

## **Comparative study on different local hydrocolloids on quality of set-type yoghurt made from bovine milk**

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### **Abstract**

*Yoghurt is fermented and coagulated milk product which is often produced with the incorporation of stabilizer to improve the textural quality of the product. This study investigated the effects of cassava (T1), corn (T3) and potatoe (T5) starches as well as gelatin (T4, positive control) as stabilizers on the proximate, physical and antioxidant properties of set-type yoghurt. A stabilizer free yoghurt (T2) to serve as the negative control was also made. Raw milk from Bunaji cow was clarified, homogenized and pasteurized at 82°C for 3 minutes. Sucrose (5%) was then added per litre of milk, thereafter cooled to 42°C for inoculation. The corn, potatoe and cassava flour at 20 g each was dissolved into 200 mls of water and bring to a boil to form a paste. A 20 g paste of each of the stabilizer as well as the gelatin were measured into 1000mls of the milk. Starter culture (5g/L) was added and mixed thoroughly and incubated at 43°C until a coagulum was formed. Results showed that moisture significantly ( $P < 0.001$ ) increased from 67.43 - 79.67 % as the storage period progressed, while the ash, protein, fat and carbohydrate significantly decreased with storage time. Treatments had highly significant ( $P < 0.001$ ) effect on the proximate composition with highest moisture content (74.07%) in the control sample (T2). Protein and carbohydrate were highest (4.48 % and 21.06 %) in cassava starch (T1) and potatoe starch (T5) respectively. The interaction between storage periods and treatments showed highly significant ( $P < 0.0001$ ) effect.*

*Physical property revealed that water holding capacity and whey drainage had the same trend of increase as the storage time increased while syneresis and viscosity showed decrease as storage time progressed. Treatment effect showed that yoghurt with corn starch recorded the highest water holding capacity (97.89 %) and viscosity (84.18 dPas), least syneresis (9.73 %) and whey drainage (0.81%). The interaction between storage periods and treatments showed that the highest water holding capacity (98.67%) was obtained in corn starch while the syneresis (29.20%), whey drainage (3.57%) were superior in cassava starch at day 14. The utmost value for viscosity (89.04 dPa.s) was noted for corn starch at 7 days of storage. The antioxidant indicated that the DPPH had the highest (35.73 %) scavenging potential at day 1 of storage. Treatment effect showed significant ( $P < 0.0001$ ) increase in the control sample (40.76%) while the least value (28.72 %) was obtained in corn starch. Similar trend was observed in the interaction effect as the control sample at day 1 of storage had the highest (41.60 %) DPPH value. Conclusively, corn starch can be used as natural hydrocolloids in reduction of syneresis, improved viscosity and higher concentration of antioxidants in the yoghurt samples as it competed favourably with gelatin, a conventional synthetic stabiliser.*

**Key words:** *Yoghurt, stabilizers, physical, proximate, antioxidant, synthetic*

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Date of Submission: 03-02-2023

Date of Acceptance: 15-02-2023

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### **I. Introduction**

Milk is the origin of all dairy products. Due to its high nutrient concentration, it easily undergoes spoilage, hence can be processed into other products like yoghurt in order to retain its basic nutrients [1]. Yogurt and its related products are popular worldwide. [2]. The major concern in the yoghurt industry is the production and maintenance of a product with optimum consistency and stability [3]. The highest production or consumption of yoghurt is in Mediterranean, Asian countries and in Central Europe [4]. The consumption of functional food products is currently on the rise [5]. Yoghurt is a fermented milk product, which is produced by fermenting milk with lactic acid bacteria, which are responsible for development of typical yoghurt flavor [6]. It is highly nutritious, cherished and easily digestible due to the predigested nutrients by the starter culture. The texture of yogurt is as important as its taste and flavor in terms of consumer preferences [7].

Stabilizers, also called thickeners, gelling agents or hydrocolloids, can be obtained from different sources including animal connective tissues, sea and land plants and microorganisms [8]. They have gelling, thickener and stabilizer properties [9;6]. Stabilizers are commonly used in cultured products to control texture and reduce whey separation since they impart good resistance to syneresis and a smooth sensation in the mouth

by binding water to reduce water flow in the food matrix space [10]. Sodium caseinate and gelatin increase the density of the protein network in the gel microstructure [11;12]. Some may interact with protein in the food matrix and hence further increase hydration behaviour. The properties of the milk used in yoghurt production, the production and storage conditions or the transportation to far sales points can lead to textural defects such as viscosity variations and syneresis [13;14]. Various stabilizers are used to prevent these problems and to create desirable textural characteristics [15;16]. Stabilizers, also called thickeners, gelling agents or hydrocolloids, can be obtained from different sources including animal connective tissues, sea and land plants and microorganisms [8]. They have gelling, thickener and stabilizer properties [9;6]. Type of hydrocolloids used in yoghurt production could affect the stability and functional properties of the product. Synthetic hydrocolloids have reportedly been used in yoghurt production [17;18; 14], however, in recent years there is an increasingly negative consumers' perception on their use.

Cornstarch is a carbohydrate extracted from the endosperm of a maize grain. The increased concentration of starch enhanced the consistency index, and reduce the emulsion droplet size, thereby improving the emulsion stability [19]. Also, potato starch is useful in yoghurt production, because it serves as a binder to reduce defects and cracks on the surface curd by making textures of manufactured yoghurt appealing [20]. Cassava flour has a high hydrocolloid property which makes it a good binder when used in yoghurts, creams and desserts [21]. The thrust of this study was to investigate the effects of these different local hydrocolloids (cornstarch, potato starch and cassava starch) on some characteristics of set-type yogurt made from cow's milk.

