

Using Salivary Analysis to Confirm the Presence of Metabolites

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Abstract: The deficiencies of current diagnostic tools for Adderall are analyzed. A Java-applet was developed which analyzes delta E values from the Color Catcher app by TECHKON. This app is particularly useful for taking a picture of the color change from the reaction and then quantifying it through a delta E value. Further, the Java-applet analyzes delta E values, which are collected for the controls for each experimental sample. The values can be typed into the Java applet as inputs. A comparative study is made between several student saliva samples, against their respective controls. The applet compares metabolites in the experimental saliva against the metabolites in the respective control saliva, and finally provides meaningful conclusions. This applet helps in removing the possibility of any human error, and is thereby useful as an objective decision-making tool.

Keywords: Adderall, delta E, saliva, chromophore, Hydroxyamphetamine, b-endorphin, DOPAC

I. Introduction

The past and current research on diagnostic tools for Adderall, does not throw sufficient light and reliable conclusions about the patient. Primarily this is so due to human error which occurs during data collection and data analysis of urine samples. Furthermore, the instrument used in urine analysis, GC-MS consumes immense time and energy. In addition, the methods requiring urine samples need patients to be alone, when samples can be altered. Sputum samples are inconclusive since they analyze only one metabolite which may not even occur. Blood tests pose a safety hazard which requires a laboratory environment.

With the creation of an easy-to-use saliva drug test, the abuse of drugs would greatly decrease. Currently, the diagnostic tools for drug tests do not provide either reliable nor immediate conclusions about the patient. With a sputum drug test, the identification process for drug abusers would be more efficient. Furthermore, the methodology of analyzing delta E values using a Java-applet which compares experimental sample to the control can not only be used to diagnose Adderall in saliva, but also can be used to diagnose heroin, cocaine or other drugs found in saliva. If the right biomarkers are found, this methodology can be used to detect any drug in saliva. Before we go into the details of the developed Java applet, let us have a look at Adderall as a central nervous system stimulant.

1.1 CENTRAL NERVOUS SYSTEM STIMULANT: ADDERALL

Adderall is a powerful central nervous system stimulant, one of the most common commonly abused psychostimulant drugs. Adderall is a specific type of amphetamine. Amphetamine is a group of central nervous system stimulants known for their indirect effects on the central nervous system and peripheral nervous system. Adderall was first introduced as an effective method for treating the symptoms of narcolepsy and attention deficit hyperactivity disorder (ADHD). However, due to the fact Adderall is one of the few stimulants which are legal in America, it is also one of the most commonly abused drugs (Becker, 2015). In general, Adderall is commonly used to treat the symptoms of deficit hyperactivity disorder (ADHD), as shown in below Figure 01.



Figure 1: Adderall tablet (Rivers)

1.2 ATTENTION – DEFICIT/HYPERACTIVITY DISORDER (ADHD)

Current research suggests that ADHD is associated with functional impairments in some of the brain's neurotransmitter systems [xyz]. These functional impairments are usually distorted dopamine neurotransmission and norepinephrine neurotransmission involved with the brain's physiological responses to stress and panic. Psychostimulants such as amphetamine increase neurotransmitter activity in the brain, repairing the impaired dopamine neurotransmission and norepinephrine neurotransmission. These extr dopamine and norepinephrine neurotransmitters increase impulse control and attention span; therefore, they are consequently used as medication for ADHD, as shown below in Figure 02 [Olmez, 1988].

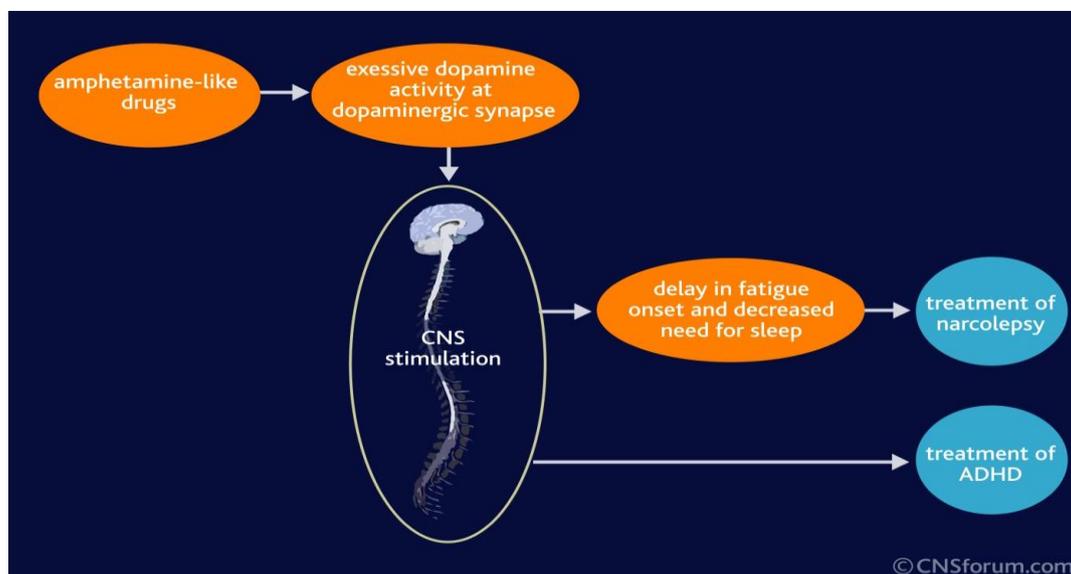


Figure 2: Diagram of how Adderall's effects counter the effects of ADHD and narcolepsy (Olmez, 1988)

