

Photoactivated Disinfection of Root Canal System with FotoSan

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Abstract: The aim of this *in vitro* study was to determine the antibacterial activity of photoactivated disinfection with FotoSan on an experimental bacterial biofilm in the root canal system consisting of *Candida albicans* and *Enterococcus faecalis*. **Material and methods:** The study was performed on 22 single root extracted teeth. The selected teeth were prepared with rotary nickel-titanium endodontic files and divided in three groups – two test groups (n=20) and one control group (n=2). According to the root canal disinfection method, the test samples (n=20) were randomized in two sub-groups: 1st group - passive irrigation, 2nd group – photoactivated disinfection with FotoSan. **Results:** After microbiological analysis no *Candida albicans* colonies were observed in test samples treated with FotoSan photoactivated disinfection and the percentage of *Enterococcus faecalis* was reduced to 99,9%.

Keywords: FotoSan, photoactivated disinfection, *Candida albicans*, *Enterococcus faecalis*.

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I. Introduction

The main objective of endodontic treatment is to eliminate the pathogenic microorganisms and their byproducts from the root canal system by shaping, cleaning and disinfecting an infected root canal space. However, chemo-mechanical procedures may fail to clean each area of the root canal system¹. The most common microbial species associated with endodontic failure are: *Enterococcus faecalis*, *Streptococci*, *Staphylococci*, *Actinomyces*, *Pseudomonas*, *Candida albicans*, *Prevotella* etc². Nowadays there are new methods such as sonic, ultrasonic irrigation, various lasers and photodynamic therapy, which provide extra disinfection of root canal system^{3, 4, 5, 6}.

Photoactivated disinfection (PAD) has been proposed as an appropriate antimicrobial treatment to maximize root canal disinfection. Light of an appropriate wavelength activates a photosensitizing molecule attached to the bacterial/fungal membrane and eventually leads to production of highly reactive oxygen species able to kill microorganisms⁷.

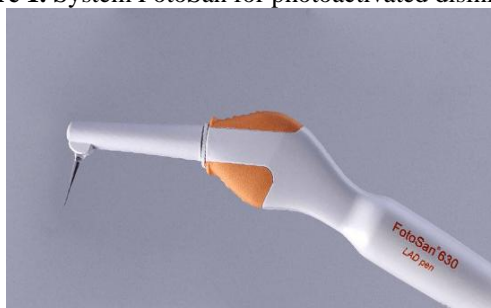
The aim of this *in vitro* study was to determine the antibacterial activity of PAD with FotoSan (CMS Dental, Denmark) on an experimental *Candida albicans* (CA) and *Enterococcus faecalis* (EF) biofilm in the root canal system of single root extracted teeth.

II. Material And Methods

The study was performed on 22 single root freshly extracted teeth. The root canals were prepared with rotary nickel-titanium endodontic files (Pro Taper, Dentsply, Maillefer) to final apical size F3 (30/09). For chemical disinfection during the root canal preparation 2.5% NaOCl was used. After the final irrigation the root canals of all extracted teeth were dried and infiltrated with experimental biofilm containing: 200 mL of LBG growth media, 1 cm³ CA and 1 cm³ EF. Teeth were placed in vials with 5 mL LBG and were incubated at 37°C for 7-10 days. As control group were used 2 teeth (n=2). After the contamination, the control samples weren't subjected to any disinfection. According to the root canal disinfection method, the test samples (n=20) were randomized in two sub-groups: 1st group (n=10): passive irrigation with 2.5% NaOCl for 1 min, distilled water, 17% ethylenediaminetetraacetic acid (EDTA) for 1 min and drying with 95° alcohol and paper points; 2nd group (n=10) - PAD with FotoSan (Fig. 1). The protocol for the second test group included: irrigation with 2.5% NaOCl for 1 min, distilled water and 17% EDTA. After drying with 95° alcohol and paper points, a photo sensitizer (PS) was applied into the root canal (Toluidine Blue) and activated with LED light source (628 nm,

2000mW/cm²) for 1 min (Fig. 2). After the activation the PS was removed from the root canal by irrigation with distilled water and drying with paper points.

Figure 1. System FotoSan for photoactivated disinfection



Dentinal debris for subsequent microbiological analysis was taken from each root canal of the test groups (n=20). The material was collected with sterile H-files and stored in sterile containers.

In the microbiological laboratory it was made ten-fold dilutions from control group. From each dilution a material was taken and seeded into 3 petri dishes with the following media: EVA for EF and Sabouraud dextrose chloramphenicol agar for CA. The samples were thermostated for 24±3 h at 37°C (for the bacteria) or 24-48 h at 32.5±2.5°C (for yeast) until single colonies appeared. The number of grown colonies was determined.

