Simultaneous Determination Of Hyoscine Butyl Bromide and Dipyrone in Their Binary Mixture By RP-HPLC Method.

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Abstract :RP-HPLC chromatographic method was developed for determination of Hyoscine Butyl Bromide (HBB) and Dipyrone (DIP). In this method, HBB and DIP were separated by using C18 (25 cm×4.6 mm i.d. 5μ m particle size) column as a stationary phase and using water: methanol (50:50) v/v pH adjusted to 7.0 with triflouroacetic acid and triethylamine) as a mobile phase, maintaining the flow rate at 1.0 mL min⁻¹ with UV detection at 210 nm. The proposed method was successfully applied for determination of HBB and DIP in pure form and in their pharmaceutical formulations.

Keywords: RP-HPL, Chromatography, Hyoscine Butyl Bromide, Dipyrone

I. Introduction

Hyoscine butyl bromide is a quaternary ammonium anticholinergic agent. It has antispasmodic action on the smooth muscles of the gastrointestinal, biliary, and urinary tracts [1].

Dipyrone is pyrazolone derivative with a strong analgesic, antipyretic, very weak antiinflammatory effect and spasmolytic activity [1]. The structural formulas of HBB and DIP are shown in Figure 1.

There are many reports for the determination of HBB and DIP either separately or in combination with other drugs including spectrophotometric methods [2-6], chromatographic methods [7-9], electrochemical methods [10-13], Capillary electrophoresis methods [14,15] and titrimetric method [16-18].

Few methods have been mentioned for analysis of HBB and DIP in binary mixture. Erk and Onur [19] developed two spectrophotometric methods for the simultaneous determination of DIP and HBB in their binary mixture. The first method depend on derivative spectrophotometry, the determination of these drugs were performed by measuring the dA/d λ values at 291.8 nm and 219.8 nm in the first derivative spectra of the mixture for DIP and HBB respectively. In the second method, the determination of these compounds in mixture was realized by precipitating HBB with ammonium reineckate at pH 6.0 selectively and reading the absorbance of the solution of the precipitate in acetone at 532.2 nm for HBB and by measuring the dA/d λ values at 306.2 nm in the first derivative spectra of the remaining solution for DIP.

Up to our knowledge, no chromatographic methods have been published for determination of HBB and DIP mixture.

Therefore, the objective of this work is to develop sensitive, selective and reproducible RP-HPLC method for simultaneous determination of HBB and DIP for routine quality control analysis of these drugs either in bulk powder or in pharmaceutical formulations. Chromatographic methods are well known for providing high selectivity and sensitivity when used for determination of pharmaceutical drugs.

II. Experimental

II.1. Apparatus

1-Shimadzu Class - LC 10 AD Liquid Chromatography supplied with Shimadzu SPD - 10 A UV - VIS Detector (Shimadzu Corporation ,Japan).Phenomenex C18 (25 cm x 4.6 mm i.d, 5 μm particle size) column was used as a stationary phase for HPLC determinations (USA).

2-Sonix TV ss-series ultrasonicator (USA).

II.2. Materials

II.2.1. Pure samples

Dipyrone (DIP) and hyoscine N butyl bromide (HBB) were kindly supplied by CID Co. Chemical Industries Development, Giza, Egypt. Their purity was found to be 100.22 ± 0.544 and 99.21 ± 1.012 , respectively, according to the company analysis certificate (HPLC).

II.2.2. Market samples

1- Buscopan compositum[®] tablets (Batch No 115815T) labeled to contain 250 mg of (**DIP**) and 10 mg of (**HBB**), CID Co. Chemical Industries Development, Giza, Egypt.

2-Buscopan compositum[®] ampoules (Batch No 116551A), each 5 mL ampoule labeled to contain 2500 mg of (**DIP**) and 20 mg of (**HBB**), CID Co. Chemical Industries Development, Giza, Egypt.

II.2.3. Reagents

All reagents and chemicals used were of analytical grade and were used without further purification 1- Methanol HPLC grade and triethylamine (Sigma Aldrich, Germany).

2-Deionised water (SEDICO pharmaceutical Co., 6th October City, Egypt).

3-Orthophosphoric acid and glacial acetic acid from (EL - NASR Pharmaceutical Chemicals Co., Abu - Zabaal, Cairo, Egypt).

4- Trifluoroacetic acid from Spectrochem, India.

II.3. Preparation of standard solutions

A-Dipyrone (DIP) and Hyoscine N butyl bromide (HBB) stock standard solutions (1 mg.mL⁻¹).

Weigh accurately 0.1 gram of each drug into a two separate 100-mL volumetric flask, 50 mL methanol was added, shake to dissolve then complete the volume to the mark with methanol.