1.3 SOME FACTS ABOUT ADDERALL ABUSE

Since amphetamines behave like dopamine, they also have the ability to bind to the same enzymes as dopamine, and can release euphoria. Furthermore, the increase in euphoria and expanded attention span, make Adderall one of the most commonly abused drugs among students. Abuse of amphetamine medication, such as Adderall, is a concern on college campuses. Students abuse Adderall for many reasons, however, it is mostly used for improving attention to do help improve grades. Some other common reasons to abuse Adderall include recreational use, and reducing hyperactivity [Olmez, 1988], show in the below Figure 03.

TABLE 1. Sample and Population Demographics, in Percentages

Characteristic	University (N = 11,897†)	Total sample (N = 1,025)	ADHD diagnosed students (n = 68; 6.6%)
Age (y)			
17	< 1	< 1	0
18	16	24	22
19	19	19	24
20	18	18	12
21	16	17	13
22–23	12	11	7
24+	19	11	22
Female	58	66	52
Year in school			
Freshman	22	31	32
Sophomore	22	20	19
Junior	20	19	16
Senior	20	21	25
Graduate	19	10	7

†From University of New Hampshire 2001 Fact Book. University of New Hampshire Office of Institutional Research. Available at: http://www.unh.edu/it/master_factbook_01PDF.pdf. Accessed July 15, 2004.

Figure 3: Table which demonstrates the number of students at the university of New Hampshire who use Adderall and those who actually need it. (Olmez, 1988)

The recent study collected data in patient students of people who used Ritalin, Adderall, Cylert, Dexedrine, Concerta, or any other stimulant medication. This was an online survey to be sent to the students of New Hampshire University. It was concluded that 95.4% of the students can be concluded that were on stimulant medication illegally.

This study emphasizes not only the college students but also the truck drivers who are also known to be common Adderall abusers. Amphetamine is the most commonly consumed drug by truck drivers. Urine samples of Mexican truck drivers were evaluated. In a recent study, 109 had amphetamine (90%) and 12 had methamphetamine (10%). The metabolite 4-hydroxyamphetamine was also tested for, and those which tested positive for it were quantified in the amphetamine positive group [Barceloux, 2012], as shown below Figure 04

Amphetamine in Urine Range (ng/mL)	Number of Individuals	Methamphetamine in Urine Range (ng/mL)	Number of Individuals	4-Hydroxyamphetamine in Urine Range (ng/mL)	Number of Individuals
1000 to 5000	19	1000 to 5000	1	0 to 500	26
5000 to 10,000	44	5000 to 10,000	4	500 to 1000	25
10,000 to 20,000	33	10,000 to 20,000	4	1000 to 2000	30
20,000 to 50,000	12	20,000 to 50,000	2	2000 to 5000	18
> 50,000	1	> 50,000	1	> 5000	10
Total	109	Total	12	Total	109

Figure 4: Table which displays the Amphetamine, Methamphetamine, and 4-Hydroxyamphetamine levels in the urine of Truck Drivers. (Barceloux, 2012)

1.4 PHARMACOKINETICS

Pharmacokinetics is the study of how Adderall is metabolized in the body. Adderall is primarily metabolized in the liver. To form 4-hydroxyamphetamine, amphetamine is either oxidized at the benzene ring to form alpha-hydroxy-amphetamine. Or, it is oxidized on the side chain of β carbons side to form norephedrine. Norephedrine and 4-hydroxyamphetamine are both metabolites of amphetamine and are both oxidized to form 4-hydroxy-norephedrine. Then the alpha-hydroxy-amphetamine goes through deamination to form phenylacetone. Phenylacetone then forms benzoic acid and its glucuronide and the glycine conjugate hippuric acid; however this is a common reaction of phenylacetone; it is not specific to amphetamine metabolism. CYP2D6, dopamine β -hydroxylase, 6-l-aminocaproic acid oxidase, butyrate-CoA ligase, and glycine N-acyltransferase are the enzymes that metabolize amphetamine [Chaing, 1986], as shown below.

