Antioxidant Activity Of Two Different Extracts From Doum(*Hyphaenethebaica*) Fruits

*Lamiaa A. Gharb¹, Laith Z.Fadhel²

1,2(Department of biology, College of science/University of Baghdad, Iraq) *Corresponding Author: Lamiaa A. Gharb

Abstract: In the present study we have investigated antioxidant activity ofdoum(Hyphaenethebaica) fruit by using soxhlet apparatus for 9 hours to each of ethanol and ethyl acetate solvents. The antioxidant activity was done by using(DPPH) 2, 2 - diphenyl, 1- picrylhydrazyland (FRAP) ferric reducing antioxidant power assays.(BHT)butylatedhydroxytoluene was used as control. The results show that the scavenging effects of both extracts from our plant on DPPH radicals increased by increasing theconcentration. Ethyl acetate extract of the H. thebaica showed strong DPPH scavenging activity at concentrations (400, 600, 800 μ g/ml)more than ethanolic extract and BHT which were 70.20, 88.70 and 95.80% respectively. Ethyl acetate extract showed highest FRAP scavenging activity at concentration 600 μ g/mlwhich was 92.70 % more than BHT and ethanolic extract. The lower IC50 indicates a stronger free radical inhibition, However the IC50 of ethyl acetate extract was (229.383 and 205.507) in both DPPH and FRAP assays respectively. The results also revealed that doum fruits can be used as a natural antioxidants as well as the possibility of using this plant as food additives.

Date of Submission: 29-06-2019

Date of acceptance: 15-07-2019

I. Introduction

Herbal medicine is still the most common source for Human health careof about 65-80% of the world's population, because of better cultural acceptability, better compatibility with the human body and fewer side effects. Roots, flowers, bark, leaves, fruits, seeds and stem can all be constituents of herbal remedies. The medicinal values of these plants lie in their phytochemical components which produce definite physiological actions on the human body. The most important of these components are alkaloids, tannins, flavonoid and phenolics compounds¹. In present day the studies focus on natural antioxidants especially on plant phenolics^{2,3}. which act as an important free radicals removal and prevention disease, therefore the interest in plant products and extracts as a source of antioxidants is growing worldwide^{4,5}. Doum palm (*HyphaenethebaicaL.*) is a desert palm belonging to the family of Arecaceae. It is widespread in the sub-Saharan Africa, west India and tends to grow in areas where groundwater is present and is found along the Nile River in Egypt and Sudan. It is registered as one of the beneficial plants of the world^{6,7.} Various studies have revealed the fact that the doum fruit contain high levels of essential minerals such as potassium, sodium, calcium, magnesium, and phosphorus. As well as, Doum fruit contains B-complexvitamins, carbohydrates, and dietary fiber, which is essential for good nutrition^{7,8}. Numerous studies have emphasized that the doum fruit extracts contain high levels of phenols and flavonoids, and possess significant antioxidantand antimicrobial activities^{7,9.} Previous studies on doum had focused on the fruit because, besides its nutritional value, the fruit drink brewed from hot water infusion of the dried fruit pulp is widely consumed as a health tonic and has been valued in the Turkana region of Kenya, for its many anecdotal medicinal properties for centuries^{10,11}. The water extract of doum fruits can reduce hyperlipidaemia in nephritic syndrome and leads to decrease the risk of glomerulosclerosis and atherosclerosis and consequently the natural, safe and nontoxic H.thebaica fruit could be of great merit for use as hypolipidaemic drug as found by¹². This extract also is used in the treatment of bilharziasis, haematuria, and bleeding especially after child birth and as haematinic agent ^{13,14}. According to the previous studies, few scientific evaluations were done concerning the characterization of alcoholic doum fruit (*H.thebaica*) extracts.

Plant materials and chemicals

II. Material and Methods

Doum fruits were purchased from the local markets in Saudi Arabia. Chemicals were obtained from Sigma Chemicals Co. (USA).