B- Dipyrone (DIP) and Hyoscine N butyl bromide (HBB) working standard solutions (100 μ g.mL⁻¹)

Transfer accurately 10 mL of the stock solution of each drug into two separate 100-mL volumetric flasks and complete to volume with methanol to get 100 μ g.mL⁻¹ working solution for each drug.

II.4. Procedures

II.4.1. Linearity and construction of calibration curves:

Transfer accurate aliquots equivalent to $(20 - 500) \mu g$ of **HBB** and $(50 - 1200) \mu g$ of **DIP** from their corresponding working solutions (100 $\mu g.mL^{-1}$) or stock solutions (1000 $\mu g.mL^{-1}$) into two separate sets of a series of 10-mL volumetric flasks. Complete the volume with methanol. Make triplicate 20 μ L injections for each concentration. The separation was done on a C₁₈ column using (water: methanol 50: 50 v/v, pH adjusted to 7.0 with triflouroacetic acid and triethylamine) as a mobile phase. All solvents were filtered through a 0.45 μ m membrane filters before use and degassed in an ultrasonic bath for 20 min. Record the chromatograms at ambient temperature maintaining the flow rate at 1.0 mL.min⁻¹ and detect the effluent at 210 nm.

Construct the calibration curves for each compound by plotting the peak area/ 10^4 versus the corresponding concentration and then compute the regression equations.

II.4.2. Analysis of laboratory prepared mixtures:

Prepare mixtures containing HBB and DIP in different ratios. Proceed as mentioned under linearity and construction of calibration curves. Calculate the concentrations of the two compounds from their corresponding regression equations.

II.4.3. Application of the proposed methods to pharmaceutical formulations.

A) For tablet dosage form

The contents of ten tablets of Buscopan compositum[®] were thoroughly powdered and mixed then an amount of the powder equivalent to 250 mg of DIP and 10 mg of HBB was weighed accurately in 250-mL beaker, 70 mL of methanol was added, stirred for about 30 min then filtered through filter paper into a 100-mL volumetric flask, the beaker and the funnel were washed and the volume was completed with methanol to get a concentration of 2.5 and 0.10 mg.mL⁻¹ for DIP and HBB, respectively. Appropriate dilutions were made to bring up a concentration of 50.0 and 2.0 μ g.mL⁻¹ for DIP and HBB, respectively and a concentration of 5.0 and 0.2 μ g.mL⁻¹ DIP and HBB, respectively. The proposed method was applied for the analysis and calculation of HBB and DIP concentrations.

B) For ampoules dosage form

The contents of five ampoules of Buscopan compositum[®] were thoroughly mixed then volume of 1 mL of solution equivalent to 500 mg of DIP and 4 mg of HBB was accurately transferred into a 100-mL volumetric flask and the volume was completed to the mark with methanol to get a concentration of 5.0 and 0.04 mg.mL⁻¹

for DIP and HBB, respectively. Appropriate dilutions were made to bring up a concentration of 250.0 and 2.0 μ g.mL⁻¹ for DIP and HBB, respectively and a concentration of 5.0 and 0.04 μ g.mL⁻¹ DIP and HBB, respectively. The proposed method was applied for the analysis and calculation of HBB and DIP concentrations.

III. Results and discussion

III.1. Method development and optimization

The aim of this work is to develop a method that can be applied successfully for separation and quantification of the studied drugs.

To optimize the RP-HPLC method, it was necessary to test the effect of different variables:

a-The choice of the stationary phase

The reversed-phase separation was preferred to the normal phase due to the drawbacks of the normalphase mode, for example, hydration of the silica with water, which cause peak tailing .C18 column found was to be more efficient than C8 column.

b-The choice of the mobile phase

Mobile phase systems of different compositions and ratios were tried e.g. (water : methanol) using different acids (H_3PO_4 , CH_3COOH and CF_3COOH) at different pH values. Complete separation of HBB and DIP without interference within a suitable time was achieved using water: methanol (50:50, v/v pH adjusted to 7.0 with CF_3COOH acid and triethylamine). This selected mobile phase allows the determination of HBB and DIP in combination without interference and within a suitable time.

c-The effect of pH

Forked peak of HBB at basic medium was a problem. While forked peak of DIP was appeared at acidic medium. Adjustment of pH to inhibit this was tried using different acids and bases like glacial acetic acid, triflouroacetic acid, orthophosphoric acid, sodium hydroxide and triethylamine. Forked peak could be overcome by adjusting the pH at 7.0 ± 0.02 . In addition, the effect of acid type was studied by using acids of different pk_a values (H₃PO₄, CF₃COOH and CH₃COOH). It was found that CF₃COOH acid was necessary for proper peak shape of HBB. Furthermore, the effect of base type was studied by using bases of different pk_b values (NaOH and triethylamine). It was found that triethylamine was necessary for proper peak shape of DIP.

d-Flow rate and the scanning wavelength

Different flow rates were tried of which 1.0 mL.min⁻¹ proved to be of choice providing good separation within 9 minutes.