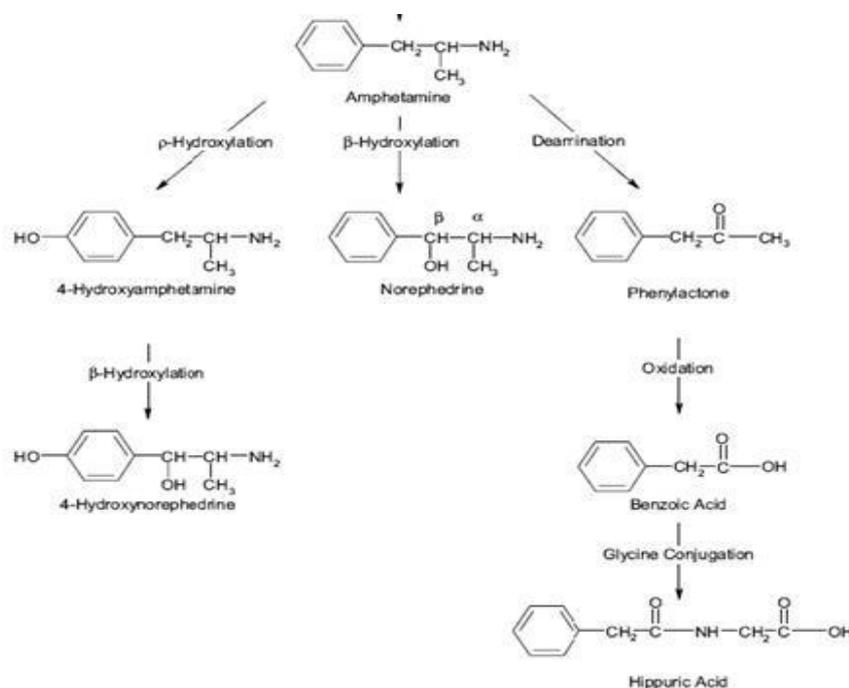


Figure 5: Metabolism of Adderall in body (Chaing, 1986)

The gastrointestinal pH determines the oral bioavailability of amphetamine. If amphetamine is well absorbed from the liver, then the oral bioavailability is over 75% for dextroamphetamine. Amphetamines are a weak base. The pKa is about 9–10. When the pH is more basic, more of the drug is in its lipid soluble free base form. This allows more of amphetamine to get absorbed through the lipid-rich cell membranes of the liver. It is realized that 15–40% of amphetamine in the bloodstream is bound to plasma proteins [Becker, 2015].

1.5 MAJOR EFFECTS OF AMPHETAMINE

The metabolites of amphetamine, *d*-amphetamine (*d*-AMPH) and *l*-amphetamine (*l*-AMPH), are reversible monoamine oxidase inhibitors. Reversible inhibitors of monoamine oxidase (RIMAs) are a class of drugs which are very selective of the enzyme monoamine oxidase (MAO) that they choose to reversibly inhibit. Monoamine oxidase type A (MAOA) inhibitors are an enzyme encoded by the MAOA gene. The human body has two neighboring gene family members that encode for mitochondrial enzymes. MAOA is one of the two. It catalyzes the oxidative deamination of amines; for example, serotonin, dopamine, and norepinephrine. Because of their reversibility and selectivity, RIMAs are safer than monoamine oxidase inhibitors (MAOIs) like phenelzine and tranylcypromine, as shown below

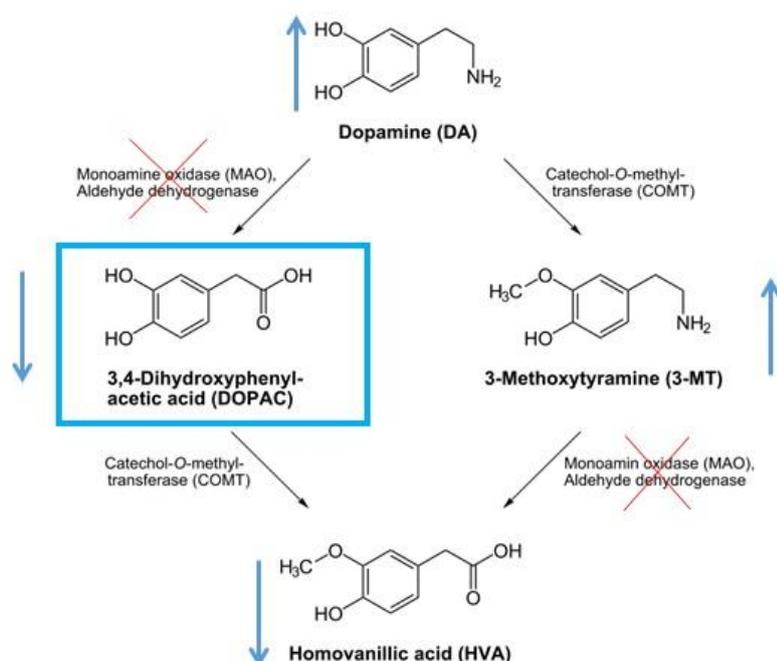


Figure 6: Diagram of how Dopamine levels are affected with Amphetamine

Some known molecules are known to increase the concentration of the ionized species of the amphetamine molecule. This increase in amphetamine's ionized species increases urinary excretion. These molecules also lower blood levels and efficacy of amphetamines. Some of the known molecules include ammonium chloride, sodium acid phosphate. (Gulovli, 1988). Amphetamines also inhibit adrenergic antagonist. Adrenergic antagonist is a pharmaceutical substance which inhibits catecholamines, a group of monoamines which have specific physical organic compound properties. These adrenergic antagonists inhibit catecholamines at the adrenergic receptors. Similar to most pharmacological receptor antagonists, the receptors' effect can only be seen when the receptor's effector is present.

