Evaluation of the Anti-malarial effect of the methanolic leaf extract of *Vernonia glaberrima* (Asteraceae)

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Abstract: The emergence of drug-resistant strains has compromised the efficacy of several anti-malarial drugs, including the artemisinins. Many species of Vernonia, family Asteraceae, have been employed in traditional medicine for the management of diabetes mellitus, microbial infections and malaria. The anti-malarial effect of the methanolic leaf extract of Vernonia glaberrima was investigated using Chloroquine-sensitive Plasmodium berghei (NK65) infected mice, according to Riley and Peters curative test model. Three test groups of mice were orally administered with the suspension of the graded doses of the extract at 300, 150 and 75 mg/Kg body weight for four consecutive days; a reference drug, Chloroquine phosphate (5mg/Kg) group, and the negative control group (0.2ml distilled water) were similarly administered. The level of chemo-suppression obtained on the eighth day was a function of the reduction in parasitemia as estimated from the mice tail blood smear. The mean survival time (in days) for each group of mice was also determined over a period of 28 days postinfection. The results showed a dose-dependent chemo-suppression by the three extract groups 300, 150 and 75 mg/Kg at 62.1%, 54.3% and 32.2% respectively, while Chloroquine (5mg/Kg) exhibited a higher suppression at 73.4%. Only the Chloroquine and 300mg/kg groups produced statistically significant (p<0.05) mean survival time of 19 and 17 days when compared to the negative control. The study demonstrated the anti-malarial effect of the leaf extract of V. glaberrima and lends credence to its ethno-medicinal use in treating malaria infection. Keywords: Vernonia glaberrima extract, Plasmodium berghei, anti-malarial, Curative test

I. Introduction

Malaria is a major public health problem affecting not less than 40% of the world's population (Snow et al., 2005). An estimated 1.2 billion are at high risk of transmission (≥ 1 case per 1000 population), more than half of which live in the African regions and Nigeria alone accounts for a quarter of all malaria cases in Africa (WHO, 2008). Malaria is the most common disease in Nigeria affecting more than 100 million people annually (Reyburn, 2010). In 2010, about 216 million people were infected with 655,000 mortality, worldwide. African region accounts for 81% and 91% of the cases and deaths respectively, with 86% of the mortalities observed among the under-fives (WHO, 2014). The emergence of drug-resistant strains has compromised the efficacy of several anti-malarial drugs, including artemisinins, thus necessitating the need for the discovery of other novel anti-malarial agents (Phyo, 2012; Nkhoma et al., 2012). Many anti-malarials including artemisinins were discovered from plant sources (Klayman 1985; Cragg, Newman et al., 1997; Newman and Cragg 2007; Newman and Cragg, 2012). Ninety-nine plant species in Brazil were reportedly claimed to have antimalarial activity (Milliken, 1997). In Nigeria, over 100 plant species including Vernonia cinera and Vernonia amygdalina are used in herbal medicine as remedies for malaria (Adebayo and Krettli, 2010). The genus Vernonia, family Asteraceae, has been employed in traditional medicine for the management of varied diseases including malaria (Muregi et al., 2003); microbial infections (Al-Magboul et al., 1988; Erastor et al., 2006), as analgesics and anti-inflammatory (Njan et al., 2008), cytotoxic (Kuo et al., 2003; Williams et al., 2005) and, many species have been reported to amend the hyperketonaemia, hyperlipidaemia and hypercholesterolaemia associated with diabetes mellitus (Nimenibo-Uadia, 2005). Sesquiterpene lactones are the major bioactive constituents isolated from the Vernonia species. They have been reported to exhibit antimalaria, antileishmania, antischistosoma, cytotoxic, anti-microbial and anti-inflammatory effects (Toyang and Verpoorte, 2013).

Vernonia glaberrima Welw. Ex O. Hoffm (family Asteraceae), Shìwaákár-ján-gágári (Hausa language - N. Nigeria) is an erect shrub, 2 meters high, found on hillside grassland in Guinea to Northern Nigeria, Western Cameroon and Central Africa to Angola. It is reported to be used traditionally against malaria, migraine, psoric and dysmenorrhoea (Burkill, 1985); it is also employed for treating pain, inflammation, vertigo

and microbial infections (Abdullahi *et al.*, 2015). This study was aimed at evaluating the anti-malarial properties of the crude methanol leaf extract of *V. glaberrima*.