III. Result

After the microbiological analysis, the following numbers of isolated microorganisms were found:

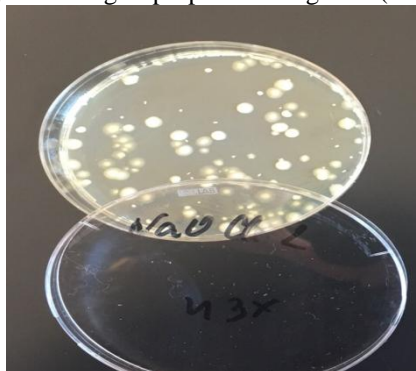
- control group:

CA - 4×10^4 chm/cm³, EF - 6.1×10^5 chm/cm³;

- 1st group (Fig. 2):

petri dish 1: CA - 4 chm/cm³, EF - 28 chm/cm³;
petri dish 2: CA - 30 chm/cm³, EF - 82 chm/cm³;
petri dish 3: CA - <1 chm/cm³, EF - 36 chm/cm³;
petri dish 4: CA - 15 chm/cm³, EF - 56 chm/cm³;
petri dish 5: CA - 47 chm/cm³, EF - 18 chm/cm³;
petri dish 6: CA - 6 chm/cm³, EF - 45 chm/cm³;
petri dish 7: CA - <1 chm/cm³, EF - 37 chm/cm³;
petri dish 8: CA - 11 chm/cm³, EF - 39 chm/cm³;
petri dish 9: CA - <1 chm/cm³, EF - 20 chm/cm³;
petri dish 10: CA - <1 chm/cm³, EF - 74 chm/cm³;

Figure 2. 1st group - passive irrigation (results)

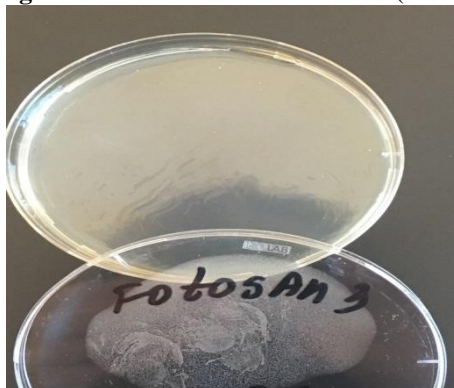


- 2nd group (Fig. 3):

petri dish 1: CA - <1 chm/cm³, EF - 2 chm/cm³;
petri dish 2: CA - <1 chm/cm³, EF - 5 chm/cm³;
petri dish 3: CA - <1 chm/cm³, EF - 6 chm/cm³;
petri dish 4: CA - <1 chm/cm³, EF - 10 chm/cm³;
petri dish 5: CA - <1 chm/cm³, EF - 8 chm/cm³;
petri dish 6: CA - <1 chm/cm³, EF - 15 chm/cm³;

petri dish 7: CA - <1 chm/cm³, EF - 7 chm/cm³;
petri dish 8: CA - <1 chm/cm³, EF - 3 chm/cm³;
petri dish 9: CA - <1 chm/cm³, EF - 20 chm/cm³;
petri dish 10: CA - <1 chm/cm³, EF - 12 chm/cm³;

Figure 3. Photoactivated disinfection (result)



The study found that no CA colonies were observed in FotoSan PAD, and the percentage of EF was reduced to 99,9%.

IV. Discussion

Treatment failure and persistent periapical pathology can result from microbial infections because of inadequate disinfection of the root canals. Root canals function as an incubator since they are closed spaces with low oxygen concentration and therefore they promote the growth of microorganisms. As a result of limited access to different regions in the root canal system, preparing a canal completely devoid of microorganisms in an infected tooth is extremely difficult or even impossible despite proper instrumentation and irrigation. Resistant microorganisms such as EF, gram-negative facultative anaerobic bacilli, CA can have a major role in the persistence of infection in the root canal system⁸.

In the last few years new irrigating solutions or additional techniques for endodontic disinfection have been proposed and tested. A modern device with the aim to minimize or eliminate the bacterial population in the root canal is PAD^{9, 10, 11, 12}. PAD is not only effective against bacteria, but also against other microorganisms including viruses, fungi, and protozoa^{7, 13, 14, 15, 16, 17}. Many studies have shown its efficacy in endodontics. In 2006 Williams et al measured the antibacterial action of PAD on different endodontic bacteria (*Fusobacterium nucleatum*, *Peptostreptococcus micros*, *Prevotella intermedia* and *Streptococcus intermedius*) in planktonic suspension and in root canals, and concluded that PAD kills endodontic bacteria at statistically significant levels¹⁸. Soukos et al investigated the effects of PAD on endodontic pathogens in planktonic phase as well as on EF biofilms in experimentally infected root canals of extracted teeth⁹. In 2010 Schlafer et al tested the antimicrobial effect of FotoSan on endodontic pathogens (*Escherichia coli*, CA, EF, *Fusobacterium nucleatum*, and *Streptococcus intermedius*) in planktonic suspension and after inoculation of *S. intermedius* into prepared root canals of extracted molars. In the test they verified a strong reduction of the number of viable endodontic pathogens both in planktonic suspension and in root canals⁷.