## **II. Materials And Method**

### **Experimental site**

The study was carried out in the Nutrition and Microbiology Laboratories of the Department of Animal Production and Health, Federal University of Technology, Akure, (FUTA), Ondo State. Akure is located on latitude 7.491780 °N and Longitude 4.944055 °E and 5.82864 °E with the annual rainfall ranging between 1,300mm and 1650mm average maximum and minimum daily temperature of 38 °C and 27 °C respectively [22].

### **Experimental Materials**

Fresh milk from lactating White Fulani cows was obtained from the Fulani herdsmen at Ipinsa, Akure, Ondo State. Sucrose was purchased from reputable store in Akure while the starter culture, gelatin and corn starch were purchased from Lagos. The potato, cassava and corn starch were hygienically produced.

### **Preparation of Local Hydrocolloids from Potatoes and Cassava tubers**

Sweet potato and cassava tubers were obtained from the market and cassava processing centre respectively. The sweet potato (white variety) was washed, peeled, sliced and sundried. The dried potato chips were milled, sieved using a filter to obtain the powder and stored in an airtight polythene bag. The cassava tuber was peeled, washed and soaked for three days to reduce the antinutrients (Hydrocyanide) and thereafter, sundried. The dried cassava was milled and sieved using a filter to obtain cassava flour and the product was packaged in an airtight polythene bag.

### **Production of yoghurt**

Fifteen liters of Fresh cow's milk was clarified, homogenized and pasteurized at 82°C for 3 minutes. Sucrose (5%) was then added as sweetener per liter of milk. The milk was thereafter cooled to 42°C for inoculation. The corn, potato, and cassava starch at 20g each was dissolved into 200 mls of water and brought to a boil to form a paste. A 20g paste of each of the stabilizer was measured into 1000mls of the milk and thoroughly mixed. Thereafter, 5g of commercially freeze-dried starter culture was added to the mixture and stirred. A 20ml of reconstituted banana flavour was added to 1 litre each of the inoculated milk. Gelatin served as positive control while the sample with no stabilizer served as negative control. Immediately, the yoghurt was kept in an incubator (Gallenkamp cooled incubator model) at 43°C until a coagulum was formed at 14hrs. The yoghurt was refrigerated at 4°C for 1, 7 and 14 days for further analysis.

### **Laboratory analyses**

#### **Proximate composition analysis**

The proximate constituents were determined according to the method of [23].

#### **Determination of viscosity**

The viscosity of the yogurt sample at 5°C was measured using a rotational viscometer (Fungilab, ALPHA H, Spain) at the speed of 100 rpm at 30 second with spindle 7 as P. The samples were analyzed by a texture profile analyzer using TA4/1000 probe [24].

#### **Whey drainage**

Whey drainage was removed from the Yogurt, using a syringe within 24h after the yoghurt fermentation was completed. The relative amount of whey drained off (in mL per 100ml of initial sample) was calculated as the whey drainage [25].

### Water Holding capacity

A 10-g sample was centrifuged at 3,000 rpm for 60 min at 10<sup>0</sup>C. The supernatant was removed within 10 min and the wet of the pellet was recorded . the water holding capacity was expressed as percentage of pellet weight relative to the original weight of yoghurt [26]

### Syneresis

An amount of 20g of the yoghurt was spread in a thin layer to cover the surface of the filter paper. The yoghurt was filtered under vacuum for 10mins. The liquid that passed through the filter paper was collected and recorded. The Percentage of Syneresis (PS) was calculated as the weight of the liquid divided by the weight of the initial sample multiplied by 100 [27].

### Antioxidant activity

The antioxidant activity of the different yoghurt samples was measured using the DPPH method. The free radical scavenging ability of the yoghurt against DPPH (1, 1- diphenyl-2-picrylhydrazyl) was determined using the method described by [28]. 1 mL of the yoghurt sample was mixed with 1 mL of the 0.4 mM methanolic solution of the DPPH. The mixture was left in the dark for 30 minutes before measuring the absorbance at 516 nm. The scavenging activity percentage was determined thus: DPPH Scavenged (%) = A Control – A test / A Control × 100.

### Experimental design and statistical analysis

The experimental design was completely randomized design in a 5×3factorial arrangement. Data obtained were subjected to two-way analysis of variance and significant means were separated using Duncan’s multiple range tests using the [29] version 9.2 software. Where there was significant difference, Duncan’s multiple range test of the same statistical package was used to separate the means.