II. Materials And Methods

Collection and Identification of the Plant Sample

The whole plant material of *Vernonia glaberrima* was collected in Nassarawa State, Northern-Nigeria in June 2012 during rainy season. It was authenticated by U.S. Gallah of the Herbarium unit, Department of Biological Sciences, Ahmadu Bello University, Zaria. A voucher specimen (No. 899) was deposited at the herbarium for future reference. The leaves were removed, air-dried, powdered, labeled and stored in air-tight container prior to extraction.

Preparation of extract

The leaves were removed, shade dried, pulverized, labelled and stored at room temperature in an airtight container prior to extraction. The Powdered leaves (2500g) was extracted with 70% methanol using maceration method for 10 days with occasional shaking. The extract was evaporated in-vacuo using rotary evaporator at 40°C to obtain a gummy greenish product (400g) subsequently referred to as the crude methanol leaf extract VGLE.

Experimental Animals

Swiss albino mice of either sex weighing (15-38g) obtained from the Animal House Facility of the Department of Pharmacology and Toxicology, Ahmadu Bello University, Zaria, Nigeria, were used for the study. They were fed with commercial feeds and water *ad libitum* and maintained under standard conditions (12hr light and 12hr dark cycle) in propylene cages at 25°C room temperature. All experimental procedures were performed in accordance with the guidelines of the Animal right Ethics Community of the university.

Acute toxicity study

The safety of the extract was evaluated as described by (Lorke, 1983). The route of administration was intra-peritoneal. In the first phase, nine mice of either sex were divided into three groups containing three mice each. The first, second and third groups received 10mg/kg, 100mg/kg and 1000mg/kg respectively. The rats were observed continuously for behavioral, neurological, autonomic and any lethality in first 24 hours. From the result of the first phase, three mice were used for the second phase. They were given different doses 1600mg/kg, 2900mg/kg and 5000mg/kg of the extract, and were observed for any sign of toxicity and possibly death during the 24 hours. The median lethal dose was calculated using the following formula;

$LD_{50} = \sqrt{minimal lethal dose \times maximal survival dose}$

Plasmodium berghei parasite

The *NK65* Chloroquine sensitive strain of *Plasmodium berghei* was purchased from National Institute for Pharmaceutical Research and Development (NIPRD), Idu, Abuja, Nigeria. The parasite was conserved in the Department of Pharmacology and Toxicology Laboratory, Usmanu Danfodiyo University Sokoto via passage of blood from infected into healthy mice.

Parasite Induction

Blood containing infected red cells (from infected donor mice) were diluted with normal saline so that 0.2mls will contain $1*10^6$ parasitized red cells. The mice were inoculated with 0.2mls of the infected blood intraperitonealy, using hypodermic needle fitted to 1-ml syringe.

Evaluation of schizontocidal activity of V. glabberima on established infection (Curative or Rane test)

Riley and Peter method (Riley, Allen *et al.*, 1991) was employed to evaluate the curative potential of *Vernonia glaberrima*. On day one (1), twenty five mice were inoculated with 0.2mls of blood containing 10^6 *Plasmodium berghei* infected red cells, intraperitoneally. Three (3) days later, the mice (with established infection) were randomly divided into 5 groups of 5 mice each and treated for four consecutive days (days 4, 5, 6 and 7). The first three groups were orally treated with 10ml/kg of the dissolved extract daily at 300, 150 and 75mg/Kg body weight respectively. Two control groups (positive and negative) were similarly dosed with 10 ml/kg of chloroquine phosphate (at 5mg/Kg) and distilled water respectively. On day 8 of the experiment (a day after completion of treatment), tail blood was collected from each mouse and thin films of the samples were stained with leishman. Buffered water (pH 6.8) was added to the film, which was kept for 8-10 minutes, cleaned with cotton wool and allowed to air dry, before it was viewed at × 100 magnifications on the microscope. The average percentage parasitaemia and percentage of parasite suppression were calculated in each of the groups as shown below (Penna-Coutinho, Cortopassi *et al.*, 2011).

