *et al.*, 2005). There is no significant change in phospholipid level between the control and the rats fed on 10%, but was significantly lowered (P<0.05), at 20-40% feed content following the administration of the compounded *Casuarina equisetifolia* bark is an indication that the plant extract may possess antihyperlipidemic properties and this may help reduce the incidence of cardiovascular diseases like atherosclerosis and hypertension.

Also there was increase in HDL – cholesterol level although insignificant in the *Casuarina equisetifolia* fed rats compared to the control. Although in the past, an increase in the serum total cholesterol level is associated with increased risk of atherosclerosis, however, recent reports indicated that the LDL/HDL ratio is a stronger index of atherogenicity of the lipoproteins rather than the lipid profile of the individual lipoprotein fraction i.e the lower the ratio, the less atherogenic the lipoprotein profile is thought to be.

The significant reduction observed in the blood glucose level (P<0.005) following the administration of the compounded diet is an indication that the plant possess anti-diabetic effect. Earlier studies made on the extract of the plant shows that some chemical constituents of the plant are responsible for its anti-diabetic effect.

### V. Conclusion

Based on the result obtained in this research, consumption of the *Casuarina esquisetifolia* (Bark) could be beneficial to people at high risk of cardiovascular disease. More investigations are required in the actual mechanism of *Casuarina esquisetifolia* (Bark) in the studied parameter.

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Reference