## III. Results

**Table1: Proximate Composition (%) of Yoghurt at different periods of storage**

Parameters	Moisture	Ash	Fat	Protein	Carbohydrate
<b>Storage Periods (SP)</b>					
1	67.43±0.26 <sup>c</sup>	1.02±0.04 <sup>a</sup>	2.68±0.12 <sup>a</sup>	5.29±0.09 <sup>a</sup>	22.82±0.39 <sup>a</sup>
7	69.53±0.28 <sup>b</sup>	0.88±0.06 <sup>b</sup>	2.13±0.07 <sup>b</sup>	4.64±0.10 <sup>b</sup>	22.46±0.35 <sup>a</sup>
14	79.67±0.44 <sup>a</sup>	0.84±0.06 <sup>b</sup>	1.93±0.15 <sup>c</sup>	3.08±0.15 <sup>c</sup>	12.43±1.15 <sup>b</sup>
	0.0001	0.0001	0.0001	0.0001	0.0001
<b>Treatments (T)</b>					
T1-Cassava starch	71.03±1.92 <sup>d</sup>	0.98±0.02 <sup>b</sup>	2.16±0.07 <sup>bc</sup>	4.48±0.24 <sup>a</sup>	20.71±1.65 <sup>ab</sup>
T2-Control	74.07±2.15 <sup>a</sup>	0.98±0.02 <sup>b</sup>	2.16±0.07 <sup>bc</sup>	4.19±0.54 <sup>b</sup>	15.38±2.83 <sup>d</sup>
T3-Corn starch	71.69±1.71 <sup>c</sup>	0.58±0.07 <sup>d</sup>	2.59±0.17 <sup>a</sup>	4.38±0.26 <sup>ab</sup>	20.18±1.44 <sup>b</sup>
T4-Gelatin	72.96±1.85 <sup>b</sup>	1.15±0.05 <sup>a</sup>	2.41±0.18 <sup>ab</sup>	4.27±0.37 <sup>ab</sup>	18.85±0.46 <sup>c</sup>
T5-Potato starch	71.30±1.83 <sup>d</sup>	0.87±0.02 <sup>c</sup>	1.91±0.28 <sup>c</sup>	4.34±0.31 <sup>ab</sup>	21.06±1.23 <sup>a</sup>
P - value	0.0001	0.0001	0.0001	0.0001	0.0001
<b>SP*T</b>					
1*T1	66.34±0.19 <sup>i</sup>	1.04±0.02 <sup>c</sup>	2.35±0.17 <sup>c</sup>	5.21±0.11 <sup>c</sup>	24.40±0.22 <sup>a</sup>
7*T1	68.11±0.06 <sup>g</sup>	0.96±0.03 <sup>d</sup>	2.05±0.03 <sup>g</sup>	4.58±0.29 <sup>b</sup>	23.60±0.30 <sup>b</sup>
14*T1	78.63±0.32 <sup>c</sup>	0.94±0.02 <sup>d</sup>	2.10±0.05 <sup>f</sup>	3.66±0.08 <sup>j</sup>	14.13±0.07 <sup>g</sup>
1*T2	68.47±0.24 <sup>g</sup>	1.04±0.02 <sup>c</sup>	2.35±0.17 <sup>c</sup>	5.51±0.26 <sup>b</sup>	21.64±0.32 <sup>d</sup>
7*T2	71.22±0.12 <sup>d</sup>	0.96±0.03 <sup>d</sup>	2.05±0.03 <sup>g</sup>	4.98±0.01 <sup>f</sup>	20.43±0.22 <sup>e</sup>
14*T2	82.53±0.27 <sup>a</sup>	0.94±0.02 <sup>c</sup>	2.10±0.05 <sup>f</sup>	2.10±0.05 <sup>n</sup>	4.08±0.04 <sup>i</sup>
1*T3	67.49±0.25 <sup>h</sup>	0.84±0.02 <sup>e</sup>	3.12±0.06 <sup>a</sup>	5.15±0.08 <sup>d</sup>	22.67±0.33 <sup>c</sup>
7*T3	69.13±0.08 <sup>f</sup>	0.46±0.03 <sup>g</sup>	2.30±0.25 <sup>d</sup>	4.55±0.28 <sup>i</sup>	23.30±1.00 <sup>b</sup>
14*T3	78.45±0.23 <sup>c</sup>	0.43±0.02 <sup>g</sup>	2.36±0.23 <sup>c</sup>	3.45±0.02 <sup>k</sup>	14.58±0.31 <sup>g</sup>
1*T4	68.45±0.23 <sup>g</sup>	1.24±0.12 <sup>a</sup>	3.11±0.05 <sup>a</sup>	5.54±0.27 <sup>a</sup>	20.99±0.05 <sup>e</sup>
7*T4	70.14±0.07 <sup>e</sup>	1.13±0.06 <sup>b</sup>	1.94±0.03 <sup>h</sup>	4.20±0.10 <sup>b</sup>	22.50±0.32 <sup>c</sup>
14*T4	80.29±0.15 <sup>b</sup>	1.07±0.03 <sup>c</sup>	2.17±0.11 <sup>e</sup>	3.05±0.03 <sup>m</sup>	13.08±0.10 <sup>h</sup>
1*T5	66.41±0.20 <sup>i</sup>	0.94±0.02 <sup>d</sup>	2.50±0.31 <sup>b</sup>	5.02±0.01 <sup>e</sup>	24.43±0.29 <sup>a</sup>

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7*T5	69.07±0.04 <sup>f</sup>	0.87±0.03 <sup>e</sup>	2.33±0.23 <sup>c</sup>	4.87±0.06 <sup>g</sup>	22.46±0.28 <sup>c</sup>
14*T5	78.43±0.22 <sup>c</sup>	0.81±0.01 <sup>f</sup>	0.90±0.02 <sup>i</sup>	3.12±0.06 <sup>l</sup>	16.30±0.15 <sup>f</sup>
Pv	0.0001	0.0001	0.0001	0.0001	0.0001

Means along the same column with different superscripts are significantly (p<0.05) different. T1= Cassava starch T2= Control, T3= Corn Starch, T4= Gelatin, T5=Potato starch

Presented in Table1 is the proximate compositions of locally stabilized yoghurt stored for 14 days. Moisture significantly (P<0.001) increased from 67.43 - 79.67 % as the storage period progressed, while the ash, protein, fat and carbohydrate significantly decreased with storage time. Treatments had highly significant (P<0.001) effect on the variables investigated. Highest moisture content (74.07%) was observed in the control sample while the least value (71.03%) was recorded in T1 (cassava starch yoghurt). Ash content was superior in T4 (1.15 %), fat was utmost in T3 (2.59 %) while protein and carbohydrate were highest (4.48 % and 21.06 %) in T1 and T5 respectively. The interaction between storage periods and treatments showed highly significant effect (P<0.0001) as moisture (82.53%) content was highest in T2 at day 14. Ash and protein were at their peak (1.24 and 5.54% respectively) in treatment 4 at day 1, while fat (3.12%) and carbohydrate (24.43%) were superior in T3 at day 1 and in treatment 5 at day 1 respectively.