Samples preparation and extraction

The fruits were crushed and grinded. Twenty grams of fruits powder were extracted with two different solvents(Ethanol and Ethyl- acetate) by using soxhlet apparatus for 9 hours. The extract was evaporated by using rotary evaporator and then the extract was stored at 4C prior to use^{15.}

The 2,2-diphenyl-1-picrylhydrazyl (DPPH) assay

2ml of each sample at different concentrations(200,400.600,800,1000 µg/ml) were separately added to 1ml solution of DPPH radical in methanol. The mixture was shaken and allowed to stand for 30 min of dark place. Then the absorbance of resulting solution (yellow color) was measured at 517 nm with spectrophotometer

Inhibition of free radical DPPH as percentage I% was collected as follow :

 $I \% = 100 \times \frac{A \text{ Blank} - A \text{ sample}}{A \text{ Blank} - A \text{ sample}}$ A Blank

A blank =Absorbance of control (containing all reagents except the test compound) and A sample is absorbance of test compound. IC50 value µg/ml is the effective concentration of which DPPH radical are scavenged 50%, it was calculated by using Excel programme depended on the logarithm(Log.) of each concentration. Butylatedhydroxytoluene (BHT) was used as control^{16.}

The FRAP (ferric reducing antioxidant power) method.

Various concentrations of the extracts (µg/ml) in distilled water were mixed with phosphate buffer (2.5 mL, 0.2 M, pH 6.6) and 1% of potassium ferricyanide water solution (2.5 mL, K3[Fe(CN)6]). The mixture was incubated at 50 C for 20 min. Aliquots of trichloro acetic acid (2.5 mL, 10%) were added to the mixture which was then centrifuged at 3000 rpm for 10 min . the supernatant (2.5mL) was mixed with distilled water (2.5 mL) and a freshly prepared FeCl3 solution (0.5 mL, 0.1%). The absorbance was measured at 700 nm, the reducing power of the tested samples increased with the absorbance values. BHT was used as a positive control. The reducing power of the (doum fruits extracts) were determined according to the method of ¹

III. Results and Discussion

The results in Table no (1) show that the antioxidant activity increased by increasing the concentrations. The activity of doum ethyl acetate extract was higher than ethanolic extract in the two different assays. At the concentration (400,600,800µg/ml), the highest DPPH activity was observed with theethyl acetate extract as compared with control.

	BHT		DoumEthanolic extract		DoumEthyl acetate extract	
Concentrations µg/ml	I % - DPPH	I % - FRAP	I % - DPPH	I% - FRAP	I % - DPPH	I % - FRAP
200	46.40	48.70	20.30	42.90	43.40	48.50
400	66.30	81.20	40.70	71.20	70.20	80.40
600	81.40	89.70	60.20	78.90	88.70	92.70
800	93.00	95.20	71.20	88.40	95.80	95.20
1000	99.20	100.00	95.30	90.20	97.40	99.50

Table no 1: DPPH and FRAP scavenging activities of two different extracts from Hypheanthebiacafruits

The lower IC50(the half maximal inhibitory concentration) indicates a stronger free radical inhibition (strong free radical inhibitors are active at low concentrations)¹⁸. The IC50 of doum extracts for the DPPH and FRAP assays represented in table no 2, for the ethanolic extract the IC50 was (462.190 and 211.072 µg/ml) respectively as compared with BHT(223.582 and 205.623 μ g/ml). The results also revealed that the IC50 of ethyl acetate extract was (229.383 and 205.507) in both DPPH and FRAP assays respectively .

Table no 2: The IC50 of doum fruit extracts and BHT in DPPH and FRAP assays						
Assay	IC50/BHT	IC50 /DoumEthanolic extract	IC50 /Doum Ethyl acetate extract			
DPPH	223.582	462.190	229.383			
FRAP	205.623	211.072	205.507			

. . . · · IDUTE: DODU

Table no(3) shows that in DPPH and FRAP assays there was a significant difference at p < 0.05 exist between the (200 and 400 µg/ml) concentrations of ethanolic extract with each of control and ethyl acetate extract. However, there was no significance difference at p > 0.05 appeared between the ethyl acetate extract and control in FRAP assay in these two concentrations in addition to the 200 µg/ml concentration in DPPH assay. On the other hands, ethyl acetate extract of doum fruit appeared to be higher than control and ethanolic extract in the concentration 400 μ g/ml in DPPPH test. Figure(1)

HypneanineDiacaffuits							
	DPPH Test			FRAP Test			
Concentration Treatments	200 µg/ml	400 µg/ml	200 µg/ml	400 μg/ml			
ВНТ	46.400 a ± 1.039	66.300 b ± 0.924	48.700 a ± 1.212	81.200 a ± 1.039			
Ethanolic	20.300 b ± 0.808	40.700 c ± 0.924	42.900 b ± 1.097	71.200 b ± 0.866			
Ethyl acetate	43.400 a ± 1.097	70.200 a ± 1.039	48.500 a ± 0.693	80.400 a ± 0.981			
LSD P ≤ 0.05	3.424	3.335	3.548	3.339			

