Nutrients decrease Al toxicity to resistant variety of wheat (Raj-3077)

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Abstract: Wheat is a second major staple food crop in south Asia after, rice. The wheat acreage to South Asia (India, Pakistan, Nepal, and Bangladesh) is more than 36 million hectare (FAO 2007). But it is also seems less to fulfill the needs of population. Different environmental factors and pollutants decrease its yield further. Aluminum decreases its root- shoot length and biomass at different concentrations; but nutrient supply decreases the toxic effects.

I. Introduction

Aluminum is found natural in soil, water, air (CEPA, 2000; ATSDR, 2006). Al ion contributes to soil acidity through their tendency to hydrolyze. The hydrogen ions released give a very low pH value in the soil solution and are a major source of hydrogen in most acid soils (Richard, 1998). Aluminum (Al) is toxic to plants at low pH and can begin to inhibit root growth within 3 h in solution experiments and finally decrease in yield. Al interferes with uptake or transport and utilization of essential nutrients like Ca, Mn, P, Mg, B, Fe, Cu, K, and Zn (Keltjens and Tan, 1993; Keltjens, 1995; Lukaszewski and Belvins, 1996; Slaski *et al*, 1996; Taylor *et al*, 1998; Lidon *et al*, 2000; Guo *et al*, 2003, 2007; Olivares *et al*, 2009).

Plant Materials

II. Material and method

Wheat (*Triticum aestivum*) Raj-3077 is an early maturity (115-120 days) medium height (90 cm) variety, released in Rajasthan, 1989 under wheat Breeding Scheme, RAU, ARS, Durgapura. It is resistant to brown and yellow rust, tolerant to Saline and alkaline conditions and well adapted to drought. Its grains are lustrous amber medium bold semi-hard with good chapatti making quality.

Seeds were surface sterilized with 0.1% mercuric chloride (HgCl₂). Equal sized seeds were sown at equal distance in Petri dishes lined with filter paper and germinated in dark at $25\pm5^{\circ}$ for 24 h.

Preparation of stock solution of Aluminum

1000 ppm stock solution was prepared with Analar grade aluminum sulphate (Al₂ (SO_4)₃.16H₂O). Various dilutions were prepared using distilled water and Hoagland's Solution respectively.

Hoagland's' medium composition

KH₂PO₄, KNO₃, MgSO₄.7H₂O, Boric acid, MnCl₂.H₂O, ZnSO₄.7H₂O, CuSO₄.5H₂O, Molybdic acid, Fe-EDTA make up to one liter of solution.

Treatments for wheat

Group A- Al solutions (100, 250, 500, 750, 1000ppm) made in distilled water; Group B- Al solutions (100, 250, 500, 750, 1000ppm) made in Hoagland's nutrient medium. Group A control was set in distilled water whereas of Group B in Hoagland's nutrient solution.

Growth conditions

The plants were grown under 500 watt fluorescent light bulb. This light stayed on for 10 hours a day for 10 days. Harvesting was after 10 days. Root numbers were counted while root-shoot lengths were measured with the help of scale. Roots and shoots were separated and oven dried at 60°C for two days. Dry weight was taken after two days on electronic balance.

III. Result

Toxicity of different concentrations of Al on wheat

Germination was recorded 95-100% at all concentrations i.e. no adverse effect on germination. Symptoms of toxicity were noted at high concentrations of Al, reduced development of the roots, stubby appearance and were brownish color. Above ground portion of the plant, typical symptoms are small leaves, and shortened and thickened internodes

There was dose dependent reduction in root number, shoot and root length and their dry weight, with the exception at 100 ppm at which both shoot and root growth were least affected (Table 1). I found 100ppm Al concentration to be non toxic to seedlings. Rather it was favorable to shoot growth.

Toxicity of different concentrations (diluent Hoagland's solution) on wheat

There was no change in shoot number of wheat (*T. aestivum*) but root numbers were higher than control. Maximum (29.0%) number of roots was found at 750 ppm.

Shoot length increased a little at 100ppm and 250ppm but decreased markedly at 750 and 1000 ppm (Table. 2). The root length decreased gradually with increased Al concentration, with exception at 100 ppm showing a little increase (14.5%). The smallest roots were found in seedlings growing at 1000 ppm (Table. 2).

Compared with control, both shoot and root dry weights increased at 100 and 250ppm but decreased at higher concentrations, being minimum at 1000 ppm (Table 2). Similar trend was noted for total dry biomass of seedlings. R/S ratios declined at lower concentrations (100-500ppm) but increased at higher concentrations (750-1000ppm).

Al toxicity was low when different dilutions of Al were made in Hoagland's medium suggesting plant nutrients in the medium provides protection to seedlings.