**Table 2: Physical Properties of Yoghurt at different periods of storage**

Parameters	Water Capacity (%)	Holding	Syneresis (%)	Whey Drainage (%)	Viscosity (dPa.s)
<b>Storage period (SP)</b>			80.21±0.86 <sup>c</sup>		
1	79.87±4.27 <sup>c</sup>		19.86±1.85 <sup>a</sup>	1.05±0.22 <sup>c</sup>	80.24±1.14 <sup>b</sup>
7	84.27±4.13 <sup>b</sup>		14.37±1.44 <sup>b</sup>	1.25±0.34 <sup>b</sup>	83.88±1.18 <sup>a</sup>
14	87.73±3.13 <sup>a</sup>		11.97±2.13 <sup>c</sup>	2.85±0.14 <sup>a</sup>	76.58±0.94 <sup>c</sup>
	0.0001		0.0001	0.0001	0.0001
<b>Treatments (T)</b>					
T1- Cassava starch	69.01±3.60 <sup>d</sup>		16.69±2.50 <sup>b</sup>	2.38±0.28 <sup>b</sup>	79.20±1.84 <sup>d</sup>
T2- Control	66.56±2.07 <sup>e</sup>		21.51±2.10 <sup>a</sup>	3.08±0.21 <sup>a</sup>	72.65±0.62 <sup>e</sup>
T3- -Corn starch	97.89±0.26 <sup>a</sup>		9.73±1.69 <sup>d</sup>	0.81±0.34 <sup>e</sup>	84.18±1.27 <sup>a</sup>
T4- -Gelatin	96.56±0.47 <sup>b</sup>		14.52±3.18 <sup>c</sup>	0.97±0.39 <sup>d</sup>	73.75±0.60 <sup>a</sup>
T5- Potato starch	89.78±2.61 <sup>c</sup>		14.53±1.82 <sup>c</sup>	1.34±0.31 <sup>c</sup>	83.82±1.01 <sup>b</sup>
	0.0001		0.0001	0.0001	0.0001
<b>SP*T</b>					
1*T1	54.67±0.33 <sup>j</sup>		14.77±0.39 <sup>g</sup>	2.33±0.03 <sup>c</sup>	72.07±0.07 <sup>i</sup>
7*T1	75.33±0.33 <sup>g</sup>		20.57±0.30 <sup>e</sup>	3.33±0.33 <sup>a</sup>	76.07±0.07 <sup>g</sup>
14*T1	77.00±0.58 <sup>f</sup>		29.20±0.20 <sup>a</sup>	3.57±0.03 <sup>a</sup>	73.11±0.11 <sup>b</sup>
1*T2	95.67±0.33 <sup>c</sup>		27.19±0.19 <sup>b</sup>	0.23±0.03 <sup>b</sup>	80.41±0.03 <sup>f</sup>
7*T2	98.33±0.33 <sup>a</sup>		9.31±0.31 <sup>k</sup>	0.13±0.03 <sup>i</sup>	83.08±0.08 <sup>d</sup>
14*T2	95.67±0.33 <sup>c</sup>		7.07±0.07 <sup>l</sup>	2.53±0.03 <sup>b</sup>	77.14±0.14 <sup>g</sup>
1*T3	97.67±0.33 <sup>b</sup>		26.6±0.31 <sup>c</sup>	0.13±0.03 <sup>i</sup>	83.04±0.04 <sup>d</sup>
7*T3	98.67±0.33 <sup>a</sup>		12.77±0.39 <sup>i</sup>	0.13±0.03 <sup>i</sup>	89.04±0.04 <sup>a</sup>
14*T3	97.33±0.33 <sup>b</sup>		10.71±0.29 <sup>j</sup>	2.17±0.03 <sup>d</sup>	80.43±0.29 <sup>f</sup>
1*T4	71.00±0.58 <sup>h</sup>		9.31±0.31 <sup>k</sup>	1.57±0.03 <sup>e</sup>	81.59±0.30 <sup>e</sup>
7*T4	58.33±0.33 <sup>i</sup>		4.11±0.11 <sup>m</sup>	2.13±0.03 <sup>d</sup>	84.04±0.04 <sup>c</sup>
14*T4	70.33±0.33 <sup>h</sup>		15.77±0.40 <sup>f</sup>	3.43±0.13 <sup>a</sup>	71.10±0.02 <sup>j</sup>
1*T5	80.33±0.33 <sup>e</sup>		21.44±0.29 <sup>d</sup>	1.00±0.00 <sup>f</sup>	84.11±0.11 <sup>c</sup>
7*T5	90.66±0.33 <sup>d</sup>		13.08±0.08 <sup>h</sup>	0.50±0.00 <sup>g</sup>	87.15±0.15 <sup>b</sup>
14*T5	98.33±0.33 <sup>a</sup>		9.08±0.07 <sup>k</sup>	2.53±0.03 <sup>b</sup>	80.21±0.21 <sup>f</sup>
PV	0.0001		0.0001	0.0001	0.0001

Means along the same column with different superscripts are significantly (p<0.05) different. T1= Cassava starch T2= Control, T3= Corn Starch, T4= Gelatin, T5=Potato starch

Presented in Table 2 is the physical property of yoghurt stabilized with different local hydrocolloids at 1, 7 and 14 days storage periods. Storage period had highly significant (P<0.0001) effect on the parameters. Water holding capacity and whey drainage had the same trend of increase as the storage time increased while syneresis and viscosity showed decrease as storage time progressed. The highest water holding capacity and whey drainage were 87.73 and 2.85 % respectively at day 14 while syneresis and viscosity had the least values of 11.97 and 76.58 dPa.s (%) at day 14. Treatment effect showed that yoghurt with corn starch recorded the highest

water holding capacity value of 97.89%, least syneresis (9.73 %), whey drainage (0.81%) and superior viscosity (84.18dPa.s). The interaction between storage periods and treatments showed that the highest water holding capacity (98.67%) was obtained in T3 while the syneresis (29.20%), whey drainage (3.57%) were superior in T1 at day 14. The utmost value for viscosity (89.04 dPa.s) was noted for T3 at 7 days of storage.