The two drugs under investigation were dissolved in methanol separately and examined by the spectrophotometer. it was found that detection at 210 nm gave good sensitivity for both compounds.

Finally, a satisfactory separation was obtained by using C18 (25cm×4.6mm i.d. 5µm particle size) column as a stationary phase and using water: methanol (50:50) v/v pH adjusted to 7.0 with CF₃COOH acid and triethylamine) as a mobile phase, maintaining the flow rate at 1.0 mL min⁻¹ with UV detection at 210 nm. The retention times for HBB and DIP were 3.57 and 8.68 min respectively, Figure 2.

III.2. Method validation

Method validation was performed according to ICH guidelines ⁽²⁰⁾.

Linearity of the proposed methods was evaluated and it was evident in the concentration range of 2.0-50.0 μ g.mL⁻¹ for HBB and 5.0-120.0 μ g.mL⁻¹ for DIP, Figures (3-4). Good linearity was evident by the high value of the correlation coefficient and the low intercept value, (Table 4).

The regression equations were calculated and found to be:

YI = 3.522 C1 + 45.92 r1 = 0.9997

 $Y2= 5.326 C2 - 63.28 \qquad r2 = 0.9996$

Where Y1 and Y2 are the peak area /100, C1 and C2 are HBB and DIP concentrations in μ g.mL⁻¹ respectively and r1 and r2 are the correlation coefficients.

Precision of the proposed RP-HPLC method was evident as shown in Table (4).

Accuracy of the proposed method was checked by applying the proposed method for determination of different blind samples of HBB and DIP. The concentrations were calculated from the corresponding regression equations. The results obtained as shown in Table (1-2).

Accuracy of the method was assured by applying the standard addition technique on Buscopan compositium[®] tablets and Buscopan compositium[®] ampoules where good recoveries were obtained as shown in Table (3) revealing no interference from excipients and good accuracy of the proposed method.

Specificity of the proposed method is evident from the RP-HPLC chromatogram as shown in Figure (2). Robustness of the proposed method was evaluated in the development phase by making small changes in the composition of mobile phase and detection wavelength. The low value of %RSD shows that the methods are robust and that deliberate small changes in the studied factors did not lead to a significant change in retention values, area or symmetry of the peaks.

System suitability tests are based on the concept that the equipment, electronics, analytical operations and samples constitute an integral system that can be evaluated as whole. System suitability is used to ensure system performance before or during the analysis of the drugs. System suitability was checked by calculating the capacity factor(K'), tailing factor(T),column efficiency (N), the selectivity factor(γ) and resolution(Rs), where the system was found to be suitable as shown in Table (5).

IV. Conclusion

The proposed method are efficient for providing sensitive, accurate and reproducible quantitative analysis for simultaneous determination of HBB and DIP in bulk powder and in pharmaceutical formulations, without any interference from excipients. RP-HPLC method has the advantages of short analysis time and the availability of the device in every quality control unit so it is suitable for routine analysis.

Statistical analysis was performed by comparing the results of the proposed method with those of manufacturer method. No significant difference was observed regarding accuracy and precision, as shown in Table (6).

The suggested method provides selective, accurate and sensitive analytical procedures for the determination of HBB and DIP. It is suitable for routine analysis and quality control of HBB and DIP in its pharmaceutical formulations.



Molecular formulaC21 H30 Br N O4Molecular formula C13H16N3Na SO4.H2OMolar mass440.4Molar mass351.4Figure (1) chemical structure of HBB (A) and DIP (B)







Figure (3): Linearity of the peak area at 210 nm to the corresponding concentration of HBB $(2-50 \ \mu g \ mL^{-1})$ using HPLC method.



Figure (4): Linearity of the peak area to the corresponding concentration of DIP $(5 - 120 \ \mu g \ mL^{-1})$ using HPLC method.

 Table (1): Results of accuracy for determination of pure authentic of HBB and DIP by the proposed

 RP-HPLC method.

