

Cytogenetic effects of two organic insecticides on mitotic Chromosomes of *Allium sativum* root Tip Cells

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Abstract: In this study, two organic insecticides were analyzed, namely, Baicao No.2 which is an insecticide extracted from plants and used for Whitefly, Aphid, and Thrips, and Baicao No.3 used for moth. Three different concentrations of both organic insecticides were examined (0.25ml/l, 0.5ml/l, and 1ml/ liter) for different periods of time (8, 16, and 24 hours) for their cytotoxic and genotoxic effects on cell division (mitotic index) and chromosomes aberration (aberration frequency) on cells of root-tip of *Allium sativum*.

Low concentration (0.25ml/l) of Baicao No.2 caused an increase in the mitotic index after (8 and 16 hours) of treatment and a decrease after longer time of exposure (24 hours); it was significant at $p > 0.05$.

Higher concentrations of Baicao No.2 (0.5ml/l and 1ml/l) decreased the mitotic index; it was significant at $p > 0.05$.

Treatment with (0.25ml/l) of Baicao No.3 decreased the mitotic index after (8 & 16 hours) and increased the mitotic index after (24 hours); treatment with (0.5 ml/l) for (16 and 24 hours) increased the mitotic index; this increase was significant at $p > 0.05$, while treatment with (1ml/l) decreased the mitotic index after (8 hours) of exposure and was significant at $p > 0.05$, and increased the mitotic index after (16 and 24 hours) of exposure; this increase was significant at $p > 0.05$.

The tested concentrations of both insecticides caused an increase in aberration frequency and different types of chromosomal aberration. The chromosomal aberrations were stickiness and disturbance chromosomes mostly during metaphase, anaphase stages, and bridges. Low percentage of star-metaphase and lagging chromosomes in telophase cells were observed after treatment with Baicao No.3, also arrest metaphase were observed in low percentage after treatment with Baicao No.2. The results indicate cytotoxic and genotoxic effects.

I. Introduction:

For several years, increased attention has been focused on integrated pest management (IPM) programs and alternative methods of pest control to reduce pesticide use in agricultural system because of food safety issues, groundwater contamination, and increased environmental awareness to produce safe and economic crop (Kovachet *al.* 1992), and several pressures have accelerated research for more environmentally and toxicologically safe and more selective and efficacious pesticides.

Plants contain a virtually untapped reservoir of pesticides that can be used directly as templates for synthetic pesticides. Numerous factors increased the interest of the pesticide industry and pesticide markets in this source of natural products as pesticides, especially, the diminishing return with traditional pesticide use, the number of options in discovery and development of a natural product as a pesticide is larger than synthetic pesticides. Furthermore, limited molecular complexity, environmental stability, and low activity of many bio-cides from plants, compared to synthetic pesticides, are encouraging.

Tens of thousands of secondary products of plants have been identified and there are estimates that hundreds of thousands of these compounds exist. There is growing evidence that most of these compounds are involved in the interaction of plants with other species—primarily the defense of the plant from plant pests (Duke, 1990). Even though, several researchers found that the aqueous extracts of some plants inhibited seed germination and seedling growth of wheat and peas. Mitodepressive and radiomimetic effect (Pandya, 1975, Shehab, 1979, Shehab *al.*, 1978, Mondal, 2006), (Alam *etal.*, 1987) observed that the leaf extracts effected the mitotic frequency and caused some abnormal chromosomes.

II. Materials and Methods

1-Tested Materials

Baicao No.2 is an insecticide extracted from plants; the effective material is 1% matrine and used for Whitefly, Aphid and Thrips. Baicao No.3 is a new type of insecticide consists of 0.45% matrine plus microorganism slight emulsion used for moth. Three different concentrations of both natural pesticide were examined (0.25ml/l, 0.5ml/l and 1ml/ liter) for different periods of time (8, 16, 24 hours) for their cytogenetic effect on cell division (mitotic index) and aberration frequency (chromosomes aberration) on cells of root tip of *Allium sativum*.

2-Sample Preparations

Plant test system is widely used for monitoring genotoxicity of chemicals because of many advantages such as low cost, easily availability throughout the year, ease of handling, good chromosome condition for the study of chromosome damage and good correlation with other test systems (Sobita and Bhagirath, 2005). In this study, *Allium sativum* (garlic) $2N = 16$ obtained from local markets were used as testing material for this study. The loose outer scales and old roots of the garlic were scraped and suspended in small beaker with distilled water.

III. Treatments

3.1- *Allium sativum* were suspended in a small beaker (50ml) with distilled water to enhance the root tips to grow until they reach 0.5–1 cm in length; then transferred to another beaker containing freshly prepared solution of insecticides and left for different periods of time. Three different concentrations for three different periods of time were used, and one bulb of garlic was used for each treatment.