**Table 3: Antioxidant Properties (%) of Yoghurt at different periods of storage**

Parameters DPPH	
<b>Storage Periods (SP)</b>	
1	35.73±1.11 <sup>a</sup>
7	33.06±1.25 <sup>b</sup>
14	32.77±1.25 <sup>c</sup>
	0.0001
<b>Treatments (T)</b>	
T1-Control	40.76±0.25 <sup>a</sup>
T2-Cassava starch	34.92±0.30 <sup>c</sup>
T3-Corn Starch	28.72±0.34 <sup>e</sup>
T4-Gelatin	29.28±0.78 <sup>d</sup>
T5-Potato starch	35.61±0.76 <sup>b</sup>
PV	0.0001
<b>SP*T</b>	
1*T1	41.60±0.30 <sup>a</sup>
7*T1	40.47±0.24 <sup>b</sup>
14*T1	40.20±0.20 <sup>b</sup>
1*T2	36.03±0.03 <sup>d</sup>
7*T2	34.70±0.15 <sup>e</sup>
14*T2	34.04±0.04 <sup>e</sup>
1*T3	30.07±0.07 <sup>g</sup>
7*T3	28.05±0.05 <sup>h</sup>
14*T3	28.02±0.02 <sup>h</sup>
1*T4	32.34±0.33 <sup>f</sup>
7*T4	27.97±0.03 <sup>i</sup>
14*T4	27.52±0.29 <sup>i</sup>
1*T5	38.61± 0.31 <sup>c</sup>
7*T5	34.11±0.11 <sup>e</sup>
14* T5	34.10±0.10 <sup>e</sup>
PV	0.0001

The antioxidant property (Table 3) of yoghurt stabilized with different local hydrocolloids at 1, 7 and 14 days storage periods revealed the highest (35.73 %) scavenging potential at day 1 of storage. However, significant reduction was observed in the antioxidant capabilities from 35.73 -32.77 % as storage period increased. Treatment effect showed significant (P<0.0001) increase in the control sample (40.76%) while the least value (28.72 %) was obtained in corn starch. Similar trend was observed in the interaction effect as the control sample at day 1 of storage had the highest (41.60 %) value.

#### IV. Discussion

##### Proximate Composition

The moisture content is used to assess the storability of a product and provides a measure of the water content [30]. The increases in moisture content as storage period increased may be due to the gain of moisture or water from the internal atmosphere of the refrigerator during storage period. [31] reported increase in moisture content due to long period of storage. The moisture content in this study is similar to the findings of [32] who reported values ranging from 71.170- 87.36 % in yoghurt treated with local stabilizers. However, the decline in carbohydrate could be due to the conversion of the carbohydrate (lactose) to lactic acid. This corroborates the reports of [33]. [31] also reported decrease in carbohydrate as storage period increases in carrot and pineapple flavoured yoghurt production. The decline in fat content as storage progressed, corroborated the report of [34]; these authors reported decrease in fat (from 2.50-2.92%) as storage period increases in papaya yoghurt and cactus pear flavoured yoghurt production. The protein content decreases as the storage period progressed. This result disagreed with the report of [35] who reported that during fermentation process, the *Lactobacillus bulgaricus* and *Streptococcus thermophiles* microbe biomass were increased, thus the sum of microbe protein was increased, that automatically led to increased protein in their yoghurt. The reduction in protein content observed in this study was probably due to activity of proteolytic agents on protein degradation as opined by [36]. The highest concentration of ash noted in the gelatin fortified yoghurt could be connected to the mineral content of gelatin which reflected in the product.

### **Physical Properties**

The increase in water holding capacity and whey drainage obtained in this study during storage agrees with the findings of [37] who reported an increase in water holding capacity and whey drainage as storage period increases. The current findings disagreed with the results of [38] and [39] Lubna *et al.* (2020) who reported that water holding capacity values of yoghurt samples decreased due to increase in syneresis during increased storage time. The highest water holding capacity and least syneresis observed in gelatin fortified yoghurt, a conventional stabilizer could as a result of the proteinous nature of gelatin. Proteins have water binding properties and reduce syneresis by increasing the water holding capacity of the yoghurt [40].

Syneresis is considered as a very important physical test for yoghurt quality and is related to the instability of the yoghurt gel network and the impossibility of trapping the serum phase in its gel network [41]. Syneresis in yoghurt occurs due to compression of three-dimensional structure of the protein network that results in decreasing the protein binding power and exiting the water from yoghurt. Adding the hydrocolloids reduced syneresis of the yoghurt because the hydrocolloids are able to establish the stronger bonds with free water molecules due to high molecular weight.

This increase in viscosity during storage from day 1 to 14 may be due to changes in protein-protein binding in a three-dimensional protein network of yoghurt and their rearrangement [4]. Also, [4] opined that the increase in starch concentration resulted in an increase of viscosity and elastic modulus ( $G'$ ) due to the uptake of water by swollen starch granules, resulting in the thickening of the continuous phase, and the formation of more particle-particle interactions. [42] reported an increase in the apparent viscosity of concentrated yoghurt during storage and opined that it could be due to the development of gel structure during storage [42; 43]. [44] also reported that the viscosity of the fruit-flavored yogurt (by adding cornelian cherry paste and sugar at different ratios) increase rapidly up to day 7, and continued to increase slowly up to day 14 of storage and afterward decreased slowly.