Table.	1 I UMER	of unterent concentrations of 211 (undeed with distinct water) on wheat seconds.							
	Shoot Number	Root Number	Shoot length (cm)	Root Length (cm)	Dry weight of shoot	Dry weight of root	Total dry weight (mg)	Root/Shoot dry wt. ratio	
					(mg)	(mg)	× 0,		
Control	1.0+0.0	6.4±0.2	13.2±0.7	11.7±0.8	11.7±0.8	11.2±0.9	22.9±0.9	0.9	
100	1.0±0.0	6.0±0.3	15.6±0.7*	9.2±0.7	14.1±1.1	11.7±0.1	25.8±1.9	0.8	
ppm	(Nil)	(-6.3%)	(+18.2%)	(-21.4%)	(+20.5%)	(+4.5%)	(+12.7%)	(-15.6%)	
250	1.0±0.0	7.4±0.6	11.9±1.3	6.2±1.1**	11.5±1.4	9.9±0.6	21.4±1.8	0.9	
ppm	(Nil)	(+15.6%)	(-12.1%)	(-47.0%)	(-1.7%)	(-11.6%)	(-6.6%)	(-10.4%)	
500	1.0±0.0	7.2±0.4*	10.6±0.6	5.3±0.4***	9.2±0.5*	8.4±0.6**	17.7±0.7**	0.9	
ppm	(Nil)	(+12.5%)	(-19.7%)	(-54.7%)	(-21.4)	(-25.0%)	(-22.7%)	(-6.3%)	
750	1.0±0.0	7.4±0.5	1.7±0.04***	1.9±0.5**	1.9±0.5**	6.3±0.6*	8.3±0.9***	3.3	
ppm	(Nil)	(+15.6%)	(-87.1%)	(-83.8%)	(-83.8%)	(-43.8%)	(-63.8%)	(+243.8%)	
1000	1.0±0.0	7.2±0.5	1.5±0.4***	1.9±0.7***	1.9±0.7***	5.8±0.8**	7.8±1.4***	3.05	
ppm	(Nil)	(+12.5%)	(-88.6%)	(-83.8%)	(-83.8%)	(-48.2%)	(-65.9%)	(+217.7%)	

*Significance at 5% ** 1% and *** 0.1% probability, data in parenthesis indicate percent change in values in comparison to control

Table: 2 Toxicity of different concentrations of Al (diluted with Hoagland's solution) on wheat seedlings

	Shoot Number	Root Number	Shoot length (cm)	Root Length (cm)	Dry weight of shoot (mg)	Dry weight of root (mg)	Total dry weight (mg)	Root/Shoot dry wt. ratio
Control	1.0±0.0	6.2±0.2	10.1±0.4	6.2±0.6	10.1±0.7	11.7±0.8	21.8±0.9	1.2
100	1.0±0.0	6.4±0.2	10.5±0.3	7.1±1.0	10.7±0.9	10.8±0.4	21.5±1.1	1.0
ppm		(+3.2%)	(+3.9%)	(+14.5%)	(+5.9%)	(-7.7%)	(-1.4%)	(-12.9%)
250	1.0±0.0	7.0±0.4	10.9±1.2	4.8±0.5	11.2±1.4	9.7±0.6	20.8±1.7	0.9
ppm		(+12.9%)	(+7.9%)	(-22.6%)	(+10.9%)	(-17.1%)	(-4.6%)	(-25.0%)
500	1.0±0.0	6.8±0.2	10.1±1.7	4.6±0.4	9.5±1.7	8.4±0.5	17.8±1.6	0.9
ppm		(+9.7%)	(Nil)	(-25.8%)	(-5.9%)	(-28.2%)	(-18.3%)	(-24.1%)
750	1.0±0.0	8.0±0.0**	5.4±0.6**	3.7±1.0*	4.5±0.5**	6.8±0.4**	11.3±0.6**	1.5
ppm		(29.0%)	(-46.5%)	(-40.3%)	(-55.4%)	(-41.9%)	(-48.2%)	(+30.2%)
1000	1.0±0.0	7.4±0.5	1.5±0.5**	2.1±02**	2.0±0.7**	5.9±0.7**	8.0±1.3***	2.9
ppm		(+19.4%)	(-85.1%)	(-66.1%)	(-80.2%)	(-49.6%)	(+63.3%)	(+154.3%)

*Significance at 5% ** 1% and *** 0.1% probability, data in parenthesis indicate percent change in values in comparison to control

IV. Discussion

Aluminum exposure affected plant growth adversely. In the present investigation, Al stress decreased plant height, root length and plant biomass. These findings are in agreement with other workers (Mossor-Pietraszewska, 2001; Ma, 2007; Zheng *et al.*, 2007, Jiang *et al.*, 2008; Diaz, 2011).

Wheat root and shoot growth were affected greatly in Al treatments (250, 500, 750 and 1000ppm) prepared after dilution of stock solution with distilled water (Table 1). Al toxicity at similar concentrations was relatively less

when dilutions (250 and 500 ppm) of stock solution were made in Hoagland's nutrient (Table 2). This may be either on account of chelation of Al with EDTA or competition of Al with divalent cations for absorption. Bartlett and Riego (1972) reported same on maize seedlings.

The lower concentration of Al (100ppm) had no adverse effects on wheat seedlings might be due to less Al was accumulated in tolerant wheat (Darko *et al.*, 2004). Kochian (1995) reported that Al might be bound inactively to some component of the cell wall or cell membrane, or to ligands found in the cytoplasm or vacuoles.