 Table no 3: DPPH and FRAP scavenging activities of(200 and 400µg/ml) in two different extracts from

 Hypheanthebiacafruits

 $Small \ letter \ s \ indicate \ to \ comparison \ in \ column \ , \ similar \ letters \ are \ non-significantly \ differences \\ between means \ at \ (p \le 0.05) \ using \ LSD \ test$

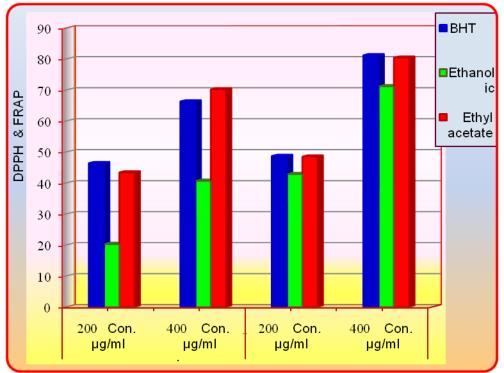


Figure 1: Antioxidant activity of (200 and 400 µg/ml) in two different extracts from Hypheanthebiacafruits

IV. Discussion

The proton radical scavenging action is known as an important mechanism of antioxidants. DPPH is usually used as a substrate to evaluate the antioxidative activity of natural antioxidants because it is a stable free radical and accepts an electron or hydrogen radical to become a stable diamagnetic molecule^{19.} The effect of antioxidants on DPPHradical scavenging was thought to result from their hydrogen donating ability²⁰. The decrease in absorbance of the DPPH radical caused by reacting between antioxidant molecules and the radical, progresses, which results in the scavenging of the radical by hydrogen donation. The scavenging effects of both extracts (Ethanol and ethylacetate) from our plant (*H. thebaica*) on DPPH radical decreased in the order of BHT, ethyl acetate extract ,Ethanolic extracts and standard on the DPPH radical decreased in the order of BHT, ethyl acetate extract ,Ethyl acetate extract of the *H. thebaica* shown strong DPPH scavenging activity at concentrations 400, 600, 800 µg/mlmore than ethanolic extract and BHT.These results indicated that ethyl acetate extract of doum fruithas a noticeable effect on scavengingfree radicals.The FRAP assaydepend on the reduction of a ferric tripyridyltriazinecomplex to ferrous $-(2,4,6-tripyridyl-s-triazine)_2$ i.e.: ferric (III) colorless will change to ferrous (II) blue color. The absorption readings are related to the reducing

DOI: 10.9790/3008-1404010104

power are related to electron-donating antioxidants present in the test compound. The scavenging effects of both extracts (Ethanol and ethyl acetate) on FRAP increased with concentration (Fig. 1). Ethyl acetate extract of the *H. thebaica* shown strong FRAP scavenging activity at concentrations 400 μ g/mlmore than ethanolic extract. These results indicated that the ethyl acetate extract of *H. thebaica* as a noticeable effect on scavengingfree radicals. Phenolic compounds of the *H. thebaica* extracts are probably involved in their antiradical activity²¹. Although the activity of ethyl acetate extract of doum fruit in some concentration is relatively more than of BHT, the extract may be viable source of bioactive compounds with better activities after fractionation.

V. Conclusion

Since the presence of free radicals, especially their increased production, appears to be a feature of most, if not all human diseases. The modern research is directed towards "Natural antioxidants" from the herbal plants due to safe therapeutic. The findings of this study support the view that some medicinal plants like doum fruits are promising sources of natural antioxidants as well as to the possibility of using this plant as food additives.