	HBB		DIP			
Taken	Found [*]	Recovery	Taken	Found [*]	Recovery	
(µg.mL⁻	(µg.mL⁻	%	(µg.mL⁻	(µg.mL⁻	%	
1)	1)		1)	1)		
2.00	2.01	100.50	5.00	4.98	99.60	
10.00	10.02	100.20	10.00	10.05	100.50	
20.00	19.96	99.80	50.00	49.97	99.94	
30.00	29.95	99.83	100.00	100.12	100.12	
40.00	40.50	101.25	110.00	110.04	100.04	
50.00	49.98	99.96	120.00	119.10	99.25	
Mean		100.26±			99.91±	
±SD		0.552			0.434	

* Average of three determinations

Mix.	Ratio		HBB	0	DIP			
No.	HBB:DIP	Taken (µg. band ⁻¹	Found* (µg.band ⁻ ¹)	Recovery %	Taken (µg. band ⁻¹)	Found* (µg. band ⁻¹)	Recovery %	
1	1:1	10.00	9.99	99.90	10.00	10.03	100.30	
2	1:2	10.00	10.12	101.20	20.00	19.96	99.80	
3	1:5	10.00	10.05	100.50	50.00	50.05	100.10	
4	1 :10	2.00	2.00	100.00	20.00	20.05	100.25	
5	1: 25**	2.00	2.02	101.00	50.00	49.85	99.70	
6	1:125***	2.00 0.96	1.98	99.00	250.00 120.00	122.00	101.67	
Mean ±SD				100.27 ±0.809	s	5	100.30 ±0.711	

Table (2): Determination of HBB and DIP in laboratory prepared mixtures by the proposed RP-HPLC method.

Average of three determinations

** The ratio present in Buscopan compositium[®] tablets. *** The ratio present in Buscopan compositium [®] ampoules.

Table (3): Application of standard addition technique to analysis of HBB and DIP in dosage forms by the RP-HPI C method

HPLC method.								
Dosage form	Drug	Taken	Found*	Found %	Pure	Pure	Recovery	Mean
_	_	$(\mu g.mL^{-1})$	$(\mu g.mL^{-1})$		added	Found ^{**}	%	±SD
					$(\mu g.mL^{-1})$	$(\mu g.mL^{-1})$		
					2.00	2.04	102.00	$100.97 \pm$
Buscopan	HBB	2.00	1.98	99.00	4.00	4.01	100.25	0.914
compositum					6.00	6.04	100.67	
[®] tablets	DIP	5.00	5.02	100.40	5.00	5.00	100.00	100.50±
Batch No					10.00	10.05	100.50	0.500
115815T					15.00	15.15	101.00	
					2.00	2.05	102.50	100.67±
Buscopan	HBB	2.00	1.99	99.50	4.00	4.04	101.00	2.021
compositum ®					6.00	5.91	98.50	
ampoules	DIP	5.00	5.04	100.80	5.00	5.05	101.00	100.86±
Batch No					10.00	10.09	100.90	0.171
116551A					15.00	15.10	100.67	

* Average of six determinations

** Average of three determinations

Table (4): Results of assay validation parameters of the proposed RP-HPLC method for the determination of HBB and DIP in binary mixture.

	2		
Parameters	HBB	DIP	
	2-50	5-120	
Range ($\mu g.mL^{-1}$)	$(\mu g.mL^{-1})$	$(\mu g.mL^{-1})$	
Slope	3.522	5.326	
Intercept	45.92	63.28	
Correlation coefficient (r)	0.9997	0.9996	
Accuracy	100.26±0.552	99.91±0.434	
$(\text{mean} \pm \text{SD})$			
$(RSD\%)^{a}*$	0.973	0.721	
(RSD%) ^b *	0.753	0.688	

 $(RSD\%)^{a_*}$, $(RSD\%)^{b_*}$ the intra-day and inter-day relative standard deviations of the average of concentrations (20, 40 and 50 µg.mL⁻¹ for each).

method.						
Parameters	For RP-HPLC method					
	Obtained	value	Reference value			
	HBB	DIP				
Resolution (R _s)	11.3	6	>1.5			
Capacity factor(K')	2.59	0.48	1- 10 acceptable			
Tailing factor (T)	1.17	1.00	< 1.5-2			
Selectivity factor(γ)	5.40		> 1			
Number of Theoretical	48219136	12744900	Increases with increases efficiency			
plate(N)						
HETP	0.0518	0.1962	The smaller the value, the higher the			
(cm.plate ⁻¹)			efficiency			
Height equivalent to						
theoretical plate						

 Table (5): Statistical analysis of parameters required for system suitability testing of the proposed RP-HPLC

Table (6): Statistical analysis of the results obtained by proposed RP-HPLC method and reference method for the determination of HBB and DIP.

Parameter	RP-H metl	PLC nod	Reference method		
	HBB	DIP	HBB	DIP	
Mean %	100.26	99.91	99.21	100.22	
SD	0.552	0.434	1.012	0.544	
n	6	6	6	6	
Student 's t-test (2.23) ^b	0.061	0.304			
F-value (5.05) ^b	3.361	1.571			

a manufactured method personal communications.

b the values between parenthesis are the theoretical values for t and F at P=0.05

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