V. Conclusion

The lower concentrations were not toxic to Raj-3077 resistant variety of wheat and dilutions made with Hoagland's nutrient medium supports seedling growth and decreased Al toxicity.

Reference

- [1]. ATSDR, 2006. Draft toxicological profile for aluminum. Atlanta, US Department of Health and Human Services, Public Health Service, Agency for Toxic Substances and Disease Registry.
- [2]. Bartlett, R. J. and Riego, D. C.1972. Effects of chelation on the toxicity of aluminum. Plant & Soil 78: 79-84.
- [3]. CEPA. 2000. State of the Science Report for Aluminum chloride, Aluminium nitrate, and Aluminium sulphate. Canadian Environmental Protection Act.
- [4]. Darko, E., Ambrus, H., Banyai, E. S., Fodor, J. Bakos, F. and Barnabas, B. 2004. Aluminum toxicity, Al-tolerance and oxidative stress in an aluminum sensitive wheat genotype and in Al-tolerant lines developed by *in vitro* microspore selection. *Plant Sci.* 166: 583-591.
- [5]. Diaz, D. R. 2011 Symptoms of aluminum toxicity of wheat on low-pH soils. AG Professonal magazine
- [6]. FAO 2007. Statistical database http://www.fao-stat.fao.org
- [7]. Guo, T.R., Zhang, G.P., Lu, W.Y., Wu, H.P., Wu, F.B., Chen, J.X. and Zhou, M.X., 2003. Effect of Al on dry matter accumulation and Al and nutrients in barleys differing in Al tolerance. *Plant Nutri*. *Ferti. Sci.* 9: 324–330.
- [8]. Guo, T.R., Zhang, G.P., Zhou, M.X., Wu, F.B. and Chen, J.X., 2007. Influence of aluminum and cadmium stresses on mineral nutrition and root exudates in two barley cultivars. *Pedosphere* 17: 505–512.
- [9]. Jiang, H.X., Chen, L.S., Zheng, J.G., Han, S., Tang, N. and Smith, B.R. 2008. Aluminum-induced effects on Photosystem II photochemistry in citrus leaves assessed by the chlorophyll a fluorescence transient. *Tree Physiol*. 28:1863-1871.
- [10]. Keltjens, W.G. 1995. Magnesium uptake by Al-stressed maize plants with special emphasis on cation interactions at root exchange sites. *Plant and Soil* 171: 141–146.
- [11]. Keltjens, W. G. and Tan, K. Z. 1993. Interactions between aluminum, magnesium and calcium with different monocotyledonous and dicotyledonous plant-species. *Plant and Soil* 156: 485–488.
- [12]. Kochian, L. V. 1995. Cellular mechanisms of aluminum toxicity and resistance in plants. Ann. Rev. Plant Physiol. Plant Mol. Biol. 46: 237-260.
- [13]. Lidon, F.C., Azinheira, H.G. and Barreiro, M.G., 2000. Aluminium toxicity in maize: biomass production and nutrient uptake and translocation. *J. Plant Nutrition* 23: 151–160.
- [14]. Lukaszewski, K.M. and Blevins, D.G., 1996. Root growth inhibition in boron-deficient or aluminum-stressed squash may be a result of impaired ascorbate metabolism. *Plant Physiology* 112: 1135–1140.
- [15]. Ma, J. F. 2007. Syndrome of aluminum toxicity and diversity of aluminum resistance in higher plants. Int. Review. Cytol. 264: 225-252.
- [16]. Mossor-Pietraszewska, T. 2001. Effect of aluminium on plant growth and metabolism. Acta Biochim Pol. 3: 673 686.
- [17]. Olivares, E., Pena, E., Marcano, E., Mostacero, J., Aguiar, G., Benitez, M. and Rengifo, E. 2009. Aluminum accumulation and its relationship with mineral plant nutrients in 12 pteridophytes from Venezuela. *Environmental and Experimental Botany* 65: 132– 141.
- [18]. Richard A. F. 1998. Soil pH: Natural resources conservation service. Agronomy Tech. Note No. 150.2.
- [19]. Slaski, J.J., Zhang, G.C., Basu, U., Stephens, J.L. and Taylor, G.J. 1996. Aluminum resistance in wheat (*Triticum aestivum*) is associated with rapid, al-induced changes in activities of glucose-6-phosphate dehydrogenase and 6-pbosphogluconate dehydrogenase in root apices. *Physiol. Planta.* 98: 477-484.
- [20]. Taylor, G.J., Blamey, F. and Edwards, D. G. 1998. Antagonistic and synergistic interactions between aluminum and manganese on growth of *Vigna unguiculata* at low ionic strength. *Physiol. Planta*. 104: 183–194.
- [21]. Zheng, K., Pan, J. W., ye, L., Fu,Y., Peng, H. Z., Wan, B. Y., Gu, Q., Bian, H. W., Han, N., Wang, J. H., Kang, B., Pan, J. H., Shao, H. H., Wang, W. Z. and Zhu, M. Y. 2007. Programmed cell death involved aluminum toxicity in yeast elevated by antiapoptotic members with decreased calcium signals. *Plant Physiol*. 143: 38-49.