Furthermore, amphetamines have the potential to enhance the activity of tricyclic sympathomimetic agents. When the metabolite-amphetamine is exposed to desipramine or protriptyline and possibly other tricyclics, it increases the concentration of *d*-amphetamine in the brain. Consequently, the cardiovascular system has the potential to be affected. (Gulovli, 1988)

1.6 CHROMOPHORES

Chromophores are compounds that are produced the color in dyes. They absorb electromagnetic radiation of different wavelengths, which depend on the energy of the electron clouds, to produce a color. Chromophores are structures with atoms joined in a sequence composed of alternating single and double bonds. Chromophore configurations often exist as multiple units, having conjugated double bonds, and are more effective when they do so. This is due to the interaction between the double bonds, which causes partial de-

localization of the electrons involved in the bonds. In this case, although specific atoms are involved in the bonds, the electrons are redistributed over a larger area than the specific atoms and also involve adjacent atoms that have double bonds (Rivers, 2015). The point of this is that conjugated systems have partially delocalized electrons, and the energy in these delocalized electrons can impact on the energy of the delocalized electrons of the parent aromatic compound by extending the number of electrons involved in the system and the energy needed to keep the whole system in place.

1.7 EXISTING TEST STRIPS & SAMPLE COLLECTION

The test device consists of a chromatographic absorbent device in which the drug or drug metabolites in the sample compete with a drug conjugate immobilized on a porous membrane support for the limited antibody sites. As the test sample flows up through the absorbent device, the labeled antibody-dye conjugate binds to the free drug in the specimen forming an antibody: antigen complex. This complex competes with immobilized antigen conjugate in the positive reaction zone and will not produce a magenta color band when the drug (SalivaConfirm), as shown in the below Figure 06.

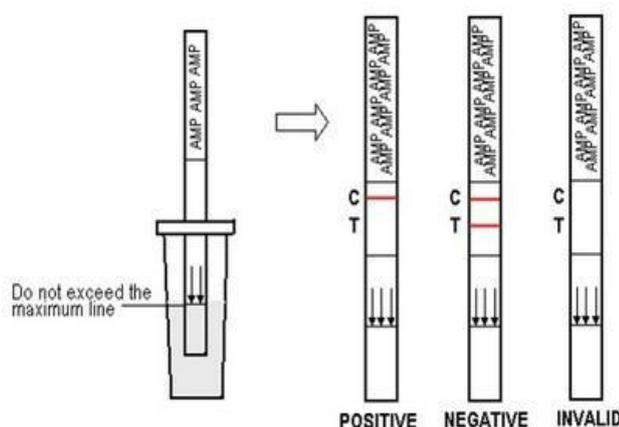


Figure 7: Illustration of current saliva test strips (SalivaConfirm)

It is also important to realize, that the ΔE value is primarily based on an algorithm originally developed by Richard Hunter in 1972 which is based on 3-dimensional Euclidean difference formulae (equation 1) that takes the root of the difference of the color coordinate squares based on the standard CIE LAB color axis [Fairchild, 2005]. If coordinate 1 is (x_1, y_1, z_1) and coordinate 2 is (x_2, y_2, z_2) , then the value of ΔE can be easily calculated using the below formula

$$\Delta E = \sqrt{(x_1 - x_2)^2 + (y_1 - y_2)^2 + (z_1 - z_2)^2}$$

This calculation of ΔE is widely used in printing and photography as it provides a common platform for costumers, print providers and suppliers to exchange their resources and requests. [Fairchild, 2005], as shown below

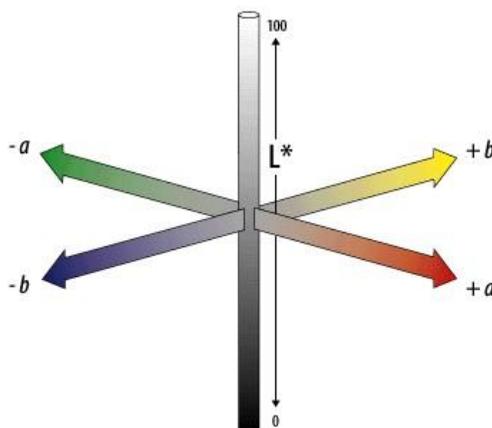


Figure 8: CIE LAB Color axis. ΔE is calculated from taking 2 points on this 3D coordinate system and then defining x, y, z according to the color axis as shown here. These axes are defined based on the simple theory that not two colors can be the same identical parts of red and green, blue and yellow, and black and white. (Fairchild, 2005)

The numerical values used in the formula come from the color-modelling system devised by Albert Henry Munsell. In his 1898 publication "A Color Notation" he describes color using a decimal system as opposed to adjectives. (Fairchild, 2005) His work remains a standard for colorimetry even today and has been adopted even in this numbering system by which ΔE was calculated.

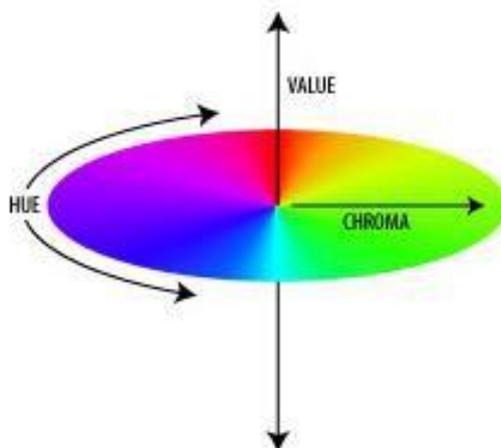


Figure 9: Overall model of Munsell. Munsell's model is based on a 3D sphere where the central slice (termed equator) carries a band of colors and the vertical axis running perpendicular to this slice is the gray scale with black and white at either pole. Moving outwards from the center towards the outside along the same gray point (same point on the vertical axis) provides different levels of saturation. (Fairchild, 2005)

1.8 UNDERLYING ENGINEERING PLAN

In this research we propose an engineering plan, detailed below, setting up fixed goals and certain procedures.