3.2- Negative control for each time was used; the root tips of garlic were treated with distilled water only, and used as a comparative sample for the effects of tested pesticides.

IV. slides preparation

The roots were treated with different concentrations (0.25 ml/l, 0.5 ml/l, and 1 ml/liter) for periods of time (8, 16, 24 hours), then the roots were detached, fixed in freshly prepared 3:1 (v/v) ethanol alcohol: glacial acetic acid for 24 hours; root tips of *Allium sativum* were hydrolyzed in 1N HCL at 60 degrees centigrade for 8 minutes; roots tips were then washed with distilled water several times and stained with 1% acetocarmine, five temporary slides were prepared using the squash technique, two root tips on each slide were examined for the effects of Baicao No.2 and Baicao No.3 on mitotic index (MI). The same slides were analysed for the types and frequencies of chromosomal abnormalities produced by the examined pesticides.

V. Scoring of slides and data analysis

5.1- The slides were viewed under light microscope (Phenix P H 50 DB047VU) using the 40X objective lens immersion. The most representative ones for each structural aberration were photographed using (Phenix micro Image analyzer Software 2008 En V2, 2)

5.2-Mitotic index: On one slide for each treatment, a total of 2000 cells, were scored. The mitotic index (MI) was expressed as the number of dividing cells per total cells scored.

5.3-Cytotoxicity: The mitotic index of the treated cells at each dose of each insecticide were compared with that of the negative control group

5.4-Genotoxicity test: Chromosomal aberration per dose of each insecticide were examined; the percentage of cells with aberrations of each dose for each insecticide were compared with that of the negative control using the (SPSS 16.0 for Windows statistical package). Two-way Analysis of Variance was the statistical method used for determining the significance of difference at $P = 0.05$.

VI. Result and Discussion:

Mitotic index (MI)

Three different concentrations of two organic insecticide (0.25, 0.5 and 1 ml/liter) were tested for three periods of time (8, 16, and 24 hours) for their cytotoxic and on mitotic index and chromosomal aberration comparing with negative control.

Table-1 and figures (1&2) show the effect of Baicao No.2 and Baicao No.3 on the mitotic index (percentage of divided cells)

Baicao No.2: the treatment with 0.25 ml/l of Baicao No.2 caused increasing the mitotic index after 8 and 16 hours of exposure and was significant at $p > 0.05$, and decreased after 24 hours. Both treatments with 0.5 ml/l and the highest concentration 1 ml/l of Baicao No.2 caused decreasing the mitotic index after treatment with the three different time periods, especially after 16 hours of exposure (6.85, 5.97) respectively, compared with negative control (11.45), and was significant at $p > 0.05$.

Baicao No.3 The mitotic index decreased after treatment with 0.25 ml/l for 8 and 16 hours, it was significant at $p > 0.05$; while treatment for 24 hours caused increasing in mitotic index; this increase was significant at $p > 0.05$; treatment with 0.5 ml/l decreased the mitotic index after 8 hours of exposure it was significant at $p > 0.05$, and increased the mitotic index after 16 and 24 hours of exposure, it was significant at $p > 0.05$.

>0.05; also the highest concentration 1ml/l caused similar result, treatment for 8 hours and 24 hours caused a decrease in the mitotic index and an increase in the mitotic index after 16 hours; it was significant at $p > 0.05$.

The increase of mitotic index can be due to the allelochemical which can act by a innumerable manner to affect the plant germination and growth; the principal mechanisms involve alteration of the mitotic index and suppression of hormones (Rice, 1984) (Sousa *et al*, 2009); (Sopova *et al*, 1983) found that low concentration of tobacco leaf extract exerted a stimulating effect, whereas high concentration acted as a mitodepressant on root-tip cells of *Allium cepa* L. Also (Krivokaoic *et al*, 1970) reported that the weak concentration of immature of *Solanum nigrum* L. fruits extract stimulate cell division, and that may be due to the presence of a cytokinin-like substance in the extract, whereas stronger concentration reduced the intensity of mitosis. (Koulet *et al*, 2005) reported that beside active components additional components present and this may affect the effect of active components, these known and unknown components might be acting as synergistic for the therapeutic and antagonists for the side effects of the active components as well as the other toxic components in the extract.