### **Antioxidant Properties**

Antioxidants are considered important nutraceuticals [45]. They scavenge free radicals generated in the body due to metabolic processes [46]. Antioxidant compounds, particularly those in the flavonoid family, are required to combat free radicals and prevent oxidative stress, leading to degenerative diseases [47]. The use of DPPH as an assay method is due to the good stability, simplicity and feasibility and the ability to form stable radicals [48]. Highest DPPH inhibition during first day of storage may be attributed to the metabolically active yoghurt bacteria even at low temperature [49]. Continued microbial growth during refrigerated storage may alter some of the phenolic compounds and hence their antioxidant activities [50]. Antioxidant activities during refrigerated storage of yoghurt is attributed to increasing degradation of phenolic compounds with antioxidant activities [51] and/or increasing milk protein polyphenol interaction [52]. In this regard, the consumption of yoghurt is highly advisable within 7 days after yoghurt-making to benefit from high live bacterial contents [53] and high antioxidant activities useful for protective cardiovascular effect [54].

This result is in accordance with the report of [55], when they studied antioxidant, some flavor components, microbiological and microstructure characteristics of corn milk yoghurt. It was also reported that the antioxidant activities of yoghurts containing both yoghurt culture and probiotic culture, determined by both DPPH and ABTS methods, generally increased during the fermentation period [56]. It has been reported that the unstable changes in ABTS, FRAP and DPPH radical scavenging activity are due to many factors, such as the activity of the microbiota and the antioxidant abilities of the many compounds formed during the fermentation process and because phenolic compounds also play an important role in antioxidant activity [57]. Hydrolysis and release of cell wall components through fermentation causes the release of phenolic compounds from food, which in turn affects antioxidant activity [58]. Higher DPPH value indicates low concentration of antioxidants in the sample. The lower values of DPPH obtained instabilized yoghurts compared with the control indicates that stabilized yoghurt samples have higher concentration of antioxidants.

## **V. Conclusion**

The results obtained from this study indicate that corn, cassava, potatoe starch made into paste can serve as stabilizer as they competed favourably with the conventional synthetic stabilizer (gelatin) in improving the textural qualities of the yoghurt. However, among the local starches used, corn starch proved to have better impact on the yoghurt quality.

## **References**

- [1]. Ibhaze, G. A., Akinbanjo, D.T., Jacob, G.T. (2021). Evaluation of Set Yoghurt Quality Enhanced with Selected Indigenous Fruits. *International Journal of Trend in Scientific Research and Development* 5 (6) : 690-699
- [2]. Wang, Y Li, D., Chitrakar, B., Zhang, X., Zhang, N., Liu, C., Li, Y., Wang, M., Tian, H., and Li, C. (2023). Copper inhibits postacidification of yogurt and affects its flavor: A study based on the Cop operon. *Journal of Dairy Science* Vol. 106 No. 2, 2023 (Article In Press) <https://doi.org/10.3168/jds.2022-22369>