Percentage Parasitaemia= Number of infected RBCS X100

Total no. of RBCS examined

% suppression = $\frac{PC - PTG}{PC}$

Where PC is the parasitaemia in the untreated group

PTG is the parasitaemia in the test group.

Drugs that reduced parasitaemia by 29-40% were considered as partially active antimalarials, while those that produced greater than 40% reduction in parasitaemia were considered active antimalarials.

Determination of Mean Survival Time

The duration of 28 days survival was recorded for each mouse. Mean survival time (MST) was calculated using the following formula (Penna-Coutinho, Cortopassi *et al.*, 2011).

MST=<u>Sum of survival time of all mice in a group (days)</u>

Total no. of mice in that group

III. Results

Acute toxicity test

The LD₅₀ was extrapolated to be 1265mg. This indicates that the experimental doses used are relatively safe.

Curative antiplasmodial activity and the mean survival time

Α high suppression (>40%) of parasites, in animals with established infection was observed in the groups treated with chloroquine, graded doses of the extract (300 and 150 mg/Kg), but only mild (29-40%) in the group treated with 75mg/kg. There was no significant difference (P>0.05) in chemosuppression between mice administered with 300mg/Kg, 150 mg/Kg and chloroquine-treated group. On the other hand, mice treated with 75mg/Kg did not differ significantly (P>0.05) in chemosuppression from the vehicle-treated control group (Table 1). The groups of mice treated with chloroquine and 300mg/Kg produced a statistically significant increase (P<0.05) in mean survival time compared with vehicle-treated animals (Table 1). No significant difference (P>0.05) was observed in the mean survival time of the groups of mice treated with either 150 or 75 mg/Kg of the plant extract when compared to the negative control group (Table 1).

IV. Discussion

In this study, the anti-malaria traditional claim of *Vernonia glaberrima* was evaluated. The rodent model was employed in the investigation because it takes into account potential pro drug formation and immune system effects in combating infection (Fidock, Rosenthal *et al.*, 2004). The study showed that *V. glaberrima* has a curative anti-malarial effect at a dose of 150mg/kg and 300mg/kg. The high percentage cure, similar to that of chloroquine was observed for the groups treated with 150 and 300mg/kg. Moreover, the positive control and the group treated with 300mg/kg showed statistically significant increase in mean survival time when compared with the negative control. The finding that the group treated with 150mg/kg produces significant curative antimalarial activity which did not translate into increased mean survival time compared to the negative control is a paradox suggesting caution and further evaluation in the interpretation of results. A possible explanation is that the extract was toxic to mice at the dose administered, thus reducing the survival period even at a low parasite load. This paradox may need to be explored in further studies.

The exact mechanism of antiplasmodium activity of the plant extract was not explicated, though antiplasmodial effect of extracts from plants have been ascribed to their phytochemical components like flavonoids, terpenes, alkaloids, saponins and glycosides (Iwu, Duncan *et al.*, 1999; Abosi and Raseroka 2003; Ayoola, Coker *et al.*, 2008; Kaur, Jain *et al.*, 2009). Bioactive sesquiterpene lactones such as Vernodalin, Vernomygdin, Vernodalol and Epivernodalol have been isolated from other Vernonia species (Kupchan *et al.*, 1969; Igile *et al.*, 1995; Ganjian *et al.*, 1983; Owoeye *et al.*, 2010) and may have contributed to the antimalarial activity. The plant *V. glaberrima* has the potential to be further explored for its anti-malaria effect through the isolation and characterization of the active principles.

V. Figures and Tables

Table 1: Curative effect and Mean survival time (MST) of chloroquine and Vernonia glaberima against P. berghei infection in mice.