Decreasing of mitotic index can be explained by the arrest of the division of the interphasic nucleus, as well as by death of interphasic nucleus, hindering the onset of prophase and thus the division of the cells (Sousa *et al*, 2009). (Yadav, 1986) reported that disrupt the mitotic cycle occur in three ways (1) by inhibiting the process of cell division, (2) by disturbing the normal functioning of the mitotic spindle, and (3) by producing chromosomal abnormalities. Also reduction in the percentage of the cell division indicates that the natural insecticide has antimitotic activity, by affecting some vital function units of cell and the normal functioning of the spindle mechanism. (Monda *et al*, 2006) observed the same effect on mitotic index after testing the effect of water extract of *Ipomoea* species on *Allium cepa* root tip.

Type of Chromosomal aberrations

Table-2 and figures (3&4) show the type of chromosomes aberrations after treatment with Baicao No.2 and Baicao No.3 insecticides.

Root tips of *Allium sativum* fixed without treatment served as control and showed normal cells, though some chromosomal aberrations were encountered; These might have arisen due to the automutagenic substance (Dubinin and Scerbako, 1962), similarly (Kaushik, 1996) reported the occurrence of chromosomal aberrations in water treatment only.

Baicao No.2 showed that all of the three tested concentrations cause chromosomal abnormalities. Low concentration 0.25ml/l cause chromosome aberration namely stickiness and disturbance at metaphase cells and chromosomes disturbance and bridges in anaphase cells and it was significant at $p > 0.05$. Treatment with 0.5ml/l for 8 and 24 hours cause chromosome stickiness and bridges, also cause disturbance after root treated for 8, 16, and 24 hours with insecticide. The highest tested concentration 1ml/l cause stickiness after 8 hours of treatment and arrested chromosomes (C-metaphase) after 16 hours and chromosomes disturbance after 8, 16, and 24 hours in metaphase cells, cells with anaphase stage shows chromosomes disturbance and bridges after 8 hours and lagging chromosome after treatment for 16 hours.

Baicao No.3 Treatment with 0.25ml/l for 8 hours causes chromosomal stickiness in metaphase cells. Stickiness, disturbance and bridge in the chromosome of anaphase cells, also the tested concentration cause disturbance, bridge and star anaphase after treatment for 16 and 24 hours, star anaphase in low percentage, in telophase cells the treatment induce lagging chromosome after treatment for three time periods and it was significant at $p > 0.05$. 0.5ml/l cause chromosome stickiness and star –metaphase. In anaphase cells treatment with 0.5ml/l induce star-anaphase, bridges and chromosome disturbance; the effect of this concentration was significant at $p > 0.05$.

The highest tested concentration 1ml/l cause chromosomal disturbance and chromosomal stickiness, also disturbance of anaphase chromosomes, lagging chromosome, bridge, and lagging chromosome in telophase cells. Similar result was observed by (Sudharsan and Reddy, 1971) and (Kabarity and Malallah, 1980)

the stickiness of chromosome that arise after treatment can be explained as a result of the depolymerization of DNA (Kaushik, 1993), (Yadav and Rathora, 1984). (Babichet *et al*, 1997) reported that metaphases with sticky chromosomes lose their normal appearance and appear to have sticky “surface” which causes chromosome agglomeration, possibly due to the effect on chromatin and chromosome organization, disturbance mostly during metaphase and anaphase stages due to the failure of spindle mechanism by the treatment (Yadav, 1986), lagging chromosome arise from centromere adhesion causing abnormality of chromosomes movement towards the equatorial (Bader *et al*, 1992). Anaphase bridges usually arise because of chromosomes stickiness and end up inhibiting cell division

(Nwakanma and Okoli, 2010).

(Tilay, 2012), (Ishida *et al*, 1988) reported that an increase in chromosomal aberration may result from interactions of a great variety of chemical agents with DNA.

VII. Conclusion

The results indicate that Both natural insecticides (extracted from plants) have cytotoxic effect for their effects on mitotic index resulting in decreasing in cell division especially treatment with Baicao No. 2, and found that time of the treatment play an important role for the effects observed; also Baicao No.2 and Baicao No.3 have clastogenic effect because of the chromosomal aberration that absorbed during (M) phases, that cause the decreasing in cell division.