- [3]. Olorunnisomo, O.A., Ososanya, T.O., and Adedeji, A.Y. (2015). Influence of Stabilizers on Composition, Sensory Properties and Microbial Load of Yoghurt Made from Zebu Milk. *International Journal of Dairy Science*, 10: 243-248.
- [4]. Sahan N., Yasar K., Hayaloglu A. 2008. Physical, chemical and flavour quality of non-fat yoghurt as affected by a  $\beta$ -glucan hydrocolloidal composite during storage. *Food Hydrocolloid*; **22**: 1291-1297. DOI: <https://doi.org/10.1016/j.foodhyd.2007.06.010>
- [5]. Indiarto, R., Reni, R., Lara, G., Edy Subroto, U., Pangawikan, A and Djali, M. (2023). The physicochemical, antioxidant, and sensory properties of chocolate biscuits incorporated with encapsulated mangosteen (*Garcinia mangostana* L.) peel extract. *International Journal of Food Properties* Vol. 26(1):122–138 <https://doi.org/10.1080/10942912.2022.2159429>
- [6]. Tamime AY, Robinson RK (2007). *Yogurt science and technology*. Boca Raton Boston New York Washington, DC. 35-41.
- [7]. Macit, E and Bakirci, I. (2017) Effect of different stabilizers on quality characteristics of the set-type yoghurt. *African Journal of Biotechnology* 16(46), pp. 2142-2151. DOI: 10.5897/AJB2017.16197
- [8]. Imeson A (2010). *Food stabilisers, thickeners and gelling agents*. Blackwell Publishing Ltd, UK.
- [9]. Lal, S.N.D, O'Connor CJ, Eyres L (2006). Application of emulsifiers/stabilizers in dairy products of high Rheology. *Adv. Colloid Interf. Sci.* 123-126:433-437.
- [10]. Amatayakul T, Sherkat F, Shah NP (2006a). Syneresis in set yoghurt as affected by starter cultures and levels of solids. *Intl. J. Dairy Technol.* 59(3):216-221.
- [11]. Amatayakul T, Sherkat F, Shah NP (2006b). Physical characteristics of set yoghurt made with altered casein to whey protein ratios and EPS-producing starter cultures at 9 and 14% total solids. 20:314-324. *Food Hydrocolloids*.
- [12]. Supavitpatana P, Wirjantoro TI, Apichartsrangkoon A, Raviyan P (2008). Addition of gelatin enhanced gelation of corn-milk yogurt. *Food Chem.* 106(1):211-216.
- [13]. Trachoo, N. (2002). Yogurt: The fermented milk. *Songklanakarin J. Sci. Technol.* 24(4):727-734.
- [14]. Hematyar N, Samarin AM, Poorazarang H, Elhamirad AH (2012). Effect of gums on yogurt characteristics. *World Appl. Sci. J.* 20:661-665. <http://dx.doi.org/10.1080/10408398.2014.903384>
- [15]. Athar IH, Shah MA, Khan UN (2000). Effect of various stabilizers on whey separation (syneresis) and quality of yogurt. *Pak. J. Biol. Sci.* 3:1336-1338.
- [16]. Mohammadifar, A.M., Musavi, S.M., Williams, P.A. (2007). Study of complex coacervation between  $\beta$ -lactoglobulin and tragacanthin (soluble part of gum tragacanth). *Milchwissenschaft.* 62:365-488. of the set-type yogurt. *African Journal of Biotechnology* Vol. 16(46), pp. 2142-2151, DOI: 10.5897/AJB2017.16197
- [17]. Ares G, Gonçalvez D, Pérez C, Reolón G, Segura N, Lema P, Gámbaro A (2007). Influence of gelatin and starch on the instrumental and sensory texture of stirred yogurt. *Int. J. Dairy Technol.* 60:263-269
- [18]. Soukoulis C, Panagiotidis P, Koureli R, Tzia C (2007). Industrial yogurt manufacture: Monitoring of fermentation process and improvement of final product quality. *J Dairy Sci.* 90(6):2641- 2654.
- [19]. Ihekoronye, A.I., Ngoddy, P.O. (1985). *Integrated Food Science and Technology for the Tropics*. Macmillan Publishers Limited, London. pp. 106-112.
- [20]. Syed, T. M., Tariq, M. I., Talat, M. I. and Maqsood, S. (2008). Effect of different additives from local source on the quality of yoghurt. *Pakistan Journal of Nutrition*, 7(5):695-699
- [21]. Melesa, T. (2015). Effect of application of stabilizers on gelatin and syneresis in yoghurt. *Food Science Quality Management*, 37(4):66-74
- [22]. Daniel, O. A. (2015). Urban extreme weather a challenge for a healthy living environment in Akure, Ondo State Nigeria climate, 3(4): 775-791.
- [23]. AOAC (2012). *Official Methods of Analysis*. Association of Official Analytical Chemists. (18th Edition). Gaithersburg, USA.
- [24]. Fox, P. F., Wallace, J. M., Morgan, S., Lynch, C. M., Niland, E. J. and Tobin, J. (2017). *Fundamental of Cheese Science*. Science daily, (4): 45-55.
- [25]. Onwuka, G. I. (2005). *Food analysis and instrumentation, Theory and Practice*. Naphthali Prints, Lagos. pp. 119-121.
- [26]. Parnell- Clunies E.M., Kakuda, Y., Mullen , K., Arnott, D.R., and deMan, J. M. (1986). Physical properties of yoghurt: A comparison of vat versus continuous heating systems of milk. *J. Dairy Sci.*; 69, 2593-2603.
- [27]. Wu, H., Hulbert, G. J. and Mount, J. R. (2001). "Effects of Ultrasound on milk homogenization and fermentation with Yoghurt starter. *innovative Food Science and emerging Technologies* 2:211-218.
- [28]. Gyamfi, M. A., Yonamine, M., & Aniya, Y. (1999). Free-radicalscavenging action of medicinal herbs from Ghana. *General Pharmacology: The Vascular System*, 32, 661–667. [http://dx.doi.org/10.1016/S0306-3623\(98\)00238-9](http://dx.doi.org/10.1016/S0306-3623(98)00238-9)
- [29]. SAS (2008). *Statistical Analysis System. SAS Version 9.2 user's guide*. Cary, NY: SAS institute.
- [30]. Aremu, M. O., Olaofe, O. and Akintayo, E. T. (2006). A comparative study on the chemical and Amino acid Composition of Some Nigerian Underutilized legume flours. *Pakistan Journal of Nutrition*, (5):34-38.
- [31]. IHEMEJE, A., Nwachukwu, C. N. and Ekwe, C. C. (2015). Production and quality evaluation of flavoured yoghurts using carrot, pineapple, and spiced yoghurts using ginger and pepper fruit. *African Journal of Food Science*, 9(3):163-169
- [32]. Mbaeyi-Nwaoha I. E., Nnagbo, C. L., Obodoechi, C. M., Nweze, B. C., Okonkwo, T. M. (2017). Production and evaluation of yoghurt contained local stabilizers - *Brachystegia eurycoma* ('Achi') and *Detarium microcarpum* ('Ofo'). *International Journal of Biotechnology and Food Science* Vol. 5(2), pp. 23-31
- [33]. Younous, S., Masud, T. and Aziz, T (2012) Quality evaluation of market yoghurt/Dahi. *Journal of Nutrition* 5, 226 -230.
- [34]. Matter, A., Eman, A., Mahmoud, A. M. and Zidan, N. S (2016). Fruit Flavored Yoghurt: Chemical, Functional and Rheological Properties *International Journal of Environmental and Agriculture Research*, 5(2) 57-66
- [35]. Tamang, J. P., Shin, D. H., Jung, S. J. and Chae, S. W. (2016). Functional Properties of Microorganisms in Fermented Foods. *Front. Microbiol.* 7(578):1-13.
- [36]. Kim, S. S., Shin, S. G., Chang, K. S., Kim, S. Y., Noh, B. S. and Bhowmik, S. R. (1997). Survival of lactic acid bacteria during microwave vacuum-drying of plain yoghurt. *Lebensm.-Wiss. U.- Technology*, 30: 573–577.
- [37]. Akubor, P. I. (2016). Physicochemical, microbiological and sensory properties of yoghurt supplemented with carrot juice. *Nigerian Journal of Nutritional Sciences*, 32(1): 15-20
- [38]. Galal, E. A., Mahmoud, A., EL-Fakhany, A. and Moawad, A. (2003). Effect of adding carrot puree on Organoleptic, chemical and microbiological quality of UF soft cheese. In: *Proceedings of the 1st International Conference. Food for better health*. Cairo, Egypt: NRC. p. 18-20
- [39]. Lubna, R., Tahir, Z., Ambreen, S., Nazia, K., Ubaid ur, R. and Atif, L. (2020) Augmenting yogurt quality attributes through hydrocolloidal gums. *Asian-Australas Journal of Animal Science*, 33(2): 323-331.
- [40]. Smit, G. (2003). *Dairy Processing*. CRC Pres. 2000 Corporate Blvd, NW, Boca Raton FL 33431, USA.
- [41]. Izadi, Z., Nasirpour, A., Garoosi, G A. and Tamjidi, F. (2014). Rheological and physical properties of yogurt enriched with phytosterol during storage. *Journal of Food Science Technology*, 1-6.