A. Engineering problem being addressed:

Currently no definitive tests for Adderall use in classrooms exist. Methods requiring urine samples need patients to be alone, whereas samples can be altered. Sputum samples are inconclusive since they analyze 1 metabolite that may not occur.

B. Engineering Goals:

The goal of this project is to engineer a methodology of analyzing ΔE values using a Java-applet that compares experimental samples to the control.

C. Detailed Description of the involved procedures:

Procedures:

Each color-changing molecule (b-endorphin, DOPAC, and Adderall metabolite) will be paired with its respective chromophore to create an invisible color change. When all three invisible distinct color changes happen, another color will be formed. This color will only be formed if all three colors are present, which will indicate Adderall is present in saliva. The Qualified Scientist will do the chromophore pairing in his lab.

Data Analysis: My saliva will be collected and tested by placing the chromophore test strip as a control. The Qualified Scientist will put the three Adderall metabolites (CYP-450 metabolite, 3,4-Dihydroxyphenylacetaldehyde, and b-endorphins) in the other sample of my saliva, and test the strip again.

Design Criteria and Secondary Engineering Goals: The test strip should only make a color if Adderall is present in saliva. Nothing else in saliva should influence that color change. Furthermore, my test strip's reactions should be independent of all other substances in saliva.

Testing: My saliva will be used as a control. Common elements of Adderall influenced saliva including CYP-450 metabolite, 3,4-Dihydroxyphenylacetaldehyde, and b-endorphins will be put into another sample of my saliva, in a lab environment.

II. Research Methodology

In this research, we have developed a research methodology which is more practical and significant, especially with reference to time effectiveness. The Color Catcher app by TECHKON© was used to capture a picture of the sputum color with the LED light turned on. The generated ΔE value was compared to the provided "normal" and "Adderall abuse" samples. Since the lighting and iPhone camera lens may be different, the normal and "Adderall abuse" samples are provided so the person collecting samples will be able to use their own

phonetogenerate thenormal and“Adderallabuse” controls underthe sameconditions as the samples being collected.

OncetheDeltaE values are collected for the controls,theycan betyped into the respectiveinputboxinthe Java-applet. Thesewillfunction as theparameters for howthestudent’s samples willbe evaluated. As the pictures of students samples aretaken usingthe app, thedeltaE valuecan betyped into the respective input area for collected samples as along with the corresponding user interface for the app created by the author is shown below:

The screenshot shows a web-based user interface for data entry. It features a 'Results' section with three sub-sections: 'Normal', 'User', and 'User'. Each sub-section contains three input fields labeled 'Plear enter delta E' (likely a typo for 'Please enter delta E') corresponding to 'Normal: Beta-Endorphin', 'Normal: Hydroxyamphetamine', and 'Normal: DOPAC'. Below these are three more input fields for 'User: Beta-Endorphin', 'User: Hydroxyamphetamine', and 'User: DOPAC'. At the bottom of the form is a 'Calculate' button.

Figure 10: User Interface for the app.

Digital recordingof thedata will protect the authenticityof thedata collected and enable immediate sharingof thedata for futurerecords. The applet willgenerate aconclusiveresultof whetherthe student maybeapotential Adderalluser ornor.Iftheresultsuggest that thestudent’s color changeswereout of thenormal range, then the next step would beto placethem in a monitored settingfor24 hours wheretheywillbe allowed to eat, drink and rest normallybut then tested again just asbefore.If theresults come to besimilar to thefirst timethen it showsthat the patient is not usingAdderall, but if therearedistinctive different color changes providing different DeltaE values,then it maysuggest thesefluxuations maybewithdrawal effects of not havingthe drugforafull24 hours.

III. Experimental Results

(ug/mL)	b-endo	4-HA	DOPAC	
Color	Orange	Blue	Green	
1	48.5	0.35	0.98	ADD
2	55.3	0.7	1.1	
3	49.2	0.23	0.87	
4	48.3	0.65	0.96	
5	55.7	0.34	0.99	
6	56.4	0.21	0.92	
7	47.9	0.57	0.95	
8	54.8	0.76	0.9	
9	49.8	0.11	0.92	
10	48.4	0.81	0.88	
Average	51.43	0.473	0.947	
SD	3.6043338	0.25408	0.0671731	
(ug/mL)				
Color	not Orange	Not Blue	Not Green	
11	44.3	0	1.33	CLEAN
12	50.2	0	1.35	
13	47.4	0	1.29	
14	45.8	0	1.15	
15	52.2	0.01	1.17	
16	49.9	0	1.19	
17	45.5	0.04	1.05	
18	52.3	0	1.07	
19	51.1	0	1.51	
20	44	0	1.42	
Average	48.27	0.005	1.253	
SD	3.2421015	0.012693	0.1515879	

Figure 11: Table thattakes noteofthe PatientNumberandwhatitsdelta Eis forHydroxyamphetamine, b-endorphin,andDOPAC.