One of the advantages of photosensitization compared to traditional antimicrobials is that as the interaction of highly reactive oxygen with organic molecules is not specific, any macromolecule within microbial cell may become a potential target, thus hindering the development of mechanisms of microbial resistance. Furthermore, the procedure can be repeated several times, as there are no reports of cumulative effects^{19, 20}.

PAD has other attributes that make it an excellent tool in intracanal bacterial reduction, such as: safety for human tissues, ability to eradicate pathogens in biofilms, easy to apply and painless²¹.

V. Conclusion

FotoSan may be considered an adjunctive procedure to kill residual bacteria in the root canal system after standard endodontic treatment.

References

- [1]. Pourhajibagher M, Bahador A. Adjunctive antimicrobial photodynamic therapy to conventional chemo-mechanical debridement of infected root canal systems: A systematic review and meta-analysis. *Photodiagnosis Photodyn Ther* 2019; 26: 19-26.

- [2]. Filipov I, Markova K, Boyadzhieva E. Efficiency of photoactivated disinfection on experimental biofilm – scanning electron microscopy results. J IMAB 2013; 19 (4): 383-6.
- [3]. Halford A, Ohl CD, Azarpazhooh A, Basrani B, Friedman S, Kishen A. Synergistic effect of microbubble emulsion and sonic ultrasonic agitation on endodontic biofilm in vitro. J Endod 2012;38(11):1530-1534.
- [4]. Macedo RG, Verhaagen B, Rivas DF, Versluis M, Wesselink P, van der Sluis L. Cavitation measurement during sonic and ultrasonic activated irrigation. J Endod 2014;40(4):580-583
- [5]. Siddiqui SH, Awan KH, Javed F. Bactericidal efficacy of photodynamic therapy against *Enterococcus faecalis* in infected root canals: a systematic literature review. Photodiagnosis Photodyn Ther 2013;10(4):632-643.
- [6]. Chrepa V, Kotsakis GA, Pagonis TC, Hargreaves KM. The effect of photodynamic therapy in root canal disinfection: a systematic review. J Endod 2014;40(7):891-898.
- [7]. Schlafer S, Vaeth M, Hørsted-Bindslev P and Frandsen EVG. Endodontic photoactivated disinfection using a conventional light source: an in vitro and ex vivo study. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2010; 109: 634-641.
- [8]. Ashraf H, Samiee M, Eslami G and Hosseini MRG. Presence of *Candida Albicans* in Root Canal System of Teeth Requiring Endodontic Retreatment with and without Periapical Lesions Iran Endod J. 2007; 2(1): 24–28.
- [9]. Soukos NS, Chen PSY, Morris JT, et al. Photodynamic therapy for endodontic disinfection. J Endod 2006;32(10):979-984.
- [10]. Jori G. Photodynamic therapy of microbial infections: state of the art and perspectives. J Environ Pathol Toxicol Oncol 2006; 25(1-2):505-519.
- [11]. Konopka K, Goslinski T. Photodynamic therapy in dentistry. J Dent Res 2007; 86(8): 694-707
- [12]. Poggio C, Dagna A, Arciola CR, Florindi F. Photoactivated disinfection (PAD) in endodontics: An in vitro microbiological evaluation. The International journal of artificial organs 2011; 34(9):889-97.
- [13]. Bonsor SJ, Nichol R, Reid TMS, Pearson GJ. Microbiological evaluation of photo-activated disinfection in endodontics (An in vivo study). Br Dent J 2006;200(6):337-341.
- [14]. Gambarini G, Plotino G, Grande NM, et al. In vitro evaluation of the cytotoxicity of FotoSan light activated disinfection on human fibroblasts. Med Sci Monit 2011;17(3):21-25.
- [15]. Kömerik N, Curnow A, MacRobert AJ, Hopper C, Speight PM, Wilson M. Fluorescence bioburden and photosensitising activity of toluidine blue on rat buccal mucosa. Lasers Med Sci 2002;17(2):86-92.
- [16]. Williams JA, Pearson GJ, John Colles M. Antibacterial action of photoactivated disinfection PAD used on endodontic bacteria in planktonic suspension and in artificial and human root canals. J Dent 2006;34(6):363-371.
- [17]. Williams J, Pearson GJ, Colles J, Wilson M. The antibacterial effect of Toluidine Blue O on bacterial colonies in a collagen matrix and carious dentine. Caries Res 2004;38(6):530-536.
- [18]. Williams JA, Pearson GJ, John Colles M. Antibacterial action of photoactivated disinfection PAD used on endodontic bacteria in planktonic suspension and in artificial and human root canals. J Dent 2006;34(6):363-371.
- [19]. Wilson M. Lethal photosensitisation of oral bacteria and its potential application in the photodynamic therapy of oral infections. Photochem Photobiol Sci 2004;3:412-8.
- [20]. Soukos NS, Goodson JM. Photodynamic therapy in the control of oral biofilms. Periodontol 2011;55:143-66.
- [21]. Gursoy H, Ozcakar-Tomruk C, Tanalp J, Yilmaz S. Photodynamic therapy in dentistry: A literature review. Clin Oral Investig 2013;17: 1113–25.

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