- [42]. Abu-Jdayil, B. and Mohameed, H. (2002). Experimental and modelling studies of the flow properties of concentrated yoghurt as affected by the storage time. *Journal of Food Engineering*, 52: 359-365.
- [43]. Lim, S. M. (2013) Microbiological, Physicochemical, and Antioxidant Properties of Plain Yogurt and Soy Yoghurt. *Korean Journal of Microbiology*, 49: 403-414
- [44]. Celik, S., Bakırcı, I. and Şat, I. (2006). Physicochemical and organoleptic properties of yoghurt with cornelian cherry paste. *International Journal of Food Properties*, 9: 401-408.
- [45]. Gul, K., Singh, A. K., & Jabeen, R. (2016). Nutraceuticals and functional foods: The foods for the future world. *Critical Reviews in Food Science and Nutrition*, 56, 2617–2627. <http://dx.doi.org/10.1080/10408398.2014.903384>
- [46]. Badejo, A.A., Osunlakin, A.P., Famakinwa, A., Idowu, A. O. and Fagbemi, T.N. (2017). Analyses of dietary fibre contents, antioxidant composition, functional and pasting properties of plantain and Moringaoleifera composite flour blends, *Cogent Food & Agriculture*, 3:1, 1278871, DOI: 10.1080/23311932.2017.1278871
- [47]. Panche, A. N.; Diwan, A. D.; Chandra, S. R. (2016). Flavonoids: An Overview. *J. Nutr. Sci.* 5. DOI: 10.1017/jns.2016.41
- [48]. Aparadh, V. T., Naik, V. V., and Karadge, B. A. (2012). Antioxidative properties (TPC, DPPH, FRAP, metal chelating ability, reducing power and TAC) within some cleome species. *Annali Di Botanica*, 2, 49–56
- [49]. Ultana, B., Anwar, F. and Przybylki, R. (2007) Antioxidants Potential of Corn cob Extracts for Stabilization of Corn Oil Subjected to Microwave Heating. *Food Chemistry*, 104, 997-1005
- [50]. Papadimitriou, C. G., Mastrojiannaki, A. V., Silva, A. V., Gomes, A. M., Malcata, F. X. and Alichanidis, E. (2007). Identification of Peptides in Traditional and Probiotic Sheep Milk Yoghurt with Angiotensin I-Converting Enzyme (ACE)-Inhibitory Activity. *Food Chemistry*, 105, 647-656.
- [51]. Yildiz, O. and Eyduran, S. P. (2009). Functional Components of Berry Fruits and Their Usage in Food Technologies. *African Journal of Agricultural Research*, 4, 422-426.
- [52]. Yuksel, Z., Avci, E. and Erdem, Y.K. (2010) Characterization of Binding Interactions between Green Tea Flavanoids and Milk Proteins. *Food Chemistry*, 121, 450-456.
- [53]. Water, J. D., Keen, C. L. and Gershwin, M. E. (1999). The Influence of Chronic Yogurt Consumption on Immunity. *Journal of Nutrition*, 129: 1492S-1495S.
- [54]. Massey, L. K. (2001). Dairy Food Consumption, Blood Pressure and Stroke. *Journal of Nutrition*, 131: 1875-1878.
- [55]. Ateteallah, A. H. and Osman, A. K. (2019) Antioxidant, Some Flavor Components, Microbiological and Microstructure Characteristics of Corn Milk Yoghurt. *Food and Nutrition Sciences*, 10, 551-560.
- [56]. Ecem, A. (2022). The Effect of Fermentation Time and Yogurt Bacteria on the Physicochemical, Microbiological and Antioxidant Properties of Probiotic Goat Yoghurts. *An Acad Bras Cienc* 94(3): e20210875 DOI 10.1590/0001-376520220210875
- [57]. Ozcan, T., Sahin, S., Akpinar-bayizit, A. and Yilmaz-ersan, L. (2019). Assessment of antioxidant capacity by method comparison and amino acid characterization in buffalo milk kefir. *International Journal of Dairy Technology*, 72: 65-73.
- [58]. Yoon, J. W., Ahn, S. I., Jhoo, J. W. and Kim, G. Y. (2019). Antioxidant Activity of Yogurt Fermented at Low Temperature and Its Anti-inflammatory Effect on DSS-induced Colitis in Mice. *Food Science Animal Resources*, 39(1): 162-176.

Ibhaze, G.A., Olatunji, D. I., Onibi, G.E, et. al. "Comparative study on different local hydrocolloids on quality of set-type yoghurt made from bovine milk." *IOSR Journal of Environmental Science, Toxicology and Food Technology (IOSR-JESTFT)*, 17(2), (2023): pp 41-48.