S/No	Treatment Groups (mg/kg)		Parasite count	% Cure	Mean Survival Time (MST)
1	Chloroquine	5	0.55±0.5*	73.4	19.20±2.69*
2	VGLE	300	0.78±0.11*	62.1	17.20±3.04*
3	VGLE	150	0.94±0.18*	54.3	16.00±3.49
4	VGLE	75	1.40±0.19**	32.2	14.20±2.75
5	Distilled water		2.06±0.69	-	10.80±2.03

Values are expressed as mean \pm SEM, n = 5; Values of the group with superscript *are statistically significant (p<0.05) compared to negative control group; Values with superscript ** are statistical significant (p<0.01)compared to negative and positive control groups.

VI. Conclusion

The study demonstrated the potent anti-plasmodium activity of the leaf extract of V. glaberrima and validates its use in traditional medicine for treating fever and pain associated with malaria infection.

References

- [1]. Abdullahi MI, Uba A, Yaro AH, Maxwell O, Yusuf AJ, Kabir S, Alhasan AM, Umar A, Bello SS, Nasir I (2015). Phytochemical Screening, Acute Toxicity study and Evaluation of Antidiabetic properties of the methanolic leaf extract of Vernonia glaberrima (Asteraceae). J Pharm Chem Biol Sci. 3(2): 169-177.
- [2]. Abosi AO and BH Raseroka (2003). "In vivo antimalarial activity of Vernonia amygdalina." British Journal of Biomedical Science. 60(2): 89.
- Adebayo JO and Krettlia AU. (2011). Potential antimalarials from Nigerian plants: A review. Journal of Ethnopharmacology, 133: 289-302 [3].
- Al-Magboul AZ, Bashir AK, Salih AM, Farouk A, Khalid SA. (1988). Antimicrobial activity of certain Sudanese plants used in folkloric [4]. medicine: screening for antibacterial activity (V). Fitoterapia. 59: 57-62.
- Ayoola G, H Coker et al., (2008). "Phytochemical screening and antioxidant activities of some selected medicinal plants used for malaria [5]. therapy in Southwestern Nigeria." Tropical Journal of Pharmaceutical Research. 7(3): 1019-1024.
- Burkill HM. (1997). The useful plants of West Tropical Africa, Royal Botanical Gardens, Kew. 4: 275. [6].
- Cragg GM, DJ Newman, et al., (1997). "Natural products in drug discovery and development." Journal of natural products. 60(1): 52-60. [7].
- [8]. Erasto P, Grierson DS, Afolayan AJ. (2006). Bioactive sesquiterpene lactones from the leaves of Vernonia amygdalina. Journal of Ethnopharmacology. 106: 117-120.
- Fidock DA, PJ Rosenthal et al., (2004). "Antimalarial drug discovery: efficacy models for compound screening." Nature Reviews Drug [9]. Discovery. 3(6): 509-520.
- Ganjian I, Kubo I, Fludzinski P. (1983). Insect antifeedant elemanolide lactones from Vernonia amygdalina. Phytochemistry. 22:2525-2526. [10]. Igile GO, W Oleszek et al., (1994). "Flavonoids from Vernonia amygdalina and their antioxidant activities." Journal of Agricultural and [11].
- Food Chemistry. 42(11): 2445-2448.
- [12]. Iwu M, AR Duncan et al., (1999). "New antimicrobials of plant origin." Perspectives on new crops and new uses. ASHS Press, Alexandria, VA· 457-462
- Kaur K, M Jain et al., (2009). "Antimalarials from nature." Bioorganic & Medicinal Chemistry. 17(9): 3229-3256. [13].
- [14]. Klayman DL (1985). "Qinghaosu (artemisinin): an antimalarial drug from China." Science. 228(4703): 1049-1055. [15]. Kuo YH, Kuo YJ, Yu AS, Wu MD, Ong CW, Kuo LMY, Huang JT, Chen CF and Li SY. (2003). Two novel sesquiterpene lactones,
- cytotoxic vernolide-A and B, from Vernonia cinerea. Chemical and Pharmaceutical Bulletin. 51: 425-426. Kupchan SM, Hemmnigway RJ, Karim A, Werner D. (1969). Tumor inhibitors. XLVII Vernodalin and Vernomygdin. Two new cytotoxic [16]. sesquiterpene lactones from Vernonia amygdalina Del. J. Org. Chem. 34:3908-3911.