References

- [1]. Shariff ZU. Modern Herbal Therapy for Common Ailments. *Nature Pharmacy Series* Vol.1, SpectrumBooks Ltd., Ibadan, Nigeria in Association with Safari Books. 2001. pp. 9-84.
- [2]. EldahshanO., Ayoub N., Singab A., Al-Azizi M. Potential superoxide anion radical scavenging activity of doum palm(*Hyphaenthebaica* L.) Leaves Extract. Rec. Nat. Prod.2008. 2, 83–93.
- [3]. Eldahshan O., Ayoub N., Singab A., Al-Azizi M. Potential antioxidant phenolic metabolites from doum palm leaves. Afr. J. Pharm. Pharmacol.2009. 3, 158–164.
- [4]. Hsu B., Coupar I.M., Ng K. Antioxidant activity of hot water extract from the fruit of the doum palm, *Hyphaenethebaica*. Food Chem.2006. 98, 317–328.
- [5]. Langley-Evans S.C. Antioxidant potential of green and black tea determined using the ferric reducing power (FRAP) assay. Int. J. Food Sci. Nutr.2000. 51, 181–188.
- [6]. Fletcher, R. Listing of useful plants of the world. Australian New Crops. 1997. http://www.newcrops.uq.edu.au/listing/hyphaenethebaica
- [7]. Aboshora W. Effect of extraction method and solvent power on polyphenol and flavonoid levels in *Hyphaenethebaica* L mart (Arecaceae)(Doum) fruit, and its antioxidant and antibacterial activities. Tropical Journal of Pharmaceutical Research, 2014. 13(12): p. 2057-2063.
- [8]. Aboshora, W. Compositional and structural analysis of epicarp, flesh and pitted sample of Doum fruit (Hyphaenethebaica L.). International Food Research Journal. 2017.
- [9]. Mohamed A. A., Khalil A. A., and H. E. El-Beltagi, Antioxidant and antimicrobial properties of kaffmaryam(*Anastaticahierochuntica*) and doum palm (Hyphaenethebaica). Grasas Y Aceites, 2010. 61(1): p. 67-75.
- [10]. Martin F.W., 1999. Palm for stable foods. In: Elevitch, C. (Ed.), Multipurpose Palms You Can Grow. Also available electronically http://www.agroforestry.net/pubs/palmbk/Chapter4.html>.
- [11]. Cook J.A., Vander Jagt, D.J., Pastuszyn, A., Mounkaila, G., Glew, R. S., Millison, M. Nutritional and chemical composition of 13 wild plant foods of Niger. J. Food. Compos. Anal.2000. 13, 83–92.
- [12]. Habib, D.F., Michael, H.N., Salib, J.Y., Ahmed, N.M., Agaibyi, M. H.Hypolipidemic efficacy of hyphaenethebaica (doum) in experimental nephrotic syndrome. IJ.P.2014. 4, 28–34.
- [13]. Adaya, A.L., Bitrus, H.H., Fanjoji, M. Eaton, Gambo, D., 1977. Hidden harvest project in research series. Compiled by IIED and HNNCP. pp. 14–27; 47–53.
- [14]. Burkill, H.M., 1997. The useful plants of West Tropical Africa, vol. 4, second ed. Royal Botanical Garden, Kew, pp. 371–373.
- [15]. Dosumu O.O., Nwosu F.O. and NwoguC.D.Antimicrobial studies and phytochemical screening of extracts of *Hyphaenethebaica*(Linn) Mart Fruits.International journal of Tropical Medicine.2006.1(4):186-189.
- [16]. Sanchez-moreno C., Larrauri J.A. and saura-calixto F. A procedure to measure the antiradical efficiency of plant extracts. Journal of the Science of Food and Agriculture .1998.76(2): 270-276.
- [17]. Oyaizu, M. Studies one products of browning reaction prepared from glucose amine. Japanese Journal of Nutrition.1988. 44 (6): 307-315.
- [18]. Ghasemzadeh A., Jaafar H. Z. E., Ashkani S., A. Rahmat, Juraimi A. S., Puteh A. and Mohamed M. T. M.Variation in secondary metabolite production as well as antioxidant and antibacterial activities of *Zingiberzerumbet* (L.) at different stages of growth. BMC Complementary and Alternative Medicine.2016.16:104
- [19]. Soares JR, Dinis TCP, Cunha AP, Almeida LM. Antioxidant activities of some extracts of Thymus. *Free Radical Research*. 1997. **26**, 469-478.
- [20]. Shimada K, Fujikawa K, Yahara K, Nakamura T. 1992. Antioxidative properties of xanthone on the autooxidation of soybean in cylcodextrin emulsion. *Journal of Agricultural and Food Chemistry* **40**, 945-948.
- [21]. Hsu B, Coupar IM, Ng K. 2006. Antioxidant activity of hot water extract from the fruit of the Doumpalm, Hyphaenethebaica. *Food Chemistry* 98, 317-328.

Lamiaa A. Gharb" Antioxidant Activity Of Two Different Extracts From Doum(Hyphaenethebaica) Fruits . " IOSR Journal of Pharmacy and Biological Sciences (IOSR-JPBS) 14.4 (2019): 01-04.

DOI: 10.9790/3008-1404010104