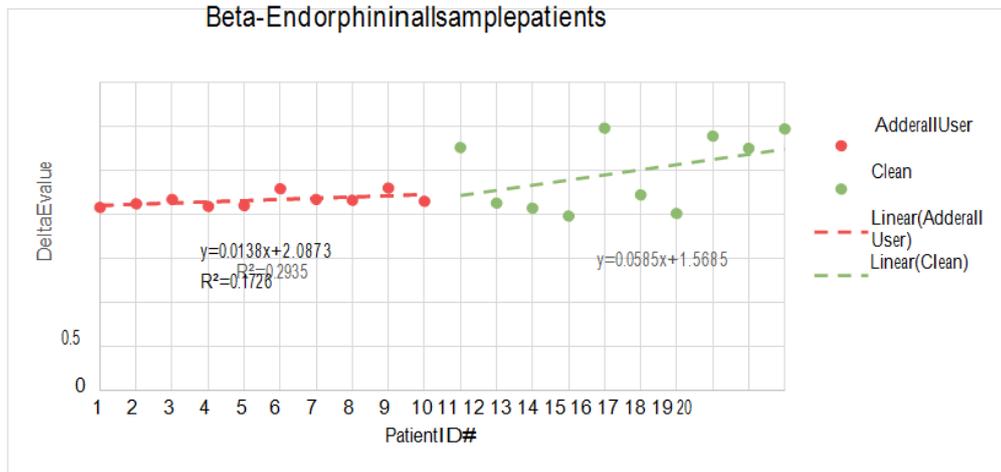


Figure 12: Graph of Delta Values of Beta-endorphin levels in the saliva of Patients

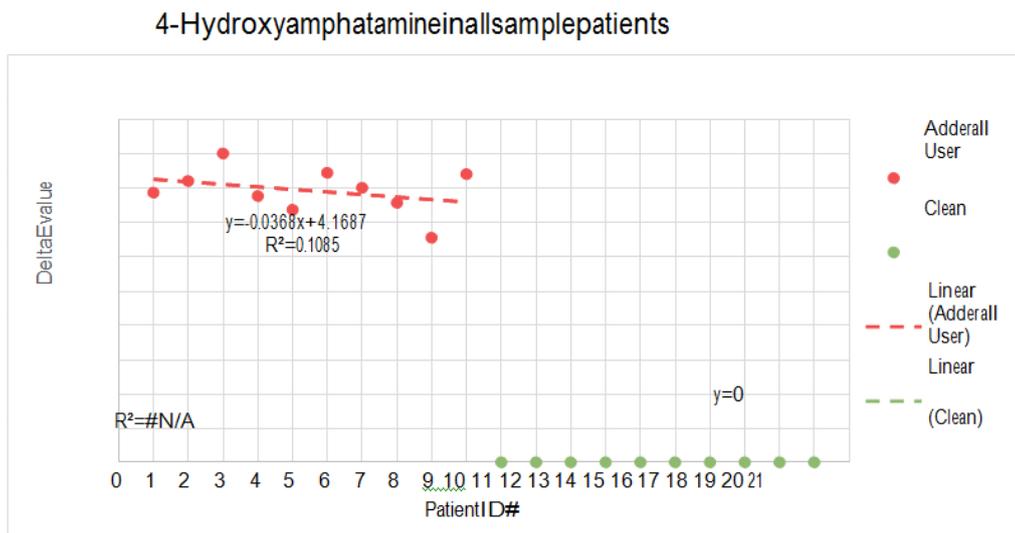


Figure 13: Graph of Delta Values of 4-Hydroxyamphetamine levels in the saliva of Patients.

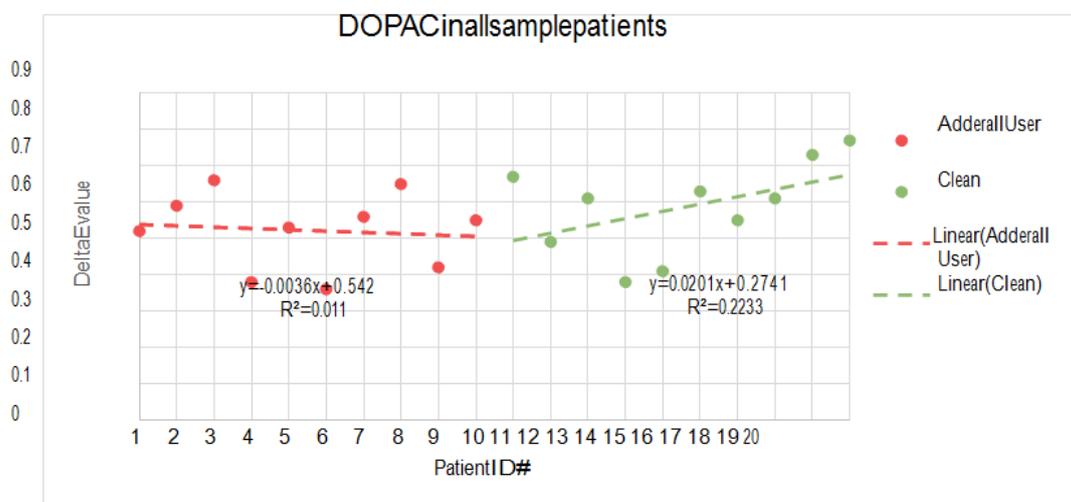


Figure 14: Graph of Delta Values of DOPAC levels in the saliva of Patients.