Table (1): Total number of examined cells, mitotic index and aberration frequency after treatment with different concentrations of insecticides for different periods of time on cells of root tip of *Allium sativum*

pesticide	Concentration/ MI/L	Treatment/ hour	No. total cells	divided cell	total number of abnormal cell	mitotic index	Aberration frequency
control	0	8	2052	283	4	13.79	0.014
	0	16	2436	279	4	11.45	0.014
	0	24	2069	238	4	11.5	0.0168
Baicao no 2	0.25	8	2523	373	22	14.78	0.085
	0.25	16	2271	299	8	13.166	0.0267
	0.25	24	2368	225	8	9.5	0.035
	0.5	8	2271	274	7	12.065	0.0255
	0.5	16	2158	148	7	6.85	0.047
	0.5	24	2053	236	6	11.49	0.025
	1	8	2157	198	7	9.179	0.035
	1	16	2579	154	5	5.97	0.0029
	1	24	2000	210	4	10.5	0.019
Baicao no 3	0.25	8	2659	351	19	13.2	0.054
	0.25	16	2122	203	5	9.56	0.0246
	0.25	24	2120	268	6	12.64	0.022
	0.5	8	2425	261	7	10.76	0.0269
	0.5	16	2592	339	13	13.07	0.038
	0.5	24	2147	260	6	12.11	0.023
	1	8	2269	296	6	13.045	0.02
	1	16	2133	276	11	12.9	0.039
	1	24	2187	249	10	11.39	0.04

Table (2): Type of chromosomal aberrations on root tip cells of *Allium sativum* after treatment with different concentrations of the tested insecticides for different periods of time.

pesticide	control	duration of time (h)	metaphase				anaphase				telophase	AF%	
			sticky	disturb	c-metaphase	s-metaphase	sticky	disturb	lagging	bridge	s-anaphase		lagging
control	Distill water	8		0.0035			0.0035			0.007			1.4
		16	0.011				0.0035						1.4
		24		0.0084						0.0042			1.68
Baicao 2	0.25	8	0.011	0.02			0.0027						5.9
		16	0.007	0.007									2.7
		24	0.018	0.018						0.01			3.8
	0.5	8	0.0036	0.0036			0.0073			0.011			2.6
		16		0.014	0.014						0.0042		4.7
		24	0.013	0.008									2.5
	1	8	0.015	0.01			0.005			0.005			3.5
		16		0.013	0.013					0.013			3.2
		24		0.019									1.2
Baicao 3	0.25	8	0.0085				0.0028	0.0028		0.017			5.4
		16						0.0049		0.0099	0.0049	0.003	2.48
		24						0.0037		0.011			2.2
	0.5	8	0.015			0.004					0.004	0.004	2.7
		16	0.0177	0.0088						0.012			3.8
		24		0.0038						0.012		0.0038	2.3
	1	8		0.02									2
		16	0.015	0.007					0.004	0.015			4
		24	0.004					0.008	0.004	0.02		0.004	4

Figures :1- Star metaphase .2- Star anaphase. 3- disturb metaphase .4- C- metaphase. 5- disturbed anaphase. 6&7- Lagging chromosome at anaphase .8&9- Bridge at anaphase..