- Lorke, D. (1983). "A new approach to practical acute toxicity testing." Archives of Toxicology. 54(4): 275-287. [17].
- [18]. Milliken W. (1997). "Traditional anti-malarial medicine in Roraima, Brazil." Economic Botany. 51(3): 212-237.
- [19]. Muregi FW, Chhabra SC, Njagi ENM, Lang'atThoruwa CC, Njue WM, Orago ASS, Omar SA, Ndiege IO. (2003). Invitro antiplasmodial activity of some plants used in Kisii, Kenya against malaria and their chloroquine potentiation effects. J Ethnopharmacol. 84235-84239.
- [20]. Newman DJ and GM Cragg (2012). "Natural products as sources of new drugs over the 30 years from 1981 to 2010." Journal of natural products.75(3): 311-335
- [21]. Newman DJ and GM Cragg. (2007). "Natural Products as Sources of New Drugs over the Last 25 Years." Journal of natural products. 70(3): 461-477.
- Nimenibo-Uadia, R. (2005). "Effects of Vernonia amygdalina in alloxan-induced diabetic albino rats." Journal of Medical Laboratory [22]. Science. 12(1): 25-31.
- Njan AA, Adzu B, Agaba AG, Byrugaba D, Diaz-Llera S and Bangsberg DR. (2008). Analgesic and Antiplasmodial activities and [23]. Toxicology of Vernonia amygalina. Journal of Medical foods. 11: 574-581.
- [24]. Nkhoma SC, Nair S, Al-Saai S, Ashley E, McGready R, Phyo AP, Nosten F, Anderson TJ. (2013). Population genetic correlates of declining transmission in a human pathogen. Mol Ecol. 22(2):273-85.
- Owoeye O, Yousuf S, Akhtar MN, Qamar K, Dar A, Farombi EO, Onwuka SK, Choudhary MI. (2010). Another Anticancer Elemanolide [25]. from Vernonia amygdalina Del. Int. J. Biol. Chem. Sci. 4:226-234.
- [26]. Penna-Coutinho J, WA Cortopassi et al., (2011). "Antimalarial activity of potential inhibitors of Plasmodium falciparum lactate dehydrogenase enzyme selected by docking studies." *PloS one* 6(7): 21237. Phyo AP, S Nkhoma, *et al.*, (2012). "Emergence of artemisinin-resistant malaria on the western border of Thailand: a longitudinal study."
- [27]. The Lancet, 379(9830): 1960-1966.
- Reyburn, H. (2010). "New WHO guidelines for the treatment of malaria." BMJ340. [28].
- [29]. Riley EM, SJ Allen et al., (1991). "Association between immune recognition of the malaria vaccine candidate antigen Pf155/RESA and resistance to clinical disease: a prospective study in a malaria-endemic region of west Africa." Transactions of the Royal Society of Tropical Medicine and Hygiene. 85(4): 436-443.
- [30]. Snow RW, Guerra CA, Noor AM, Myint HY, Hay S I (2005). The global distribution of clinical episodes of Plasmodium falciparum malaria. Nature, 434, 214-217.
- Tor-Anyiin T, R Sha'ato et al., (2003). "Ethnobotanical survey of anti-malarial medicinal plants amongst the Tiv people of Nigeria." Journal [31]. of herbs, spices & medicinal plants. 10(3): 61-74. Toyang NJ and R Verpoorte. (2013). "A review of the medicinal potentials of plants of the genus Vernonia (Asteraceae)." Journal of
- [32]. Ethnopharmacology. 146(3): 681-723.
- Trease GE and Evans WC. (1983). Textbook of Pharmacognosy, 12th (Ed) Balliers Tindall, London. 343-384. [33].
- WHO, 2008. World Malaria Report 2008. World Health Organization, Geneva, pp. 7-15, 99-101. [34].
- Williams RB, Norris A, Slebodnick C, Merola J, Miller JS, Andriantsiferana R, Rasamison, VE and Kingston DGI. (2005). Cytotoxic [35]. sesquiterpene lactones from Vernonia pachyclada from the Madagascar Rain forest. Journal of Natural Products. 68: 1371–1374.
- [36]. World Health Organization. (2014). Malaria Factsheet. Geneva, Switzerland.