IV. Research Experimental Analysis

A particular research experimental analysis is detailed below to meet the goal of this project. It is basically an engineered methodology of analyzing delta E values using a Java-applet that compares experimental

sample to the control, and have used Adderall as an example to explain my methodology.

The ΔE values from all sample patients were used to generate three separate graphs for each of the three metabolites: Beta-endorphins (b-endo), 4-Hydroxyamphetamine (4-HA), and DOPAC. By plotting Adderall users as well as clean patients on the same graph, it becomes visually evident how differently the chromophores react to their sputum metabolites. The R^2 values for all of the graphs depict a poor linear regression. This, in combination to a very slight slope apparent on all the scatter plots, points to the fact that there is no relationship between the patient ID #'s and their respective metabolite values. When these ΔE values are inputted into the JAVA applet, the fact that they are individual values not dependent on any other variable enables the algorithm to produce a conclusive and definitive statement about the metabolites present.

The trends between the sputum metabolite plot and the ΔE plot are trending very similarly, confirming the strong correlation between the collected ΔE values from the color changing reaction in the sputum and the actual metabolite content in the sputum. The JAVA applet enables this easily administered test outside a standard laboratory setting to be helpful in definitively identifying students who might be taking Adderall without a prescription.

The standard deviation (SD) reveals that the variance of the data values collected was low and the overall procedure of data collection was reliable. In the sample serum studies, the SD was smallest in the 4-HA and DOPAC data values for both Adderall abusing patients as well as clean patients. This shows that the data collection method of the b-endo was not as reliable as the data collection method of the other 2 metabolites. This finding is supported by prior literature because beta-endorphins are found in everyone at a constant amount. But depending on the lifestyle factors of the individual: athletic nature, prone to depression, and of course use of recreational stimulants, the amount of b-endo may be slightly higher or lower. The overall serum average b-endo for sample clean patients was 48.27 ug/mL with an average $\Delta E = 2.476$, indicating a light orange color. The overall serum average b-endo for Adderall users was 51.43 ug/mL with an average $\Delta E = 2.163$, indicating a bright orange color. The other two metabolites also demonstrate a similar trend between serum quantities and ΔE from the color. The fact that there was a distinguishable difference visually and quantitatively points to the strong predictive nature of using ΔE as an analysis tool for studying the presence of metabolites in sputum. The ΔE value is nothing more than the difference between the darkest shade and the lightest shade of the snaptaken, in the case of poor lighting, there will be a difference that is erroneously small which can be misinterpreted. The ΔE value is being used as the variable input into the JAVA applet. This is precisely why the success of the JAVA applet is highly dependent on the lighting in the room where the app is used to be as close to sunlight (broad spectrum) as possible.

V. Useful Computational Results

When Adderall users as well as clean patients on the same graph, trends depicting a poor linear regression and a very slight slope became apparent on all plots made. This demonstrates the independent nature of the data collected for each of the 3 metabolites. These ΔE values are inputted into the JAVA applet, their independent nature enables the algorithm to produce a conclusive and definitive statement about the metabolites present.

The JAVA applet analyzes the ΔE values, obtained via chromophore-metabolite reaction in the sputum, by comparing them to controls. By using ΔE values to quantitatively measure presence of molecules, color detection technology can move further from an opinioned personal belief to a more tangible measurement tool that can revolutionize the way we share data. The assignment of numerical values to color enables programs like the JAVA applet to take these inputs and generate a conclusive result about the data. This will take the responsibility of decision making from the hands of a tired lab technician, into the objective algorithmic fingers of IT. Medical imaging, tissue cultures, heat measurement studies and many more color-based assays used today will be able to be better communicated and thereby improve inter- and intra-disciplinary collaboration.

VI. Conclusions And Future Possibilities

The methodology of analyzing ΔE values using a Java applet which compares experimental sample to the control can not only be used to diagnose Adderall in saliva, but also can be used to diagnose heroin, cocaine or other drugs in saliva. If the right biomarkers are found this methodology can be used to detect any drug in saliva.

The next step for the saliva test would be to test out the chromophore-molecule reactions in a lab, and to engineer the physical saliva test strip.