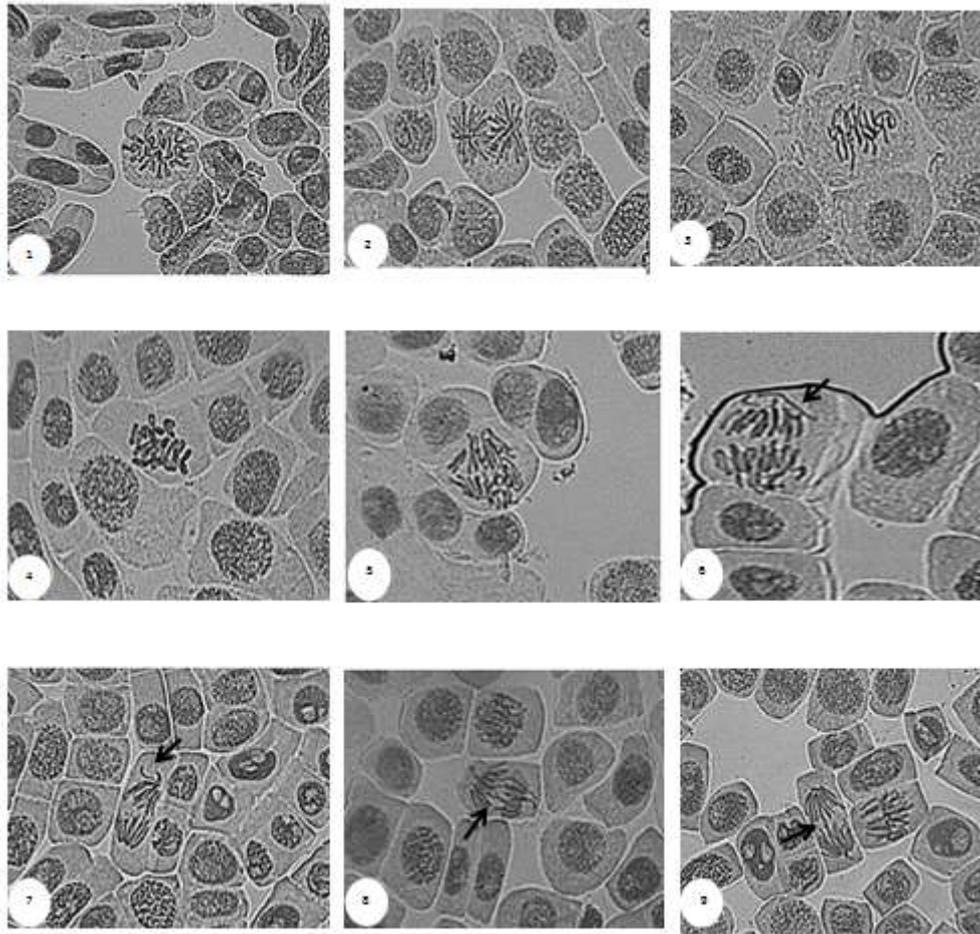


Figure 1: Effect of different concentrations of Paicao no 2 on mitotic index on root tip cells of *Allium sativum*

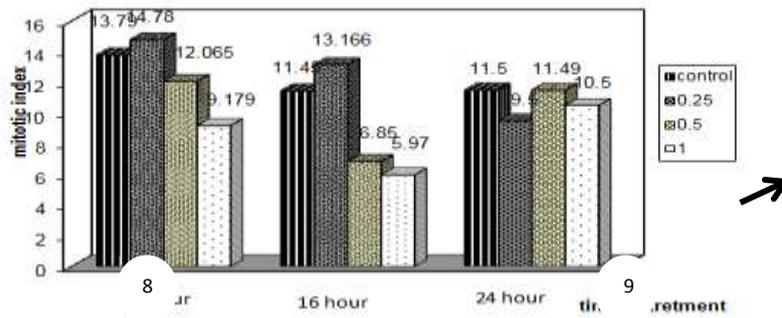
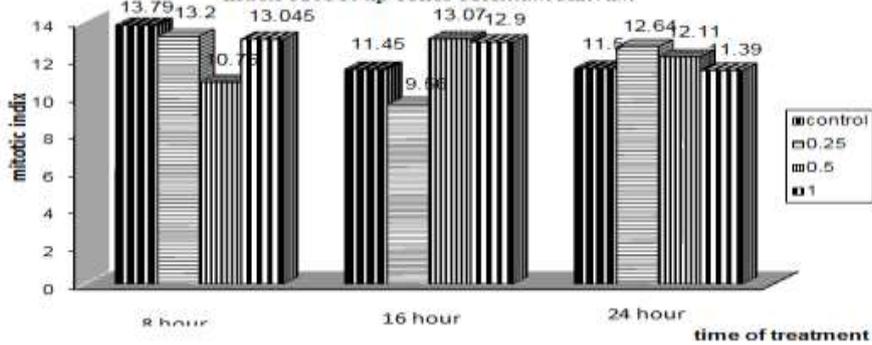
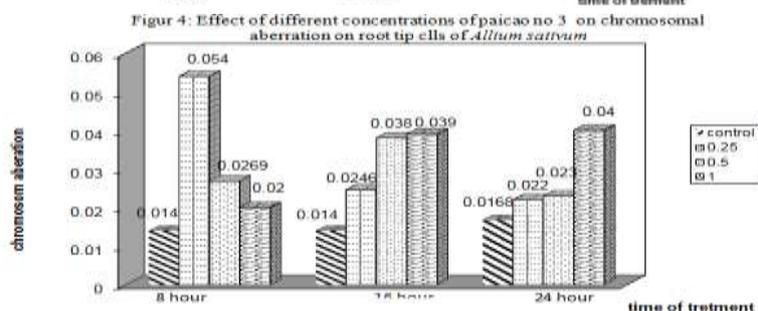
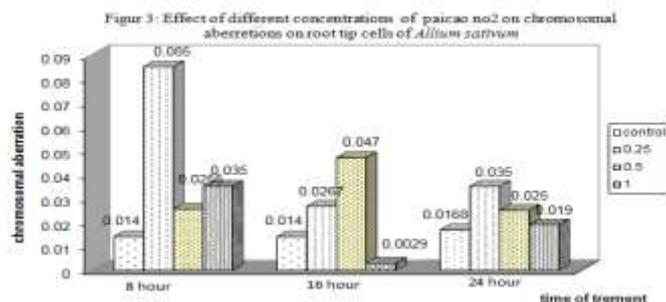


Figure 2: Effect of different concentrations of baicao no 3 on mitotic index of root tip cells of *Allium sativum*





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