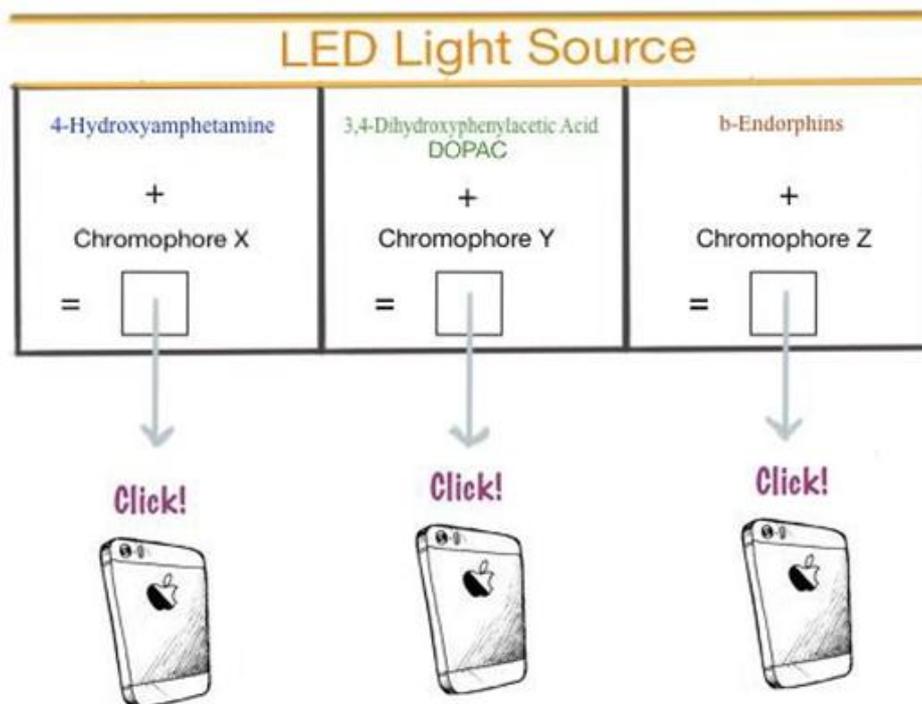


Figure 15: Model of the actual saliva test strip that has been engineered using the methodology developed in this paper

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Appendices

```

import java.awt.*;
import java.awt.event.*;
import java.applet.Applet;

import javax.swing.JTextField;

public class Gibbs extends Applet implements ActionListener
{
    Label ALabel = new Label("Alert Message");
    Button calculate = new Button("Calculate");
    Label HLabel = new Label("Enthalpy, dH [Joules]");
    TextField enthalpy = new TextField("Please enter enthalpy", 10);
    Label SLabel = new Label("Entropy, dS [Joules]");
    TextField entropy = new TextField("Please enter entropy", 10);
    Label TLabel = new Label("Temperature °K");
    TextField alert = new TextField("Hello!");
}
    
```

```
TextFieldtemperature=newTextField("PleaseenterTemprature",10);
LabelGLabel=newLabel("GibbsFreeEnergy,dG[Joules]");
TextFieldenergy=newTextField("0",10); LabelSpLabel=newLabel("Spontaneity");
TextFieldspontaneity=newTextField("Equilibrium",25); Panelp=newPanel();
privatestaticbooleanvalid=true;

publicvoidinit()
{
setBackground(Color.white); p.setLayout(newGridLayout(4,3,3,3)); p.add(ALabel);
p.add(Alert);
p.add(TLabel);
p.add(temperature); p.add(SLabel); p.add(entropy); p.add(HLabel); p.add(enthalpy); p.add(GLabel);
p.add(energy); p.add(SpLabel); p.add(spontaneity); p.add(calculate);
calculate.addActionListener(this);
add(p); Alert.setEditable(false); energy.setEditable(false);
}

intgetValue(Stringstr)
{
intnumber;
try
{
number=Integer.parseInt(str);

valid=true;
}
catch(NumberFormatException)
{
number=0;
valid=false;
}
returnnumber;
}

publicvoidactionPerformed(ActionEventevent)
{
inth=0,s=0,t=0,g=0;

StringHstr,Sstr,Tstr,Gstr,Spstr;

inti=0;
while(i<3)
{

Hstr=enthalpy.getText(); h=getValue(Hstr); if(!valid)break;
i++;

Sstr=entropy.getText(); s=getValue(Sstr); if(!valid)break;
i++;

Tstr=temperature.getText();
t=getValue(Tstr); if(!valid)break; i++;
}

switch(i)
{
case0:
```

```
{
Alert.setText("InvalidEnthalpyInput!"); Alert.setEditable(true);
spontaneity.setText("CannotBeDetermined!");
energy.setText("");
break;}

case1:
{
Alert.setText("InvalidEntropyInput."); Alert.setEditable(true);
spontaneity.setText("CannotBeDetermined!");
energy.setText("");
break;}

case2:
{
Alert.setText("Invalidtemperature."); Alert.setEditable(true);
spontaneity.setText("CannotBeDetermined!");

energy.setText("");
break;}

default:
{g=h-(t*s);
if(g==0)
{
spontaneity.setText("Equilibrium");
}

elseif(g>0)
{

happen");

happen");

}
else
{
}
spontaneity.setText("ReactionisNonSpontaneous(Cannot
spontaneity.setText("ReactionisSpontaneous(Can

Gstr=Integer.toString(g); energy.setEditable(true); energy.setText(Gstr);
}

} //switch
} //ActionPerformed
} //classGibbs
Figure 16:Code forDelta EJava Applet
```

Acknowledgements

The author wishes to thank several mentors who assisted in various aspects of this project. Dr. Gerardo Gonzalez provided ongoing guidance and support in both experiment design and execution. The author's parents willingly contributed both funding and weekend time in providing transportation and help with construction of the essential design.